

INVITED REVIEW



Synthetic microneurotrophins: Neurotrophin receptors for therapeutics of neurodegenerative diseases

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Neurodegenerative disorders are characterised by the chronic progressive degeneration of specific neuronal subtypes, neuroinflammation, myelin damage and synaptic loss. Despite their growing incidence, advancements in effective treatments remain limited, because of lack of knowledge for the aetiology of the diverse pathophysiology to design systematic therapies. Several studies highlight the role of neurotrophic factors (NTFs) as potential neuroprotective, regenerative therapies for these disorders. Although NTFs hold protective and regenerative potential for chronic neuroinflammatory and neurodegenerative conditions, major hurdles impair their clinical use, such as optimising the dosage of NTFs, minimising the invasiveness of delivery methods, overcoming blood-brain-barrier (BBB) impermeability and managing side effects. In the last two decades our group have synthesised and screened a large chemical library of steroidal analogues of **dehydroepiandrosterone** (DHEA), an endogenous steroid hormone, for their ability to mimic neurotrophin neuroprotective and neurogenic actions. Interestingly, DHEA was shown to interact with all neurotrophin receptors, acting most probably as an ancestral neurotrophin early in evolution. However, its chronic pharmacological use is questioned by its action as a major precursor of steroidogenesis. This review highlights the findings of numerous preclinical studies on these synthetic, non-toxic, BBB permeable DHEA derivatives, named microneurotrophins (MNTs), deprived of endocrine actions, activators of specific neurotrophin receptors. The multimodal actions of MNTs against neuronal death and activation of microglia, in addition to their beneficial effects in synaptogenesis and neurogenesis, place them as interesting lead molecules in the armamentarium of therapeutics for neurodegeneration.

KEYWORDS

Alzheimer's disease, amyotrophic lateral sclerosis, BDNF, diabetic retinopathy, microneurotrophins, multiple sclerosis, NGF, p75 neurotrophin receptor, Parkinson's disease, Trk receptors

Abbreviations: AD, Alzheimer's disease; ALS, Amyotrophic lateral sclerosis; APP, Amyloid precursor protein; DHEA, Dehydroepiandrosterone; DR, Diabetic retinopathy; EAE, Experimental Autoimmune Encephalomyelitis; GDNF, Glial cell-line derived neurotrophic factor; hNPC, human neural progenitor cell; MS, Multiple sclerosis; NGF, Nerve growth factor; NSC, neural stem cell; NT3, Neurotrophin 3; PD, Parkinson's disease; RGC, retinal ganglion cell; Trk, tropomyosin receptor kinase.

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1 | INTRODUCTION

Neurodegenerative diseases are characterised by the progressive neuronal loss, resulting in significant impairment of nervous system function and represent a major global health challenge. Neurodegeneration and neuroinflammation are fundamental hallmarks of central nervous system (CNS) disorders (Ransohoff, 2016). Neurodegenerative diseases, including multiple sclerosis (MS), Alzheimer's disease (AD), Parkinson's disease (PD), amyotrophic lateral sclerosis (ALS), and, in acute cases, spinal cord injury (SCI), remain incurable and impose substantial costs on both individuals and society (Lindvall & Kokaia, 2010; M. Metcalfe et al., 2017; Przedborski et al., 2003). The most common are dementias, with AD and PD, with their incidence rising significantly as the world's population ages. Currently, an estimated 50 million people worldwide are affected by neurodegenerative diseases, and this number will rise to 115 million by 2050 ('The Challenge of Neurodegenerative Diseases in an Aging Population,' 2017).

At present, available therapeutic options are insufficient to reverse or even halt the progression of neurodegenerative processes (Frozza et al., 2018; Volkman & Offen, 2017). Additionally, the development of effective therapies is hindered by the complexity of the pathogenic mechanisms underlying neuronal loss and the conflicting pathological causes of these diseases. Furthermore, the aetiology of sporadic neurodegenerative diseases is unknown, rendering the design of effective systematic therapies for each one a daunting task. The challenge is further exacerbated by the significant limitations faced by most therapeutic agents, particularly their inability to effectively cross the blood–brain barrier (BBB) (Ebrahimi et al., 2020; Zlokovic, 2008).

Neurotrophins are polypeptide growth factors that regulate key neuronal functions during development and adulthood, including cell differentiation, proliferation, survival, axonal growth and synaptic plasticity. The neurotrophin family includes **nerve growth factor** (NGF), **brain derived neurotrophic factor** (BDNF), and **neurotrophins 3 and 4/5** (NT-3, and NT-4/5). They exert their effects via binding to their specific high-affinity membrane receptors: **TrkA** (for NGF), **TrkB** (for BDNF and NT4/5) and **TrkC** (for NT3). Additionally, all neurotrophins can bind with lower affinity to the pan-neurotrophin receptor, **p75NTR** (Barker, 2004; Bibel et al., 1999), while their immature, pro-apoptotic isoforms, pro-neurotrophins, present high affinity for the p75^{NTR}-Sortilin complex (Jansen et al., 2007). Released by target tissues, they prevent presynaptic neurons from undergoing apoptosis and promote the differentiation of progenitor cells into neurons (Allen & Dawbarn, 2006; Huang & Reichardt, 2001). Their complex signalling pathways play crucial roles in cell physiology and are essential for understanding various diseases (Chao et al., 2006; Numakawa & Odaka, 2022; Reichardt, 2006). Numerous studies have demonstrated that the processing and expression levels of neurotrophins are dysregulated in MS, AD, PD and ALS, contributing significantly to degenerative pathology (Budni et al., 2015; Jiao et al., 2016; Just-Borràs et al., 2022; Karimi et al., 2022; Lorigados Pedre et al., 2002). Hence, the specific role of neurotrophins in each of these

disorders and their therapeutic potential has been extensively examined, showing promising results. However, natural neurotrophins have short half-lives, difficulty in crossing the BBB and poor pharmacokinetic profiles. Thus, research interest turned to the synthesis of small molecules that mimic neurotrophins action and selectively activate Trk receptors in order to be used as therapeutic agents in neurodegenerative disorders (Josephy-Hernandez et al., 2017). In this context, our group have synthesised and identified novel small steroidal derivatives of DHEA, an endogenous hormone precursor, that interact with the TrkA, TrkB and p75 neurotrophin receptors, exhibiting similarities to certain actions of NGF and/or BDNF, such as the protection of specific neuronal populations from degeneration (Calogeropoulou et al., 2009; Padiaditakis, Efstathopoulos, et al., 2016). These highly lipophilic molecules demonstrate significant potential as therapeutic agents against neurodegenerative diseases, as they can selectively mimic specific beneficial neurotrophic actions and pass the BBB while circumventing undesirable side effects.

Here, we provide a comprehensive overview of the role of neurotrophins and their prospective therapeutic uses in treating neurodegenerative diseases such as MS, AD, PD and ALS. We summarise the different approaches of NTFs administration to animal models or to patients suffering from neurodegenerative diseases, demonstrating the limitations holding NTFs back from successful clinical use. Building on these insights, we also review key studies that highlight the innovative potential of our small-molecule synthetic microneurotrophins. These compounds are uniquely capable of crossing the blood–brain barrier (BBB), selectively activating neurotrophin receptors, and mimicking the activity of endogenous neurotrophins—without eliciting the adverse effects typically associated with natural neurotrophins or endogenous steroids. These studies highlight the promising potential of synthetic microneurotrophins as groundbreaking therapeutic agents for various neurodegenerative disorders.

2 | NEUROTROPHINS AND THEIR RECEPTORS IN NEURODEGENERATIVE DISEASES: INVOLVEMENT IN NEURONAL DEATH, NEUROINFLAMMATION, DEMYELINATION AND NEUROGENESIS

2.1 | Alzheimer's disease (AD)

Alzheimer's Disease (AD) is a neurodegenerative disorder, the most common type of all dementias, marked by severe memory loss, cognitive decline and impaired ability to perform daily tasks (Prince et al., 2015). The main hallmarks of the disease include extracellular amyloid plaque deposition and intracellular hyperphosphorylated neurofibrillary tau tangles. Neurotrophins play a significant role in AD progression, with NGF emerging as a critical regulator of cholinergic neurons (Figure 1), whose degeneration characterises late-stage AD (J. Allen et al., 2011; Durany et al., 2000; Ginsberg et al., 2006; Hock et al., 2000; Michalski & Fahnstock, 2003; Peng et al., 2004). Reduced levels of BDNF in the AD brain imply impaired neurotrophic

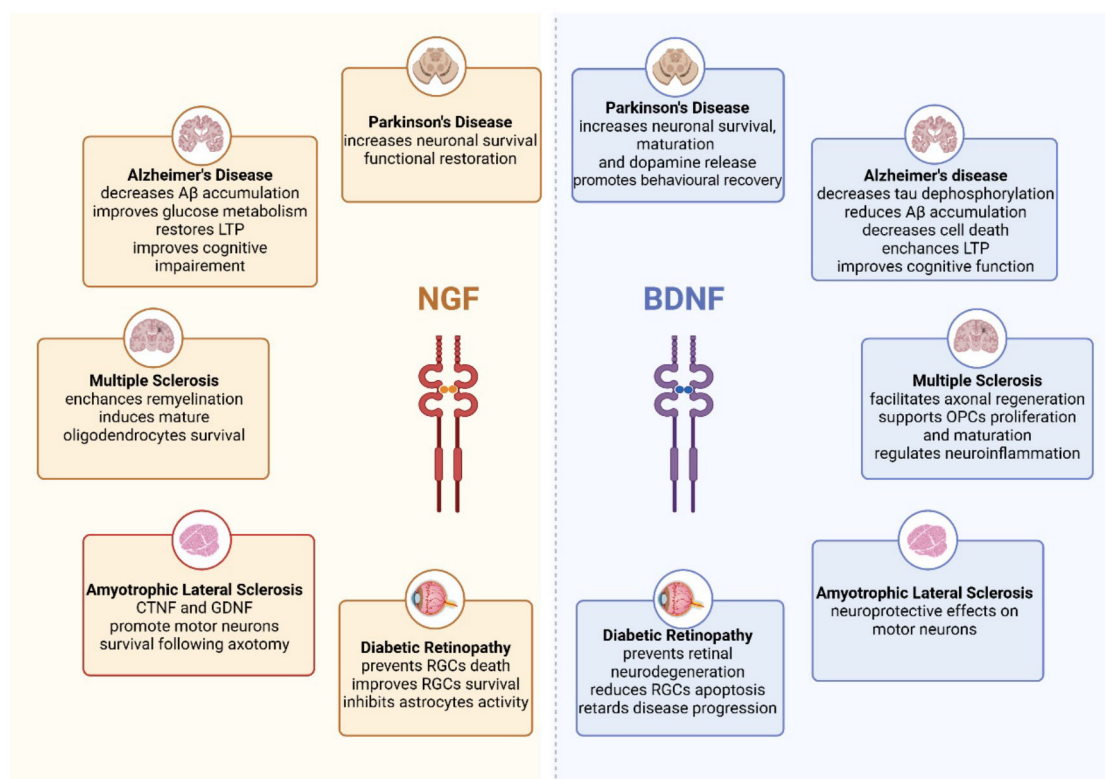


FIGURE 1 Schematic representation of the roles of neurotrophins NGF and BDNF in the treatment of neurodegenerative diseases. Created in BioRender.

support, possibly exacerbating neurodegeneration, while evidence indicates that enhancing BDNF signalling could improve cognition (Arancibia et al., 2008; Ibrahim et al., 2022; Jiao et al., 2016; Nagahara & Tuszynski, 2011). Elevated serum BDNF levels have also been linked to cognitive restoration (Weinstein et al., 2014). Moreover, signalling through p75NTR plays a key role in neurodegeneration associated with various diseases. Hence, the correlation between p75NTR and AD has been a subject of particular scrutiny in research.

Numerous studies highlight the therapeutic potential of NGF in AD (Aloe et al., 2015; Chan et al., 2004; Mitra et al., 2019; J. Wang et al., 2020). Reduced NGF levels in the nervous system and cerebrospinal fluid (CSF) of AD patients correlate with cholinergic degeneration, a hallmark of the disease (Budni et al., 2015; Francis et al., 1999). Mouse models lacking NGF or its high affinity TrkA receptor exhibit decreased choline O-acetyltransferase (ChAT) expression and cholinesterase activity in the basal forebrain and hippocampus (Crowley et al., 1994). Chronic NGF infusion restores long-term potentiation (LTP) in aged, cognitively impaired rats, further supporting its critical role in the CNS (Lübke et al., 2021; Villoslada et al., 2000). Additionally, reduced cortical TrkA levels are associated with lower cognitive performance in AD patients (Counts et al., 2004). This evidence suggests that declines in NGF and TrkA in the cortex and nucleus basalis of Meynert (nbM) could serve as early markers of AD onset (Mufson et al., 2012).

De-regulation of NGF is implicated in AD (Biernacki et al., 2005; Cattaneo & Calissano, 2012). In AD, deficits in NGF and degeneration

of the basal forebrain cholinergic system are closely associated with cognitive decline and dementia (Iulita & Cuello, 2014). NGF facilitates the non-amyloidogenic cleavage pathway of amyloid precursor protein (APP), thereby reducing amyloid β -peptide (A β) generation in mouse brains (Yang et al., 2014). Interestingly, higher NGF levels have been detected in the CSF and dentate gyrus of AD patients compared to controls (Budni et al., 2015; Faria et al., 2014). Clinical trials reveal that NGF treatment resulted in lower A β 1–42 levels in the CSF (Andreasen et al., 1999; Ferreira et al., 2015). NGF gene transfer therapy in early-stage AD patients showed axonal extensions towards the NGF source and activation of CREB and c-Fos markers (Tuszynski et al., 2015). Additionally, NGF administration improved brain activity, glucose metabolism, cognition and clinical status, while reducing brain atrophy and elevating CSF A β 1–42 levels (Ferreira et al., 2015).

ProNGF, the precursor of NGF, plays a critical role in AD, with alterations in its levels linked to cognitive dysfunction in key brain regions, including the frontal cortex, posterior cingulate, superior temporal cortex and hippocampus (Mufson et al., 2012; Perez et al., 2014; Sperling et al., 2014). Accumulation of proNGF in the preclinical stages of AD correlates with cognitive decline (Peng et al., 2004) and contributes to neuronal degeneration through its preferential binding to cell death p75NTR receptor, bypassing the pro-survival effect of TrkA interaction (Al-Shawi et al., 2008; Ioannou & Fahnestock, 2017). Pan-neurotrophin p75NTR receptor is highly expressed in adult basal forebrain cholinergic neurons, which are among the earliest to degenerate in AD. The balance of pro-survival versus pro-apoptotic

signalling in these neurons may depend on TrkA and p75NTR stoichiometry, co-receptor availability and the physiological role of proNGF in different contexts (Bruno et al., 2004; F. Longo et al., 2008; F. M. Longo et al., 2007).

Studies highlight the critical role of p75NTR in AD pathogenesis, including its involvement in A β deposition (Y. J. Wang et al., 2011; Yao et al., 2015) and tau hyperphosphorylation (Shen et al., 2019), implicating the receptor in disease progression (Zeng et al., 2011). In vitro research demonstrates that A β -amyloid, a key component in AD brains, binds p75NTR, inducing pro-apoptotic effects (Knowles et al., 2009; Saadipour et al., 2013; Yaar et al., 1997). Notably, Yao et al. (2015) observed a significant reduction in p75ECD levels in the cerebrospinal fluid (CSF) and brains of AD patients and APP/PS1 transgenic mice. Another study reported elevated serum but reduced CSF p75ECD levels in AD patients compared to a control group, suggesting p75ECD as a potential AD biomarker. Notably, a small molecule that modulates p75NTR activity has successfully completed Phase 2a in patients with AD (Shanks et al., 2024).

BDNF is in the centre of research interest in AD because of the widespread expression of its receptor, TrkB, in the CNS and its critical role in neurogenesis, synaptic plasticity and repair (De la Rosa et al., 2019; Kramár et al., 2012; Miranda et al., 2019; Pencea et al., 2001). BDNF-TrkB signalling also regulates adult hippocampal neurogenesis, a process severely affected in AD (Colucci-D'Amato et al., 2020; Moreno-Jiménez et al., 2019; Salta et al., 2023; Vilar & Mira, 2016). Elevated serum BDNF levels in early-stage AD patients correlate with reduced cognitive decline (Buchman et al., 2016; Laske et al., 2011). Additionally, BDNF has demonstrated protective effects against amyloid-beta-induced neurodegeneration and demyelination in in vitro and in vivo models (Arancibia et al., 2008; Mitroshina et al., 2020; Nagahara et al., 2009; Zota et al., 2024). Its stimulation induces the de-phosphorylation of tau protein and shifts APP processing towards a non-amyloidogenic pathway (Jiao et al., 2016; Nigam et al., 2017). These findings highlight BDNF signalling as a potential therapeutic target for AD (Azman & Zakaria, 2022; Gao et al., 2022).

Reduced BDNF levels in the AD brain indicate insufficient neurotrophic support, potentially exacerbating neurodegeneration. Evidence suggests that enhancing BDNF signalling may improve cognition in AD (Arancibia et al., 2008; Jiao et al., 2016). In a mouse model with AD-like pathology, cognitive improvement from stem cell transplantation was lost when BDNF was depleted, but ectopic BDNF expression restored cognitive function (Blurton-Jones et al., 2009). Additionally, the Val66Met polymorphism in BDNF, common in Caucasian and higher in Asian populations, impairs BDNF sorting, transport and secretion, leading to hippocampal dysfunction (Kuczewski et al., 2010). Both sporadic and familial AD cases with this polymorphism exhibit faster cognitive decline, hippocampal atrophy and altered CSF tau levels, highlighting the potential of boosting BDNF signalling to enhance synaptic plasticity and cognition in AD (Boots et al., 2017; Lim et al., 2018).

Studies suggest that blocking glutamatergic neurons with scopolamine or stimulating the GABAergic system reduces BDNF mRNA levels in the hippocampus (Connor et al., 1997; Da Penha Berzaghi

et al., 1993). The cholinergic system, known to regulate BDNF mRNA levels, also plays a role in this process (Phillips et al., 1991). Given that both the glutamatergic and cholinergic systems degenerate in AD, this highlights BDNF's potential contribution to cognitive impairment in AD (Connor et al., 1997; Coyle et al., 1983; Lübke et al., 2021). Reduced BDNF mRNA levels in the hippocampus of AD patients may contribute to basal forebrain cholinergic neuron atrophy (Lübke et al., 2021; Phillips et al., 1991). Moreover, BDNF administration has shown promising effects, such as decreased A β production, repair of A β -induced damage, reduced cell death, improved cognitive function, synaptic loss mitigation, and slowed cognitive decline (Rohe et al., 2009; Li et al., 2012). The enhancement of memory and LTP through increased TrkB signalling and BDNF expression further suggests the therapeutic potential of BDNF in AD (Ji et al., 2010; Monteggia et al., 2004; Wan et al., 2014).

Postmortem studies consistently show reduced levels of BDNF in the brains of AD patients (Meng et al., 2013; Michalski & Fahnstock, 2003). Elevating BDNF levels through gene transfer in the entorhinal cortex enhances memory and spatial learning in various models, including APP transgenic mice, aged rats and APP/PS1/tau transgenic mice following neuronal stem cell transplantation in the hippocampus (Blurton-Jones et al., 2009; Lattanzio et al., 2014; Nagahara et al., 2009). Studies in APP/PS1 mice show increased neurogenesis and BDNF mRNA levels in the hippocampus (Hsiao et al., 2014). In AD hippocampus, postmortem analysis reveals lower levels of BDNF mRNA. A β protein, a hallmark of AD pathology, inhibits pro-BDNF conversion to BDNF and affects BDNF levels through tau hyperphosphorylation via calcineurin activation (Ramser et al., 2013). A β also impairs retrograde transport of the BDNF-TrkB complex via the enzyme ubiquitin C-terminal hydrolase L1 (Poon et al., 2013). In vitro, oligomeric A β significantly reduces BDNF expression (Garzon and Fahnstock, 2007; DaRocha-Souto et al., 2012; Rosa & Fahnstock, 2015). A β interaction with PKA activation also decreases CREB phosphorylation and BDNF expression (Rosa & Fahnstock, 2015; Colucci-D'Amato et al., 2020). BDNF may also influence APP processing via the non-amyloidogenic α -secretase pathway in neuronal cell lines (Holback et al., 2005). Although the exact effects of BDNF on A β production remain unclear, co-treatment with BDNF in the hippocampus or entorhinal cortex prevents A β 1-42-induced impairment of LTP, highlighting its neuroprotective potential (Arancibia et al., 2008; Criscuolo et al., 2015; Kitiyanant et al., 2012; Lübke et al., 2021). However, delivery of BDNF through intracerebroventricular injections or other non-invasive methods has been unsuccessful because of side effects or low effectiveness (Givalois et al., 2004; Kopec et al., 2020).

Neurotrophin-3 (NT-3) levels remain stable in both AD brain and cerebrospinal fluid (Durany et al., 2000; Hock et al., 1998, 2000a, 2000b; Murase et al., 1994; Phillips et al., 1991). NT-3 increases APP promoter activity by 3.82-fold in PC12 cells, compared to NGF, which causes a 5.81-fold increase (Ge & Lahiri, 2002) and protects primary cortical neurons from amyloid-beta toxicity by inhibiting caspase cleavage and enhancing Akt phosphorylation through PI-3K signalling (Lesné et al., 2005). Additionally, NT-3 promotes the expression of

neuronal apoptosis inhibitory protein-1, preventing amyloid-beta-induced apoptosis. In lesion models mimicking AD, NT-3 prevents degeneration of noradrenergic neurons in the locus coeruleus (Arenas & Persson, 1994). Moreover, NT-3 overexpression facilitates the differentiation of bone marrow-derived mesenchymal stem cells (BMSCs) into neurons, improving cognitive function in AD model rats, whereas NT-3 silencing inhibits differentiation and exacerbates cognitive deficits (Yan et al., 2021). These findings suggest that NT-3 may have a therapeutic potential in neurogenesis and cognitive improvement in AD. However, its role appears less pivotal and versatile than NGF and BDNF in AD pathophysiology and treatment.

2.2 | Parkinson's disease (PD)

PD is an age-related, highly heterogeneous neurodegenerative condition characterised by progressive loss of nigrostriatal dopaminergic (DA) neurons in the midbrain that leads to severe movement deficits and degeneration of peripheral DA neurons that mediates non-motor symptoms. The main pathological feature of PD is the presence of eosinophilic cytoplasmic inclusions in neurons, known as Lewy bodies, which primarily consist of aggregates of misfolded **alpha-synuclein** (a-syn) protein (Obeso et al., 2010; Wakabayashi et al., 2007). Currently, there are only symptomatic therapies without alleviation of disease progression.

Given the absence of a disease-modifying therapy for PD (Aldakheel et al., 2014), neurotrophins have elicited great interest as potential therapeutic agents (Figure 1). Several studies have reported reduced NGF and BDNF levels in rodent models and PD patients, correlating them with the disease progression (Fumagalli et al., 2006; Howells et al., 2000; Lorigados et al., 1992; Mogi et al., 1999). Interestingly, in the late stages of the disease, BDNF levels appear to increase, likely reflecting a compensatory mechanism to counteract neuronal damage (Teixeira et al., 2010). BDNF is essential for the maturation and survival of dopamine (DA) neurons, promoting DA release, presynaptic reuptake (Bosse et al., 2012) and protection against neurotoxic insults in both in vitro and in vivo models (Hyman et al., 1991; Tsukahara et al., 1995; Levivier et al., 1995; Yoshimoto et al., 1995). Additionally, BDNF enhances behavioural recovery in rats (Singh et al., 2006) and exhibits anti-apoptotic, anti-oxidative, autophagy-suppressing, and mitochondrial-restorative properties (S. Chen et al., 2017; Miller et al., 2021). Nevertheless, NGF showed initial promise in PD resulting in a longer graft survival after transplantation in putamen and increased functional restoration (Olson et al., 1991; Chaturvedi et al., 2006). However, poor adrenal graft survival and minimal behavioural improvements in clinical studies have stalled its therapeutic development (Hurtig et al., 1989; Peterson et al., 1989).

Glial cell-line derived neurotrophic factor (GDNF), widely recognised as the most potent neurotrophic factor for DA neurons, has shown conflicting results regarding its necessity for their survival in adulthood (Enterría-Morales et al., 2020; Kopra et al., 2015; Pascual et al., 2008). However, its receptor **RET** is essential for the maintenance of adult DA neurons (Kramer et al., 2007). GDNF demonstrates

neurorestorative potential, promoting recovery of damaged DA neurons even when administered several weeks after neurotoxin treatment (Aoi et al., 2000). It surpasses BDNF in promoting the survival of SN pars compacta (SNpc) DA neurons in lesioned animal models (Lu & Hagg, 1997; Rosenblad et al., 2000; Sun et al., 2005). Furthermore, macrophage-mediated GDNF delivery protects against neurodegeneration and reverses motor and non-motor deficits in non-toxin PD models (C. Chen et al., 2019). Various delivery approaches, including infusion (Kirik et al., 2004), GDNF-producing cell transplantation (Shingo et al., 2002) and viral-mediated delivery (Björklund et al., 2000), provide neuroprotective effects in DA neurons in 6-OHDA and MPTP models in rodents and primates (Grondin et al., 2003; Palfi et al., 2002). However, clinical trials in PD patients have yielded unsatisfactory outcomes (Lang et al., 2006; Patel et al., 2013; Whone et al., 2019). Despite these limitations, neurotrophic factor-based therapies remain promising, although their clinical translation faces significant challenges (Chmielarz & Saarma, 2020).

2.3 | Multiple Sclerosis (MS)

MS is a chronic inflammatory demyelinating and neurodegenerative disease of the central nervous system (CNS) (Dobson & Giovannoni, 2019). The aetiology of MS remains unknown, but it likely involves a combination of genetic predisposition and exogenous stimuli. MS pathology is characterised by an altered multidirectional interaction among different immune cell types in the periphery and resident CNS cells, associated with the formation of a glial scar and the focal destruction of the myelin sheath (Bennett & Stüve, 2009; Lucchinetti et al., 1996; McFarland & Martin, 2007). Despite extensive research, a compelling treatment is still elusive, especially for the progressive form of the disease (D'Amico et al., 2016; Ontaneda et al., 2015).

Several studies showed that neurotrophins NGF, BDNF and NT-3 exert an important role in the pathophysiology of MS (Gold et al., 2006) (Figure 1). Administration of BDNF and NT-3 is associated with an increase in the neuronal survival in Experimental Autoimmune Encephalomyelitis (EAE)-induced demyelination (Mo et al., 2010; Yan et al., 1992). EAE is characterised by severe loss of oligodendrocytes and consequent demyelination, probably caused by autoimmune components. Moreover, administration of NT-3 decreases demyelination volume and increases the number of mature oligodendrocytes (OL) in the lysophosphatidylcholine (LPC)-induced demyelination model (Jean et al., 2003), while NGF enhances remyelination in the same model (Althaus, 2004). NGF promotes the survival of mature oligodendrocytes (OL) (Cohen et al., 1996) and prevents the TNF α -induced oligodendrocytes (OL) cell death, limiting the loss of mitochondrial membrane potential (LMP) (Takano et al., 2000). Notably, deficient p75NTR expression alters the composition of cellular infiltrates and exacerbates symptoms in EAE (Coprav et al., 2004; Küst et al., 2006) while enhanced expression of NGF and its receptors was shown in human MS lesions (Aloe, 1998; Aloe et al., 1994; Hurtig

et al., 1989; Laudiero et al., 1992; Peterson et al., 1989; Valdo et al., 2002).

BDNF is the most extensively studied neurotrophin in MS because of its neuroprotective and anti-inflammatory roles (Nociti, 2020). It enhances oligodendrocyte lineage cells, promotes myelin protein synthesis and facilitates axonal regeneration in rodent models (Fulmer et al., 2014; Gravel et al., 1997; McTigue et al., 1998). Exogenous BDNF administration via injection of BDNF-overexpressing T cells at lesion sites reduces EAE severity and provides direct axonal protection (Linker et al., 2010). In demyelinating conditions, BDNF supports oligodendrocyte progenitor proliferation and maturation (Tsiperson et al., 2015; Vondran et al., 2011; Wong et al., 2014) and regulates neuroinflammation via hedgehog and erythropoietin pathways, influencing astrocyte-microglia endogenous crosstalk (Lai et al., 2018). It also downregulates *COX2* and pro-inflammatory cytokines in microglia (Han et al., 2021). However, the therapeutic use of natural neurotrophins is limited by their inability to cross the BBB, poor pharmacokinetic stability and side effects such as dose-dependent hyperalgesia and obesity (Sandrini et al., 2018; Yi et al., 2014). Hence, the development of novel therapeutic candidates that mimic the biological properties of natural neurotrophins while overcoming the aforementioned limitations remains a critical research priority.

2.4 | Amyotrophic lateral sclerosis (ALS)

ALS is a progressive neurodegenerative disease caused by loss of motor neurons. Patients with ALS experience rapid deterioration in muscle function, with an average life expectancy of 3 to 5 years following diagnosis. Currently, there is no cure for ALS. The most effective therapy is a benzothiazole derivative that blocks glutamatergic neurotransmission, Riluzole, which prolongs survival by a few months, thus highlighting the need for new and more efficient drugs. Reduced levels of NTFs have been reported in ALS (Anand et al., 1995; Lee et al., 1996; Nishio et al., 1998; Ono et al., 1999), suggesting that loss of trophic support could be important in disease pathophysiology.

Preclinical studies in ALS mouse models have demonstrated the potential efficacy of NTFs such as BDNF, CNTF and GDNF (Figure 1) (Acsadi et al., 2002; Bemelmans et al., 2006; Ikeda et al., 1995; Li et al., 2007; Mohajeri et al., 1999; Park et al., 2009; Sendtner et al., 1990, 1992; Suzuki et al., 2007; Yan et al., 1992). Investigations into the role of BDNF and its receptor TrkB in ALS have revealed variable expression patterns in *Sod1* mutant mice and ALS patients. In mouse models, muscle BDNF levels have been reported to decrease (Harandi et al., 2016) or increase (Just-Borràs et al., 2019). In ALS patients, BDNF levels increased in muscles but remained unchanged in the spinal cord and CSF (Grundström et al., 2000; Kawamoto et al., 1998; Riolo et al., 2022). Serum BDNF levels showed variability, being reported as increased (Riolo et al., 2022) or unaltered (Cao et al., 2022; Tremolizzo et al., 2016). TrkB levels declined in *SOD1-G93A* mouse muscle (Harandi et al., 2016; Just-Borràs et al., 2019) but increased in ALS patient spinal cords, albeit with

reduced activation (Mutoh et al., 2000). These inconsistent and even opposing changes in BDNF and TrkB expression suggest dysregulated signalling, which may represent either adaptive or maladaptive responses to ALS progression (Just-Borràs et al., 2019; Rei et al., 2022).

BDNF, known for its neuroprotective effects on motor neurons (Koliatsos et al., 1993), advanced into ALS clinical trials via subcutaneous and intrathecal administration (American Neurological Association, 1995; Ochs et al., 2000). However, neither systemic BDNF (25 or 100 µg kg⁻¹) nor its intrathecal delivery showed significant clinical benefits in phase III trials (BDNF Study Group, 1999; Kalra et al., 2003; Beck et al., 2005). Trials using BDNF-secreting cells also failed to improve motor function (Park et al., 2009; J. Wang et al., 2021). These failures may result from the limited ability of BDNF to cross the BBB (Pan et al., 1998; Pardridge, 2003; Poduslo & Curran, 1996) and its capability to activate the p75 receptor, associated with ALS pathophysiology and apoptosis (Lowry et al., 2001; Dupuis et al., 2008b). Notably, p75NTR antagonist slowed ALS progression in *SOD-1* mice (Turner et al., 2004), suggesting that proapoptotic signalling may counteract BDNF's therapeutic effects. Further, post-trial studies using gene therapy in *SOD-1* mice found no effect on disease progression or survival (Park et al., 2009), questioning BDNF's clinical utility.

Ciliary neurotrophic factor (CNTF) has long been known to play a neuroprotective role in several regions of the nervous system. CNTF promotes motor neurons survival in cell or organotypic cultures (Arakawa et al., 1990; Corse et al., 1999), and in vivo after axotomy-induced apoptosis in the rodent (Sendtner et al., 1991, 1994; Thau et al., 2012), while its deletion in mouse models leads to increased vulnerability of motor neurons and atrophy (Giess et al., 2002; Masu et al., 1993). These promising data paved the way for the first clinical trials of CNTF administration for ALS treatment. However, none of these trials had a significant effect on daily living activities outcomes and survival of ALS patients (ALS CNTF Treatment Study Group, 1996; Miller et al., 1996; Turner et al., 2001).

GDNF was investigated later than CNTF and BDNF, with preclinical studies demonstrating motor neuron rescue following axotomy and excitotoxic stress (Bilak et al., 2001; Buj-Bello et al., 1995; Giménez Y Ribotta et al., 1997; Kosuge et al., 2009). In the *SOD-1* animal model of ALS, GDNF showed motor neuron protection but yielded variable outcomes on survival and neuromuscular junction integrity (Acsadi et al., 2002; Li et al., 2007; Mohajeri et al., 1999; Park et al., 2009; Suzuki et al., 2007). Although some studies reported improved disease progression and survival, others found no significant benefits (Acsadi et al., 2002; Park et al., 2009). Clinical trials using direct GDNF protein injections via subcutaneous or intrathecal routes showed no therapeutic effects because of its short half-life and limited tissue diffusion (Alisky & Davidson, 2000). A recent phase I/II trial addressed these limitations by injecting human neural progenitor cells (hNPC) engineered to secrete GDNF into the lumbar spinal cord, demonstrating safety and localised expression (Baloh et al., 2022). Based on evidence supporting cortical delivery, a new clinical trial is evaluating hNPCs-GDNF in the motor cortex.

2.5 | Diabetic retinopathy (DR)

DR is one of the most common long-term complications of diabetes that results in loss of visual acuity and blindness. Its multifactorial pathogenesis involves endothelial dysfunction, disruption of the blood-retinal barrier, oxidative stress, and inflammation driven by reactive oxygen species, advanced glycation end products, and a plethora of inflammatory mediators (Kowluru et al., 2015; Kowluru & Chan, 2007; Yu et al., 2015). Additionally, imbalance of retina's homeostatic equilibrium significantly contributes to DR development (Antonetti et al., 2012).

The imbalance between pro-survival neurotrophic factors and inflammatory components contributes to apoptosis and pro-inflammatory responses in DR (Antonetti et al., 2006). Alterations in NGF and BDNF levels have been observed in both patients and animal models of DR (Ali et al., 2011; Kaviarasan et al., 2015; Mysona et al., 2015; Ola et al., 2013). A proNGF/NGF imbalance marked by proNGF accumulation and reduced NGF levels has been implicated in retinal inflammation, neurodegeneration and BBB integrity (Al-Gayyar et al., 2011, 2013; Ali et al., 2011; Mysona et al., 2015). NGF administration prevents ganglion cell death in diabetic rats and restores TrkA levels in the retina, protecting retinal ganglion cells (RGCs) from degeneration in experimental model of diabetes (Colafrancesco et al., 2011; Hammes et al., 1995). Topical ophthalmic administration of NGF also protects RGCs in an animal model of glaucoma, in patients with glaucoma (Lambiase et al., 2009) and in the streptozotocin (STZ) model of DR (Mantelli et al., 2014). Furthermore, topical application of recombinant human-NGF significantly improves RGC survival and inhibits astrocyte activity in the optic nerve in a rat model of DR (Guo et al., 2020). These findings highlight the potential of NGF restoration as a therapeutic strategy to counteract early diabetic retinopathy.

Hyperglycaemia-mediated increased secretion of glucocorticoids and pro-inflammatory cytokines down-regulate the levels of BDNF (Kumamaru et al., 2008; Numakawa et al., 2010). Down-regulation of BDNF neuroprotective actions under hyperglycaemia renders retinal neurons vulnerable to damaging stimuli, leading to diabetic retinopathy (Ola et al., 2013). BDNF administration protects retinal neurons from hyperglycaemia in a dose-dependent manner in vitro (Liu et al., 2013). BDNF can also efficiently rescue DA amacrine cells from neurodegeneration and counteract the down-regulation of tyrosine hydroxylase (TH) levels seen under diabetic conditions in vivo (Seki et al., 2004). Intravitreal injections of pAAV-BDNF reduced RGC apoptosis and improved function in STZ-induced diabetic rats (Gong et al., 2012). Hence, restoring BDNF levels, either by counteracting hyperglycaemia-induced pathologic factors or administering exogenous BDNF, may prevent retinal neurodegeneration and retard disease progression. However, randomised, double-blind, placebo-controlled trials failed to demonstrate significant effects of subcutaneous NGF or BDNF injections for diabetic polyneuropathy (Apfel et al., 2000; Wellmer et al., 2001).

It is of interest that during retinal degeneration because of glaucoma or optic nerve transection, treatment with a mutant NGF that only activates TrkA, or with a biological response modifier that prevents

endogenous NGF and pro-NGF from binding to the p75NTR receptor, affords significant neuroprotection. On the contrary, treatment of normal eyes with an NGF mutant-selective p75NTR agonist causes progressive RGC death, and in injured eyes accelerated RGC death (Bay et al., 2010). The intravitreal or subconjunctival depot of THX-B, a small molecule antagonist of the p75NTR receptor, was therapeutic and resolved the inflammatory, vascular and neurodegenerative phases of the retinal pathology of diabetic retinopathy (Galan et al., 2017; Platón-Corchado et al., 2017). Another approach used an engineered NT3-based mutant that broadly activates TrkA, TrkB and TrkC receptors while minimising p75^{NTR} receptor binding provides enhanced neuroprotection in vivo by selectively promoting Trk-mediated pro-survival signalling and avoiding p75^{NTR}-induced toxicity in neurodegenerative disease models (Brahimi et al., 2021). Furthermore, targeting TrkC.T1 with isoform-selective small-molecule antagonists effectively reduces NT3-induced neurotoxicity, suppresses TNF- α elevation and preserves photoreceptors in a retinitis pigmentosa model, highlighting a novel therapeutic strategy centred on the modulation of receptor isoform-specific signalling to achieve neuroprotection (Brahimi et al., 2025).

3 | NEUROTROPHIN MIMETICS: A NOVEL APPROACH IN PHARMACOLOGY OF NEURODEGENERATION

3.1 | DHEA as an ancestral neurotrophin

In the early 1980s, Etienne Baulieu discovered the ability of neurons and glia to locally produce steroids by metabolising hormone precursors or synthesising steroids from cholesterol (Baulieu & Robel, 1990). Neurosteroids, produced by neurons, astrocytes and neuroglial cells, in the central and peripheral nervous systems, influence neuronal activity and differentiation, provide neuroprotection and promote neurogenesis (Compagnone & Mellon, 2000). The initiation of neurosteroid activities is thought to occur through the activation of receptors such as GABA_A, NMDA and sigma-1 (Charalampopoulos, Margioris, & Gravanis, 2008; Charalampopoulos, Remboutsika, et al., 2008; Compagnone & Mellon, 2000).

Dehydroepiandrosterone (DHEA), the first characterised neurosteroid, serves as a precursor for androgens and oestrogens. Synthesised by the cytochrome 17 enzyme (CYP17) from cholesterol, its biosynthesis occurs in the gonads, adrenal glands and locally in the brain. Adrenal DHEA has a systemic function, while brain-generated DHEA acts locally in a paracrine manner, with age-dependent synthesis peaking in early adulthood and declining over time, especially in neurodegenerative diseases like Alzheimer's (Baulieu, 1998; Schumacher et al., 2003; Weill-Engerer et al., 2002).

Although no specific DHEA receptor was identified during the first decades of its characterisation, early studies revealed that DHEA is recognised with high affinity and a K_d at nanomolar level by membrane binding sites, mediating the activation of MEK/ERK and PI3K/Akt post-receptor signalling pathways, leading to the phosphorylation and mobilisation of transcription factors CREB and NFkappaB. The

latter control the transcription of anti-apoptotic **Bcl-2** proteins or the phosphorylation/deactivation of pro-apoptotic Bad proteins, protecting neuronal cells from apoptosis (Charalampopoulos et al., 2004, 2006; Charalampopoulos, Remboutsika, et al., 2008). PC12 cells, lacking GABA_A or NMDA receptors and unable to metabolise DHEA into sex hormones, serve as a model for studying the membrane effects of DHEA in neuronal cell types (Greene & Tischler, 1976). DHEA, like NGF, prevents apoptosis through strikingly similar signal transduction pathways, suggesting a role for NGF receptors in the anti-apoptotic action of DHEA (Lazaridis et al., 2011; Riccio et al., 1999). Our group provided evidence that DHEA binds with high affinity (K_d at nanomolar level) and activates both membrane receptors of NGF, specifically TrkA and p75NTR, marking the first evidence of direct steroid binding to neurotrophin receptors (Lazaridis et al., 2011). Upon DHEA binding to TrkA, the receptor undergoes autophosphorylation at tyrosine residues, initiating a signalling pathway involving Shc-PI3K-Akt and Src-MEK-ERK kinases. Additionally, DHEA binding to the p75NTR receptor influences its interaction with effector proteins, such as TRAF6, **RIP2** and **RhoGDI**. The balance between TrkA and p75NTR receptors ultimately determines whether the cell will undergo apoptosis or survival (Lazaridis et al., 2011).

DHEA exhibits neuroprotective and pro-survival effects, promoting neurogenesis by increasing newly formed neurons in both rats and human neural stem cell (NSC) cultures. Its impact on NMDA and Sigma-1 receptors contributes to long-term NSC proliferation (Charalampopoulos, Remboutsika, et al., 2008). Pediaditakis et al. (2015) identified DHEA as an ancestral ligand for neurotrophin receptors, demonstrating its ability to interact with and activate vertebrate Trk receptors (TrkA and TrkC) and all invertebrate Trk receptors (Pediaditakis et al., 2015). Administration of DHEA to animal models of MS or PD had beneficial effects (Aggelakopoulou & Kourepini, 2016; Bélanger et al., 2006, 2006), which could be attributed to its neuroprotective (Charalampopoulos et al., 2004, 2006; Charalampopoulos, Margioris, & Gravanis, 2008; Charalampopoulos, Remboutsika, et al., 2008; Gravanis et al., 2012; Lazaridis et al., 2011; Maninger et al., 2009; Xie et al., 2019) as well as anti-inflammatory properties (Du et al., 2001; Maninger et al., 2009; Saijo et al., 2011; Straub et al., 1998; Hoyk et al., 2004; Hazeldine et al., 2010), including mitigation of microglia-mediated neuroinflammation (Alexaki et al., 2018; Saijo & Glass, 2011). However, the long-term administration of DHEA in these conditions has not proven beneficial, as it is metabolised in humans into oestrogens and androgens and their numerous metabolites, scrambling the endocrine system with potential side effects and increasing the risk for hormone-dependent neoplasias (Calogeropoulou et al., 2009; Compagnone & Mellon, 2000).

3.2 | DHEA derivatives as neurotrophin mimetics (microneurotrophins)

Based on the intriguing interactions of DHEA with the neurotrophin receptors system, our group designed, synthesised and screened a large chemical library of 17-spirocyclic steroidal analogues of DHEA for their ability to protect neurons against apoptosis and activate

neurotrophin receptors (Figure 2). The rational design of the compound series left unchanged the 3 β -OH group of DHEA because it is important for its neuroprotective properties (Calogeropoulou et al., 2009; Charalampopoulos et al., 2004), and the modifications were focussed on the C17 position important for steroidogenesis, thus avoiding steroid-like side effects. Synthetic 17-spiro derivatives of DHEA, devoid of endocrine effects (Calogeropoulou et al., 2009), form a novel group of non-toxic compounds capable of crossing the BBB and activating neurotrophin receptors, exerting in turn strong neuroprotective effects. Two BBB-permeable, 17-spiro-epoxy derivatives of DHEA, BNN27 and BNN20, were proven very effective as neuroprotective agents in various neuronal cell systems (Calogeropoulou et al., 2009). These analogues, termed 'microneurotrophins', act as activators of neurotrophin receptors.

BNN27 (17 α ,20R-epoxy-5-pregnene-3 β ,21-diol), the most studied microneurotrophin, derived from DHEA (Figure 2), activates the TrkA receptor by inducing its phosphorylation at tyrosine residues, initiating pro-survival signalling in cells (Pediaditakis, Efstathopoulos, et al., 2016). Binding studies confirm high affinity of BNN27 for both TrkA and p75NTR receptors, preventing apoptosis through the regulation of RhoGDI, TRAF6 and RIP2 protein interactions (Pediaditakis, Kourgiantaki, et al., 2016). BNN27 enhances the efficacy of low levels of NGF, promoting axonal outgrowth by facilitating the fast membrane re-entry of internalised TrkA receptor (Pediaditakis, Efstathopoulos, et al., 2016). Furthermore, BNN27 physically interacts with the p75NTR receptor in specific amino-residues of its extracellular domain, inducing survival of primary neuronal cells through the recruitment of p75NTR downstream effectors RIP2 and RhoGDI (Pediaditakis, Kourgiantaki, et al., 2016). Activation of p75NTR by BNN27 resulted also to anti-apoptotic effects, because of the decrease of phosphorylation of pro-apoptotic JNK kinase and to the cleavage of Caspase-3 in cerebellar granule neurons (Pediaditakis, Kourgiantaki, et al., 2016). Importantly, BNN27 does not show the hyperalgesic effects of NGF (Poulaki et al., 2021). BNN27 was also effective to enhance memory, to interact with the cholinergic system, to protect oligodendrocytes and myelin in demyelinating disorders, and provide therapeutic benefits for diabetic retinopathy by addressing neurodegeneration and inflammation (Bonetto et al., 2017; Ibán-Arias et al., 2018; Pitsikas & Gravanis, 2017).

3.2.1 | Microneurotrophins in animal models of neurodegenerative diseases

The aforementioned promising data of BNN27 action paved the way for extensive research on this microneurotrophin, as a potential lead molecule to develop a therapeutic strategy for a number of neurodegenerative diseases. Various microneurotrophins were thus tested in various animal models of human neurodegenerative diseases (mechanism of actions and effects are summarised in Figure 3). BNN27 inhibits apoptosis of NGF-deprived sympathetic neurons and reverses apoptosis of TrkA-sensory neurons in E13.5 NGF-deficient mouse embryos (Pediaditakis, Efstathopoulos, et al., 2016). Systemic

FIGURE 2 Chemical structure of potent synthetic microneurotrophins.

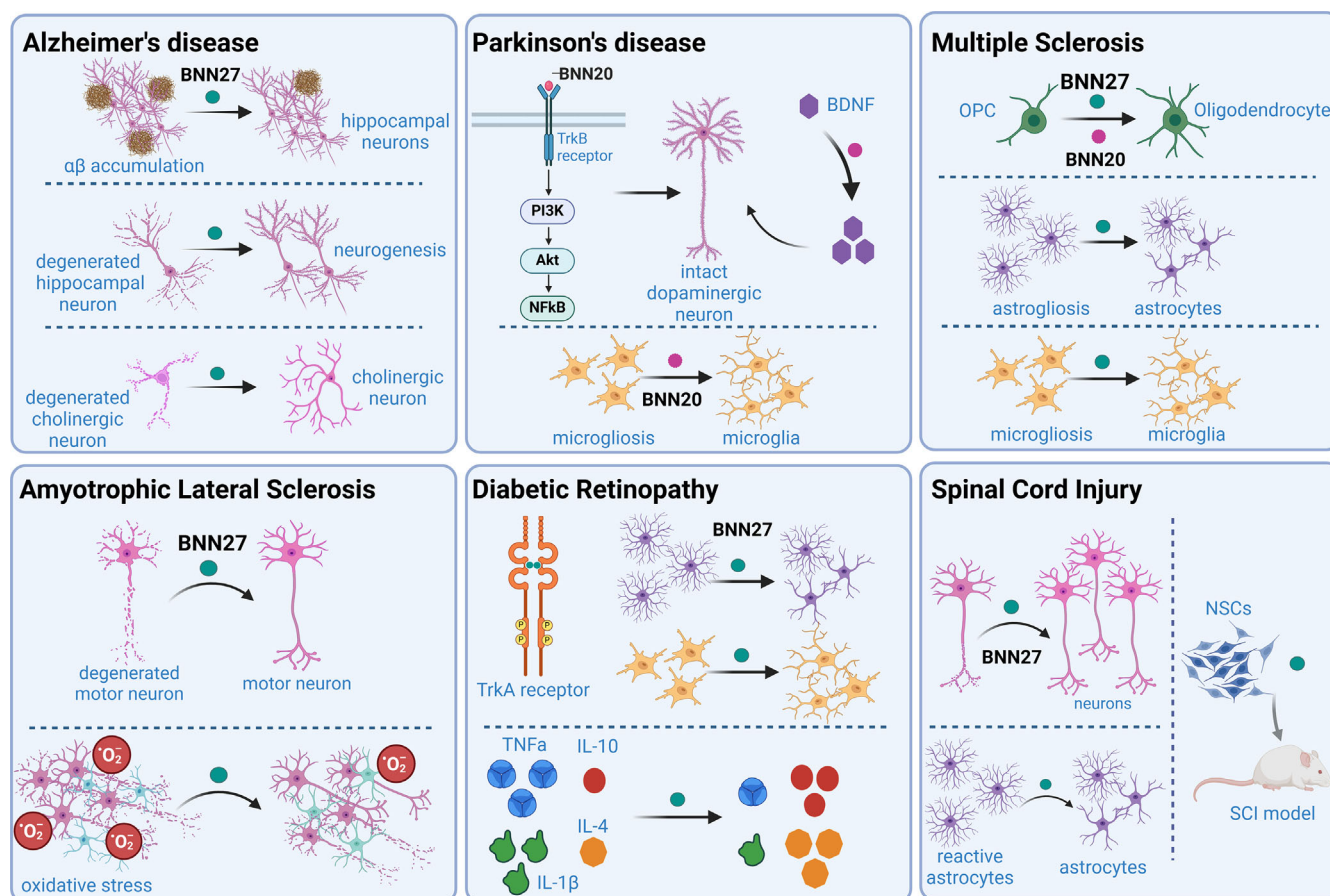
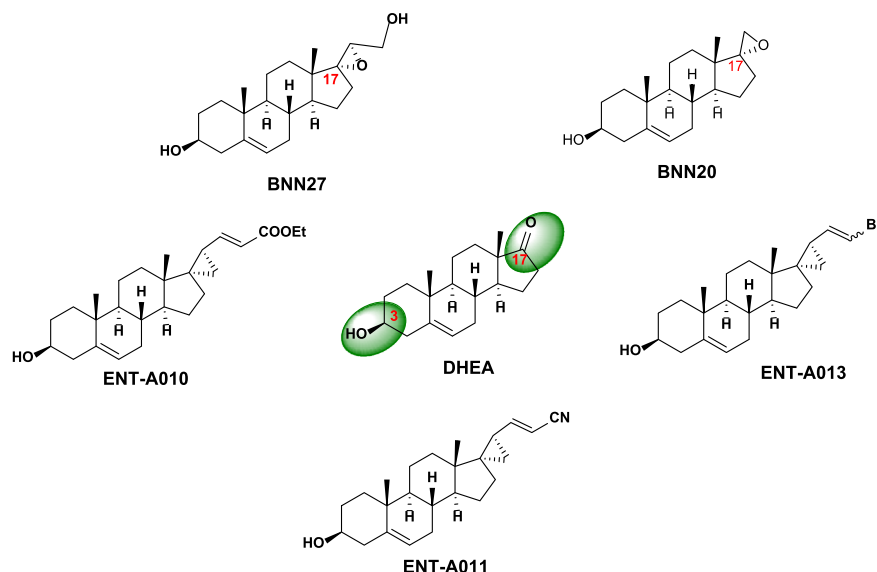


FIGURE 3 Schematic overview of the multifunctional roles of microneurotrophins BNN27 and BNN20 in neurodegenerative diseases. BNN27 exhibits diverse therapeutic potential across multiple neurodegenerative conditions. In Alzheimer's disease models, BNN27 reduces amyloid- β accumulation, promotes neurogenesis and provides neuroprotection. In models simulating multiple sclerosis, it facilitates oligodendrocyte precursor cells (OPC) maturation and attenuates inflammatory responses. In ALS models, BNN27 demonstrates neuroprotective and antioxidant properties. Additionally, in diabetic retinopathy, BNN27 reduces microgliosis, astroglia and pro-inflammatory cytokines while increasing anti-inflammatory cytokine levels. It enhances neuronal survival and mitigates astroglia in spinal cord injury (SCI) models, showing promise as a candidate for combinatorial therapies with neural stem cell (NSC) transplants. BNN20 exerts neuroprotection in Parkinson's disease models by activating the PI3K/Akt/NF- κ B pathway and increasing BDNF levels. Moreover, BNN20 promotes OPC maturation highlighting its potential in the treatment of demyelinating diseases such as multiple sclerosis. Created in BioRender.

administration of BNN27 can also significantly reduce astrogliosis and increase neuronal density following dorsal column spinal cord injury (SCI) in mice at 12 weeks post injury. Furthermore, combined administration of BNN27 with NSC-seeded collagen-based scaffold grafts leads to increased density of survived implanted NSC-derived cells, making it an appealing candidate for combinatorial neuroregenerative therapeutic strategies (Georgelou et al., 2023).

3.2.2 | Microneurotrophins in the rat streptozotocin model of diabetic retinopathy

BNN27 was shown to partially but statistically significantly reverse diabetes-induced retinal damage in the experimental rat streptozotocin (STZ) model of diabetic retinopathy by activating the TrkA receptor, decreasing pro-death p75NTR receptor levels, inducing cleavage of pro-apoptotic **caspase-3** and reducing glial activation. BNN27 systemic intraperitoneal administration also reduced the pro-inflammatory **TNF α** and **IL-1b** and increased the anti-inflammatory **IL-10** and **IL-4** cytokine levels (Ib  n-Arias et al., 2018). Targeting both the neurodegenerative and inflammatory components of the disease, microneurotrophin BNN27 seems to have the pharmacological profile for the design of promising novel therapeutic agent for DR. To follow up on these findings, the efficacy of BNN27 when administered topically as eye drops in diabetic rats was also investigated. Indeed, BNN27 reversed diabetes-induced damage in RGC axons and amacrine neurons expressing NFL and bNOS/TH, respectively, in a dose-dependent manner. The involvement of TrkA receptor activation and subsequent phosphorylation of the downstream ERK1/2 kinase signalling pathway in mediating the pro-survival effects of BNN27 was confirmed in this system, consistent with previous findings of systemic BNN27 administration (Ib  n-Arias et al., 2019). Hence, topical administration of BNN27 may offer an effective and less invasive treatment for DR by restoring the balance between pro-survival neurotrophic and inflammatory factors while minimising systemic effects (Ib  n-Arias et al., 2019).

On the contrary, systemic BNN27 administration in an experimental retinal detachment-induced mouse model resulted in increased gliosis and was insufficient to protect photoreceptors from cell death (Tsoka et al., 2018). It is possible that a different dose of BNN27 accessing the retina may be responsible for the lack of protection when added systemically. Moreover, differences in response to BNN27 may be because of the different pathophysiology pathways and cell types involved in DR and retinal detachment. Indeed retinal detachment has characteristics of an acute ischaemic trauma to the retina, while DR is a chronic metabolic diabetic condition.

3.2.3 | Microneurotrophins in the rat model of pharmacologically-induced schizophrenia

Given the antioxidant and anti-inflammatory properties of BNN27, along with its modulatory action on neurotrophins levels (Glajch et al., 2016; Ib  n-Arias et al., 2018; Padiaditakis, Efstathopoulos, et al.,

2016; Padiaditakis, Kourgiantaki, et al., 2016) and on factors which have been associated with schizophrenia (Bitanirwe & Woo, 2011; Khandaker et al., 2015; Rodrigues-Amorim et al., 2018), the ability of BNN27 to counteract schizophrenia-like behavioural deficits produced by ketamine (an NMDA receptor antagonist) in rats has also been investigated. Interestingly, BNN27 was capable of attenuating ketamine-induced ataxia and hypermobility and counteracting ketamine-induced non-spatial and spatial recognition memory deficits. Simultaneously, it reduced social isolation produced by glutamate hypofunction (Zoupa et al., 2019). A functional interaction between BNN27 and the DA system has also been illustrated by the effectiveness of BNN27 in repairing DA dysfunction caused by apomorphine, attenuating cognitive impairments induced by this D1/D2 receptor agonist in rats (Pitsikas et al., 2021). These findings support a potential role of BNN27 in the alleviation of some behavioural alterations related to schizophrenia. Notably, BNN27 appears to lack anxiolytic or antidepressant properties (Kokras et al., 2020), aligning with findings from a clinical trial involving DHEA in patients with schizophrenia, where DHEA improved negative symptoms but not depressive symptoms or anxiety (Cai et al., 2018). However, DHEA affects locomotion, progesterone and testosterone levels, as well as the glutamatergic and GABAergic systems of the hippocampus and prefrontal cortex in a sex-dependent manner (Kokras et al., 2020).

3.2.4 | Microneurotrophins in the cuprizone mouse model of demyelination

Research on the role of BNN27 on mouse glial population has revealed that it exerts trophic effects on mature oligodendrocytes, protecting them from cuprizone-induced apoptosis, in a TrkA-dependent manner. In vitro, BNN27 promotes oligodendrocyte maturation and reduces microglial activation. In the in vivo cuprizone-induced mouse demyelination model, BNN27 preserves mature oligodendrocytes and mitigates microgliosis and astrogliosis, while it has no impact on the remyelination process (Bonetto et al., 2017). Furthermore, microneurotrophin BNN20, a C17-spiroepoxy DHEA analogue (Figure 2), was shown to promote oligodendrocyte maturation possibly through TrkB receptors, while it diminished astrocytic accumulation, leading to a faster remyelination process in the LPC-induced demyelination mouse model (Kalafatakis et al., 2021). Our findings suggest that microneurotrophins BNN27 and BNN20 may serve as lead molecules to develop non-toxic, BBB-permeable synthetic activators of neurotrophin receptors that could prove effective in demyelinating and neurodegenerative diseases, such as MS.

3.2.5 | Microneurotrophins in the SOD1 mouse model of amyotrophic lateral sclerosis (ALS)

The promising neuroprotective effects of BNN27 in various neuronal cell types drove us to test its effects in an animal model of devastating ALS. In the SOD1-G93A mouse model of ALS, BNN27 administration,

via subcutaneously pellet implants containing either placebo or BNN27, mitigated motor impairments in the Paw grip endurance (PaGE) and rotarod tasks in female mice, while no effects were observed in males. However, BNN27 treatment did not affect other behavioural or neuropathological markers in either sex (Glajch et al., 2016).

However, BNN27 attenuated the loss of motor neurons and promoted their survival in co-cultures with astrocytes derived from ALS patients with SOD1 mutations, via the reduction of oxidative stress (Glajch et al., 2016). The differences in the effects of BNN27 in rat and human models of ALS might be because of species differences in BNN27 bioavailability and metabolism, with much faster degradation in mouse hepatocytes than in humans.

3.2.6 | Microneurotrophins in the 5xFAD mouse model of Alzheimer's disease

The neurogenic and neuroprotective properties of the microneurotrophin BNN27 have been explored in the 5xFAD mouse model of AD. The compound significantly reduced toxic amyloid- β deposition in whole brain of the animals and decreased inflammation. BNN27 treatment restored the size of cholinergic neurons and decreased the population of p75NTR⁺/ChAT⁺ neurons, exerting a neuroprotective effect. BNN27 also enhanced adult hippocampal neurogenesis and synaptogenesis, finally restoring working memory and leading to cognitive improvement. Moreover, BNN27 administration in both isolated hippocampal NSC and hippocampal neuronal cultures exhibited a strong anti-apoptotic effect against amyloid- β -induced toxicity (Kokkali et al., 2024). Overall, BNN27 offers a multifactorial, BBB-permeable, non-endocrine, non-toxic lead molecule to design NGF analogues with beneficial effects in the neuronal loss and memory deficits associated to AD (Kokkali et al., 2024; Bennett et al., 2016; Pediaditakis, Efsthathopoulos, et al., 2016; Pediaditakis, Kourgiantaki, et al., 2016). Additionally, the successful intranasal delivery of BNN27 via mucoadhesive liposomes or nanoemulsions enables a rapid and enhanced nose-to-brain compound transport, presenting a noninvasive, patient-friendly strategy for an efficient therapeutic administration (Kannavou et al., 2023).

It is of note that BNN27 plays a crucial role in recognition memory, and its interaction with the cholinergic system seems to be relevant to cognition. Indeed, the compound effectively counteracted delay dependent and scopolamine-induced (a muscarinic cholinergic receptor antagonist) non-spatial and spatial recognition memory deficits in rats (Pitsikas & Gravanis, 2017).

Recent studies introduced novel NGF mimetics possessing a substituted 17-spirocyclopropyl functionality, namely ENT-A013 and ENT-A010 (Figure 2), which surpass BNN27 in metabolic stability and potency (Rogdakis et al., 2022; Yilmaz et al., 2022). These recent microneurotrophins prevent synaptic loss and amyloid-induced apoptosis in hippocampal neurons and promote survival of primary dorsal root ganglion (DRG) neurons after NGF removal (Rogdakis et al., 2022; Yilmaz et al., 2022). ENT-A013 selectively

activates the TrkA receptor, exhibiting neuroprotective and anti-amyloid effects by activating TrkA and downstream kinases Akt and Erk1/2 (Rogdakis et al., 2022). Nevertheless, systemically administered ENT-A010 affects the morphology, transcriptional profile and homeostasis of hippocampal microglia in LPS-treated mice in a TrkA-dependent manner. It also increases amyloid-beta uptake and NGF expression in primary microglia in vitro (Yilmaz et al., 2022). These findings highlight these small molecules as promising candidates for targeting TrkA-mediated pro-survival signalling in neurodegenerative diseases including AD.

Both TrkA and p75NTR receptors for NGF play significant roles in AD pathology, with altered expression in the hippocampus, particularly in cholinergic neurons (Bruno et al., 2023; Ginsberg et al., 2019; Hefti & Will, 1987). TrkA ablation in the basal forebrain disrupts cholinergic function (Sanchez-Ortiz et al., 2014), while p75NTR deletion can rescue synaptic loss, amyloid- β accumulation and cognitive decline in AD models (Demuth et al., 2023; Qian et al., 2019). Notably, p75NTR mediates neurotoxic and pro-inflammatory effects of A β , exacerbating AD progression (Knowles et al., 2009; Perini et al., 2002). ENT-A044, the novel neurotrophin analogue, exhibits context-dependent effects on p75NTR, inducing apoptosis in human NSCs but promoting survival of mouse neurons, offering a potential new class of pharmacological agents in AD therapeutics (Papadopoulou et al., 2023).

It is well established that BDNF holds a pivotal role in neurogenesis, synaptic plasticity and repair (De la Rosa et al., 2019; Kramár et al., 2012; Miranda et al., 2019). Hence, research efforts have increasingly focussed on the development of neuroprotective small molecules that selectively target TrkB receptor (Antonijevic et al., 2024; Narducci et al., 2023). Our synthetic compound ENT-A011, a novel BDNF mimetic, demonstrated the ability to protect mouse and human neurons and their precursors against amyloid- β (A β)-induced toxicity while it propagates the proliferation of mouse hippocampal and human-induced pluripotent stem cell-derived neurons, mitigating cell death in human neural progenitor cells (hNPCs) exposed to A β toxicity, primarily by modulating a core gene network overlapping with that of BDNF, as identified through RNA sequencing (Charou et al., 2024).

3.2.7 | Microneurotrophins in the Weaver mouse model of PD

Microneurotrophin BNN20 (Figure 2) has demonstrated neuroprotective effects both in vitro and in vivo in Weaver mice, an animal model of PD. BNN20 was shown to activate both TrkA and TrkB receptors mimicking the effects of NGF and BDNF, respectively, and efficiently inducing the association of the p75NTR receptor with its effector protein RIP2 (Botsakis et al., 2017; Calogeropoulou et al., 2009; Lazaridis et al., 2011). It was effective in protecting DA neurons and their terminals in vivo, in 'Weaver' mice by activating neurotrophin receptor TrkB, inducing its pro-survival, post-receptor PI3K-Akt-NF κ B signalling pathway. BNN-20 exerted its beneficial effect by strongly

TABLE 1 Neuroprotective, anti-neuroinflammatory and neurogenic effects of microneurotrophins.

In vivo animal model	Human disease	Microneurotrophins
Cuprizone mice Bonetto et al. <i>Glia</i> 2017 Kalafatakis et al. <i>J Neurosci Res</i> 2021 Poulai et al. <i>Biomedicines</i> 2021	Multiple sclerosis demyelination	BNN27 BNN20
Weaver transgenic mice Botsakis et al. <i>Neuropharmacology</i> 2017 Panagiotakopoulou et al. <i>Neuropharmacology</i> 2020	Parkinson's disease	BNN20
Streptozotocin rats Iban-Arias et al. <i>Diabetes</i> 2017 Iban-Arias et al. <i>Arch Clin Exp Ophthalmol</i> 2021 Tsika et al. <i>Pharmacol Res Perc</i> 2021	Diabetic retinopathy	BNN27
SOD1 transgenic mice	Amyotrophic lateral sclerosis	BNN27
5xFAD transgenic mice Kokkali et al. <i>Mol Psychiatry</i> 2024 Pitsikas & Gravanis <i>Neurobiol Learn Mem</i> 2017 Zoupa et al. <i>Neuropharmacology</i> 2019 Pitsikas et al. <i>Psychopharmacology</i> 2021 Pitsikas et al. <i>Behav Brain Res</i> 2022 Rogdakis et al. <i>Biomedicines</i> 2022 Yilmaz et al. <i>Biomolecules</i> 2022 Papadopoulou et al. <i>Int J Mol Sci</i> 2023 Charou et al. <i>Stem Cell Res and Ther</i> 2024	Alzheimer's disease, cognitive impairment	BNN27 ENT-A010 ENT-A011 ENT-A013 ENT-A044

elevating the anti-apoptotic Bcl-2/Bax ratio, inducing an anti-inflammatory and antioxidant activity and restoring BDNF levels (Botsakis et al., 2017). Notably, the strong anti-neuroinflammatory effect of BNN20 in Weaver mice seems to be also mediated through microglia, because it effectively counteracted the activation of microglia, inducing its shift towards an M2 neuroprotective stage (Panagiotakopoulou et al., 2020).

4 | CONCLUSION

The mechanisms underlying the onset of most neurodegenerative diseases remain unclear, limiting the development of systematic therapies effective enough to control or even reverse these disorders. The important, although complex, involvement of neurotrophins in promoting neuronal protection and neuronal and myelin regeneration, in addition to their anti-neuroinflammatory effects controlling the activation of glia, offers a promising avenue for a symptomatic therapeutic approach of neurodegenerative and de-myelinating disorders. However, limitations such as neurotrophin short half-lives, their difficulty crossing the BBB and their low pharmacokinetic stability hinder their clinical applications, particularly for long-term administration, necessary for these chronic diseases. Synthetic mimetics of endogenous neurotrophins and activators of their receptors, such as microneurotrophins hold potential as a multimodal symptomatic strategy to enhance in parallel neuronal survival, neuronal and myelin regeneration, inhibition of gliosis and neuro-inflammation, targeting the main neurobiological endpoints of these diseases. Microneurotrophins have been pre-clinically tested in various animal models of various neurodegenerative diseases (relevant publications summarised in Table 1) and may offer new therapeutic agents to reverse or compensate deficits and impairments associated with neurodegenerative diseases and demyelinating disorders in a more effective and less invasive way than the exogenous transplantation of neural and oligodendrocyte precursors or administration of natural neurotrophins, with better pharmacokinetic and pharmacodynamic profiles. Multifaceted neurotrophin mimetics and their combinations provide the pharmacological basis for proposing a brand-new symptomatic therapeutic approach to neurodegenerative diseases to simultaneously control neuronal death, demyelination and neuroinflammation while boosting neuronal repair through neurogenesis. Experimental evidence suggests a role for neurotrophic factors, including NGF and BDNF, in the progression of pancreatic cancer and the mechanisms by which the sympathetic and parasympathetic nervous systems regulate tumour cell growth, migration and invasion (Xu et al., 2024). It is thus important to test the effects of microneurotrophins, activators of TrkA and TrkB receptors for NGF and BDNF, in pathophysiology models of this devastating cancer.

4.1 | Nomenclature of targets and ligands

Key protein targets and ligands in this article are hyperlinked to corresponding entries in the IUPHAR/BPS Guide to PHARMACOLOGY <http://www.guidetopharmacology.org> and are permanently archived in the Concise Guide to PHARMACOLOGY 2023/24 (Alexander, Christopoulos, Davenport, Kelly, Mathie, Peters, Veale, Armstrong, Faccenda, Harding, Davies, et al., 2023; Alexander, Fabbro, Kelly, Mathie, Peters, Veale, Armstrong, Faccenda, Harding, Davies, Annett, et al., 2023; Alexander, Fabbro, Kelly, Mathie, Peters, Veale, Armstrong, Faccenda, Harding, Davies, Beuve, et al., 2023; Alexander, Kelly, Mathie, Peters, Veale, Armstrong, Buneman, Faccenda, Harding,

Spedding, Cidlowski, et al., 2023; Alexander, Mathie, Peters, Veale, Striessnig, Kelly, Armstrong, Faccenda, Harding, Davies, Aldrich, et al., 2023).

AUTHOR CONTRIBUTIONS

Ioanna Zota: Investigation, Methodology and Writing – original draft. **Theodora Calogeropoulou:** Conceptualisation, Funding acquisition, Validation, Review and Editing. **Konstantina Chanoumidou:** Investigation, Methodology and Review. **Ioannis Charalampopoulos:** Conceptualisation, Funding acquisition, Validation, Review and Editing. **Achille Gravanis:** Conceptualisation, Funding acquisition, Supervision, Validation, Writing and Editing.

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CONFLICT OF INTEREST STATEMENT

The authors declare no conflicts of interest, except Achille Gravanis as the cofounder of BioNature E.A. LTD and proprietor of compounds BNN27 and BNN20 (patented with the WO 2008/1555 34 A2 number at the World Intellectual Property Organization).

DATA AVAILABILITY STATEMENT

N/A Review.

REFERENCES

- Acsadi, G., Anguelov, R. A., Yang, H., Toth, G., Thomas, R., Jani, A., Wang, Y., Ianakova, E., Mohammad, S., Lewis, R. A., & Shy, M. E. (2002). Increased survival and function of SOD1 mice after glial cell-derived neurotrophic factor gene therapy. *Human Gene Therapy*, 13(9), 1047–1059. <https://doi.org/10.1089/104303402753812458>
- Aggelakopoulou, M., & Kourepini, E. (2016). Er β -dependent direct suppression of human and murine Th17 cells and treatment of established central nervous system autoimmunity by a neurosteroid. *The Journal of Immunology*, 197, 2598–2609. <https://doi.org/10.4049/jimmunol.1601038>
- AIDakheel, A., Kalia, L. V., & Lang, A. E. (2014). Pathogenesis-targeted, disease-modifying therapies in Parkinson disease. *Neurotherapeutics: The Journal of the American Society for Experimental NeuroTherapeutics*, 11(1), 6–23. <https://doi.org/10.1007/S13311-013-0218-1>
- Alexaki, V. I., Fodelianaki, G., Neuwirth, A., Mund, C., Kourgiantaki, A., Ieronimaki, E., Lyroni, K., Troullinaki, M., Fujii, C., Kanczkowski, W., Ziogas, A., Peitzsch, M., Grossklaus, S., Sönnichsen, B., Gravanis, A., Bornstein, S. R., Charalampopoulos, I., Tsatsanis, C., & Chavakis, T. (2018). DHEA inhibits acute microglia-mediated inflammation through activation of the TrkA-Akt1/2-CREB-Jmjd3 pathway. *Molecular Psychiatry*, 23(6), 1410–1420. <https://doi.org/10.1038/MP.2017.167>
- Alexander, S. P. H., Christopoulos, A., Davenport, A. P., Kelly, E., Mathie, A. A., Peters, J. A., Veale, E. L., Armstrong, J. F., Faccenda, E., Harding, S. D., Davies, J. A., Abbraccio, M. P., Abraham, G., Agoulnik, A., Alexander, W., Al-Hosaini, K., Bäck, M., Baker, J. G., Barnes, N. M., ... Ye, R. D. (2023). The Concise Guide to PHARMACOLOGY 2023/24: G protein-coupled receptors. *British Journal of Pharmacology*, 180, S23–S144. <https://doi.org/10.1111/bph.16177>
- Alexander, S. P. H., Fabbro, D., Kelly, E., Mathie, A. A., Peters, J. A., Veale, E. L., Armstrong, J. F., Faccenda, E., Harding, S. D., Davies, J. A., Annett, S., Boison, D., Burns, K. E., Dessauer, C., Gertsch, J., Helsby, N. A., Izzo, A. A., Ostrom, R., Papapetropoulos, A., ... Wong, S. S. (2023). The concise guide to PHARMACOLOGY 2023/24: Enzymes. *British Journal of Pharmacology*, 180, S289–S373. <https://doi.org/10.1111/bph.16181>
- Alexander, S. P. H., Fabbro, D., Kelly, E., Mathie, A. A., Peters, J. A., Veale, E. L., Armstrong, J. F., Faccenda, E., Harding, S. D., Davies, J. A., Beuve, A., Brouckaert, P., Bryant, C., Burnett, J. C., Farndale, R. W., Friebe, A., Garthwaite, J., Hobbs, A. J., Jarvis, G. E., ... Waldman, S. A. (2023). The Concise Guide to PHARMACOLOGY 2023/24: Catalytic receptors. *British Journal of Pharmacology*, 180, S241–S288. <https://doi.org/10.1111/bph.16180>
- Alexander, S. P. H., Kelly, E., Mathie, A. A., Peters, J. A., Veale, E. L., Armstrong, J. F., Buneman, O. P., Faccenda, E., Harding, S. D., Spedding, M., Cidlowski, J. A., Fabbro, D., Davenport, A. P., Striessnig, J., Davies, J. A., Ahlers-Dannen, K. E., Alqinyah, M., Arumugam, T. V., Bodle, C., ... Zolghadri, Y. (2023). The Concise Guide to PHARMACOLOGY 2023/24: Introduction and Other Protein Targets. *British Journal of Pharmacology*, 180, S1–S22. <https://doi.org/10.1111/bph.16176>
- Alexander, S. P. H., Mathie, A. A., Peters, J. A., Veale, E. L., Striessnig, J., Kelly, E., Armstrong, J. F., Faccenda, E., Harding, S. D., Davies, J. A., Aldrich, R. W., Attali, B., Baggetta, A. M., Becirovic, E., Biel, M., Bill, R. M., Caceres, A. I., Catterall, W. A., Conner, A. C., ... Zhu, M. (2023). The Concise Guide to PHARMACOLOGY 2023/24: Ion channels. *British Journal of Pharmacology*, 180, S145–S222. <https://doi.org/10.1111/bph.16178>
- Al-Gayyar, M. M. H., Matragoon, S., Pillai, B. A., Ali, T. K., Abdelsaid, M. A., & El-Remessy, A. B. (2011). Epicatechin blocks pro-nerve growth factor (proNGF)-mediated retinal neurodegeneration via inhibition of p75 neurotrophin receptor expression in a rat model of diabetes [corrected]. *Diabetologia*, 54(3), 669–680. <https://doi.org/10.1007/S00125-010-1994-3>
- Al-Gayyar, M. M. H., Mysona, B. A., Matragoon, S., Abdelsaid, M. A., El-Azab, M. F., Shanab, A. Y., Ha, Y., Smith, S. B., Bollinger, K. E., & El-Remessy, A. B. (2013). Diabetes and overexpression of proNGF cause retinal neurodegeneration via activation of RhoA pathway. *PLoS ONE*, 8(1), e54692. <https://doi.org/10.1371/JOURNAL.PONE.0054692>
- Ali, T. K., Al-Gayyar, M. M. H., Matragoon, S., Pillai, B. A., Abdelsaid, M. A., Nussbaum, J. J., & El-Remessy, A. B. (2011). Diabetes-induced peroxynitrite impairs the balance of pro-nerve growth factor and nerve growth factor, and causes neurovascular injury. *Diabetologia*, 54(3), 657–668. <https://doi.org/10.1007/S00125-010-1935-1>
- Alisky, J. M., & Davidson, B. L. (2000). Gene therapy for amyotrophic lateral sclerosis and other motor neuron diseases. *Human Gene Therapy*, 11(17), 2315–2329. <https://doi.org/10.1089/104303400750038435>
- Allen, S. J., & Dawbarn, D. (2006). Clinical relevance of the neurotrophins and their receptors. *Clinical Science (London, England : 1979)*, 110(2), 175–191. <https://doi.org/10.1042/CS20050161>

- Aloe, L. (1998). Nerve growth factor and autoimmune diseases: Role of tumor necrosis factor- α ? *Advances in Pharmacology (San Diego, Calif.)*, 42(C), 591–594. [https://doi.org/10.1016/S1054-3589\(08\)60820-0](https://doi.org/10.1016/S1054-3589(08)60820-0)
- Aloe, L., Rocco, M., Balzamino, B., & Micera, A. (2015). Nerve growth factor: A focus on neuroscience and therapy. *Current Neuropharmacology*, 13(3), 294–303. <https://doi.org/10.2174/1570159X13666150403231920>
- Aloe, L., Skaper, S. D., Leon, A., & Levi-Montalcini, R. (1994). Nerve growth factor and autoimmune diseases. *Autoimmunity*, 19(2), 141–150. <https://doi.org/10.3109/08916939409009542>
- Al-Shawi, R., Hafner, A., Olson, J., Chun, S., Raza, S., Thrasivoulou, C., Lovestone, S., Killick, R., Simons, P., & Cowen, T. (2008). Neurotoxic and neurotrophic roles of proNGF and the receptor sortilin in the adult and ageing nervous system. *European Journal of Neuroscience*, 27(8), 2103–2114. <https://doi.org/10.1111/j.1460-9568.2008.06152.x>
- Althaus, H. H. (2004). Remyelination in multiple sclerosis: A new role for neurotrophins? *Progress in Brain Research*, 146, 415–432. [https://doi.org/10.1016/S0079-6123\(03\)46026-3](https://doi.org/10.1016/S0079-6123(03)46026-3)
- Anand, P., Parrett, A., Martin, J., Zeman, S., Foley, P., Swash, M., Leigh, P. N., Cedarbaum, J. M., Lindsay, R. M., Williams-Chestnut, R. E., & Sinicropi, D. V. (1995). Regional changes of ciliary neurotrophic factor and nerve growth factor levels in post mortem spinal cord and cerebral cortex from patients with motor disease. *Nature Medicine*, 1(2), 168–172. <https://doi.org/10.1038/NM0295-168>
- Andreasen, N., Hesse, C., Davidsson, P., Minthon, L., Wallin, A., Winblad, B., Vanderstichele, H., Vanmechelen, E., & Blennow, K. (1999). Cerebrospinal fluid beta-amyloid(1–42) in Alzheimer disease: Differences between early- and late-onset Alzheimer disease and stability during the course of disease. *Archives of Neurology*, 56(6), 673–680. <https://doi.org/10.1001/ARCHNEUR.56.6.673>
- Antonetti, D. A., Barber, A. J., Bronson, S. K., Freeman, W. M., Gardner, T. W., Jefferson, L. S., Kester, M., Kimball, S. R., Krady, J. K., LaNoue, K. F., Norbury, C. C., Quinn, P. G., Sandirasegarane, L., & Simpson, I. A. (2006). Diabetic retinopathy: Seeing beyond glucose-induced microvascular disease. *Diabetes*, 55(9), 2401–2411. <https://doi.org/10.2337/DB05-1635>
- Antonetti, D. A., Klein, R., & Gardner, T. W. (2012). Diabetic retinopathy. *The New England Journal of Medicine*, 366(13), 1227–1239. <https://doi.org/10.1056/NEJMRA1005073>
- Antonijevic, M., Charou, D., Davis, A., Cudel, T., Valcarcel, M., Ramos, I., Villacé, P., Claeys, S., Dallemagne, P., Gravanis, A., Charalampopoulos, I., & Rochais, C. (2024). Development of pleiotropic TrkB and 5-HT4 receptor ligands as neuroprotective agents. *Molecules*, 29(2), 515. <https://doi.org/10.3390/molecules29020515>
- Aoi, M., Date, I., Tomita, S., & Ohmoto, T. (2000). The effect of intrastriatal single injection of GDNF on the nigrostriatal dopaminergic system in hemiparkinsonian rats: Behavioral and histological studies using two different dosages. *Neuroscience Research*, 36(4), 319–325. [https://doi.org/10.1016/S0168-0102\(00\)00097-3](https://doi.org/10.1016/S0168-0102(00)00097-3)
- Apfel, S. C., Schwartz, S., Adornato, B. T., Freeman, R., Biton, V., Rendell, M., Vinik, A., Giuliani, M., Stevens, J. C., Barbano, R., & Dyck, P. J. (2000). Efficacy and safety of recombinant human nerve growth factor in patients with diabetic polyneuropathy: A randomized controlled trial. *JAMA*, 284(17), 2215–2221. <https://doi.org/10.1001/JAMA.284.17.2215>
- Arakawa, Y., Sendtner, M., & Thoenen, H. (1990). Survival effect of ciliary neurotrophic factor (CNTF) on chick embryonic motoneurons in culture: Comparison with other neurotrophic factors and cytokines. *The Journal of Neuroscience: the Official Journal of the Society for Neuroscience*, 10(11), 3507–3515. <https://doi.org/10.1523/JNEUROSCI.10-11-03507.1990>
- Arancibia, S., Silhol, M., Moulière, F., Meffre, J., Höllinger, I., Maurice, T., & Tapia-Arancibia, L. (2008). Protective effect of BDNF against beta-amyloid induced neurotoxicity in vitro and in vivo in rats. *Neurobiology of Disease*, 31(3), 316–326. <https://doi.org/10.1016/J.NBD.2008.05.012>
- Arenas, E., & Persson, H. (1994). Neurotrophin prevents the death of adult central noradrenergic neurons in vivo. *Nature*, 367, 368–371.
- Azman, K. F., & Zakaria, R. (2022). Recent advances on the role of brain-derived neurotrophic factor (BDNF) in neurodegenerative diseases. *International Journal of Molecular Sciences*, 23(12), 6827. <https://doi.org/10.3390/IJMS23126827>
- Baloh, R. H., Johnson, J. P., Avalos, P., Allred, P., Svendsen, S., Gowing, G., Roxas, K., Wu, A., Donahue, B., Osborne, S., Lawless, G., Shelley, B., Wheeler, K., Prina, C., Fine, D., Kendra-Romito, T., Stokes, H., Manoukian, V., Muthukumaran, A., ... Svendsen, C. N. (2022). Transplantation of human neural progenitor cells secreting GDNF into the spinal cord of patients with ALS: A phase 1/2a trial. *Nature Medicine*, 28(9), 1813–1822. <https://doi.org/10.1038/s41591-022-01956-3>
- Barker, P. A. (2004). p75NTR is positively promiscuous: Novel partners and new insights. *Neuron*, 42(4), 529–533. <https://doi.org/10.1016/j.neuron.2004.04.001>
- Baulieu, E. E. (1998). Neurosteroids: A novel function of the brain. *Psychoneuroendocrinology*, 23(8), 963–987. [https://doi.org/10.1016/S0306-4530\(98\)00071-7](https://doi.org/10.1016/S0306-4530(98)00071-7)
- Baulieu, E. E., & Robel, P. (1990). Neurosteroids: A new brain function? *The Journal of Steroid Biochemistry and Molecular Biology*, 37(3), 395–403. [https://doi.org/10.1016/0960-0760\(90\)90490-C](https://doi.org/10.1016/0960-0760(90)90490-C)
- Bay, Y., Dergham, P., Nede, H., Jing, X. J., Galan, A., Rivera, J. C., ZhiHua, S., Mehta, H., Woo, S., Sarunic, M., Neet, K., & Saragovi, U. (2010). Chronic and acute models of retinal neurodegeneration TrkA activity are neuroprotective whereas p75NTR activity is neurotoxic through a paracrine mechanism. *J Biol Chem*, 285, 39392–39400. <https://doi.org/10.1074/jbc.M110.147801>
- Beck, M., Flachenecker, P., Magnus, T., Giess, R., Reiners, K., Toyka, K. V., & Naumann, M. (2005). Autonomic dysfunction in ALS: A preliminary study on the effects of intrathecal BDNF. *Amyotrophic Lateral Sclerosis*, 6(2), 100–103. <https://doi.org/10.1080/14660820510028412>
- Bélanger, N., Grégoire, L., Bédard, P. J., & Di Paolo, T. (2006). DHEA improves symptomatic treatment of moderately and severely impaired MPTP monkeys. *Neurobiology of Aging*, 27(11), 1684–1693. <https://doi.org/10.1016/J.NEUROBIOLAGING.2005.09.028>
- Bemelmans, A. P., Husson, I., Jaquet, M., Mallet, J., Kosofsky, B. E., & Gressens, P. (2006). Lentiviral-mediated gene transfer of brain-derived neurotrophic factor is neuroprotective in a mouse model of neonatal excitotoxic challenge. *Journal of Neuroscience Research*, 83(1), 50–60. <https://doi.org/10.1002/JNR.20704>
- Bennett, J. L., & Stüve, O. (2009). Update on inflammation, neurodegeneration, and immunoregulation in multiple sclerosis: Therapeutic implications. *Clinical Neuropharmacology*, 32(3), 121–132. <https://doi.org/10.1097/WNF.0B013E3181880359>
- Bennett, J. P., Jr., O'Brien, L. C., & Brohawn, D. G. (2016). Pharmacological properties of microneurotrophin drugs developed for treatment of amyotrophic lateral sclerosis. *Biochemical Pharmacology*, 117, 68–77. <https://doi.org/10.1016/j.bcp.2016.08.001>
- Bibel, M., Hoppe, E., & Barde, Y. A. (1999). Biochemical and functional interactions between the neurotrophin receptors trk and p75NTR. *EMBO J*, 18(3), 616–622. <https://doi.org/10.1093/emboj/18.3.616>
- Biernacki, K., Antel, J. P., Blain, M., Narayanan, S., Arnold, D. L., & Prat, A. (2005). Interferon beta promotes nerve growth factor secretion early in the course of multiple sclerosis. *Archives of Neurology*, 62(4), 563–568. <https://doi.org/10.1001/archneur.62.4.563>
- Bilak, M. M., Corse, A. M., & Kuncel, R. W. (2001). Additivity and potentiation of IGF-I and GDNF in the complete rescue of postnatal motor neurons. *Amyotrophic Lateral Sclerosis and Other Motor Neuron Disorders*, 2(2), 83–91. <https://doi.org/10.1080/146608201316949523>

- Bitanirhirwe, B. K., & Woo, T. U. (2011). Oxidative stress in schizophrenia: An integrated approach. *Neuroscience and Biobehavioral Reviews*, 35(3), 878–893. <https://doi.org/10.1016/j.neubiorev.2010.10.008> 20974172
- Björklund, A., Kirik, D., Rosenblad, C., Georgievska, B., Lundberg, C., & Mandel, R. J. (2000). Towards a neuroprotective gene therapy for Parkinson's disease: Use of adenovirus, AAV and lentivirus vectors for gene transfer of GDNF to the nigrostriatal system in the rat Parkinson model. *Brain Research*, 886(1–2), 82–98. [https://doi.org/10.1016/S0006-8993\(00\)02915-2](https://doi.org/10.1016/S0006-8993(00)02915-2)
- Blurton-Jones, M., Kitazawa, M., Martinez-Coria, H., Castello, N. A., Müller, F. J., Loring, J. F., Yamasaki, T. R., Poon, W. W., Green, K. N., & LaFerla, F. M. (2009). Neural stem cells improve cognition via BDNF in a transgenic model of Alzheimer disease. *Proceedings of the National Academy of Sciences of the United States of America*, 106(32), 13594–13599. <https://doi.org/10.1073/PNAS.0901402106>
- Bonetto, G., Charalampopoulos, I., Gravanis, A., & Karageorgos, D. (2017). The novel synthetic microneurotrophin BNN27 protects mature oligodendrocytes against cuprizone-induced death, through the NGF receptor TrkA. *Glia*, 65(8), 1376–1394. <https://doi.org/10.1002/glia.23170>
- Boots, E. A., Schultz, S. A., Clark, L. R., Racine, A. M., Darst, B. F., Koscik, R. L., Carlsson, C. M., Gallagher, C. L., Hogan, K. J., Bendlin, B. B., Asthana, S., Sager, M. A., Hermann, B. P., Christian, B. T., Dubal, D. B., Engelman, C. D., Johnson, S. C., & Okonkwo, O. C. (2017). BDNF Val66Met predicts cognitive decline in the Wisconsin registry for Alzheimer's prevention. *Neurology*, 88(22), 2098–2106. <https://doi.org/10.1212/WNL.0000000000003980>
- Bosse, K. E., Maina, F. K., Birbeck, J. A., France, M. M., Roberts, J. J. P., Colombo, M. L., & Mathews, T. A. (2012). Aberrant striatal dopamine transmitter dynamics in brain-derived neurotrophic factor-deficient mice. *Journal of Neurochemistry*, 120(3), 385–395. <https://doi.org/10.1111/J.1471-4159.2011.07531.X>
- Botsakis, K., Mourtzi, T., Panagiotakopoulou, V., Vreka, M., Stathopoulos, G. T., Padiatitakis, I., Charalampopoulos, I., Gravanis, A., Delis, F., Antoniou, K., Zisimopoulos, D., Georgiou, C. D., Panagopoulos, N. T., Matsakis, N., & Angelatou, F. (2017). BNN-20, a synthetic microneurotrophin, strongly protects dopaminergic neurons in the “weaver” mouse, a genetic model of dopamine-denervation, acting through the TrkB neurotrophin receptor. *Neuropharmacology*, 121, 140–157. <https://doi.org/10.1016/j.neuropharm.2017.04.043>
- Brahimi, F., Galan, A., Siegel, S., Szobota, S., Sarunic, M. V., Foster, A. C., & Saragovi, H. U. (2021). Therapeutic neuroprotection by an engineered neurotrophin that selectively activates tropomyosin receptor kinase (Trk) family neurotrophin receptors but not the p75 neurotrophin receptor. *Molecular Pharmacology*, 100(5), 491–501. <https://doi.org/10.1124/molpharm.121.000301> 34470776
- Brahimi, F., Nassour, H., Galan, A., Guruswamy, R., Ortiz, C., Nejat, A., Nedev, H., Trempe, J. F., & Saragovi, H. U. (2025). Selective inhibitors of the TrkC.T1 receptor reduce retinal inflammation and delay neuronal death in a model of retinitis pigmentosa. *PNAS Nexus*, 4(2), pgaf020. <https://doi.org/10.1093/pnasnexus/pgaf020> 39911316
- Bruno, F., Abondio, P., Montesanto, A., Luiselli, D., Bruni, A. C., & Maletta, R. (2023). The nerve growth factor receptor (NGFR/p75NTR): A major player in Alzheimer's disease. *International Journal of Molecular Sciences*, 24(4), 3200. <https://doi.org/10.3390/ijms24043200> 36834612
- Bruno, M. A., Clarke, P. B. S., Seltzer, A., Quirion, R., Burgess, K., Cuello, A. C., & Saragovi, H. U. (2004). Long-lasting Rescue of age-associated deficits in cognition and the CNS cholinergic phenotype by a partial agonist peptidomimetic ligand of TrkA. *The Journal of Neuroscience*, 24(37), 8009–8018. <https://doi.org/10.1523/JNEUROSCI.1508-04.2004>
- Buchman, A. S., Yu, L., Boyle, P. A., Schneider, J. A., De Jager, P. L., & Bennett, D. A. (2016). Higher brain BDNF gene expression is associated with slower cognitive decline in older adults. *Neurology*, 86(8), 735–741. <https://doi.org/10.1212/WNL.0000000000002387>
- Budni, J., Bellettini-Santos, T., Mina, F., Garcez, M. L., & Zugno, A. I. (2015). The involvement of BDNF, NGF and GDNF in aging and Alzheimer's disease. *Aging and Disease*, 6(5), 331–341. <https://doi.org/10.14336/AD.2015.0825>
- Buj-Bello, A., Buchman, V. L., Horton, A., Rosenthal, A., & Davies, A. M. (1995). GDNF is an age-specific survival factor for sensory and autonomic neurons. *Neuron*, 15(4), 821–828. [https://doi.org/10.1016/0896-6273\(95\)90173-6](https://doi.org/10.1016/0896-6273(95)90173-6)
- Cai, H. L., Cao, T., Zhou, X., & Yao, J. K. (2018). Neurosteroids in schizophrenia: Pathogenic and therapeutic implications. *Frontiers in Psychiatry*, 9, 73. <https://doi.org/10.3389/FPSYT.2018.00073>
- Calogeropoulou, T., Avlonitis, N., Minas, V., Alexi, X., Pantzou, A., Charalampopoulos, I., Zervou, M., Vergou, V., Katsanou, E. S., Lazaridis, I., Alexis, M. N., & Gravanis, A. (2009). Novel Dehydroepiandrosterone derivatives with antiapoptotic, neuroprotective activity. *Journal of Medicinal Chemistry*, 52(21), 6569–6587. <https://doi.org/10.1021/jm900468p>
- Cao, M. C., Cawston, E. E., Chen, G., Brooks, C., Douwes, J., McLean, D., Graham, E. S., Dragunow, M., & Scotter, E. L. (2022). Serum biomarkers of neuroinflammation and BBB leakage in amyotrophic lateral sclerosis. *BMC Neurology*, 22(1), 216. <https://doi.org/10.1186/S12883-022-02730-1>
- Cattaneo, A., & Calissano, P. (2012). Nerve growth factor and Alzheimer's disease: New facts for an old hypothesis. *Molecular Neurobiology*, 46(3), 588–604. <https://doi.org/10.1007/S12035-012-8310-9>
- Chan, J. R., Watkins, T. A., Cosgaya, J. M., Zhang, C., Chen, L., Reichardt, L. F., Shooter, E. M., & Barres, B. A. (2004). NGF controls axonal receptivity to myelination by Schwann cells or oligodendrocytes. *Neuron*, 43(2), 183–191. <https://doi.org/10.1016/j.neuron.2004.06.024>
- Chao, M. Y., Rajagopal, R., & Lee, F. S. (2006). Neurotrophin signalling in health and disease. *Clinical Science (London, England : 1979)*, 110(2), 167–173. <https://doi.org/10.1042/CS20050163>
- Charalampopoulos, I., Alexaki, V. I., Tsatsanis, C., Minas, V., Dermizaki, E., Lazaridis, I., Vardoulis, L., Stournaras, C., Margioris, A. N., Castanas, E., & Gravanis, A. (2006). Neurosteroids as endogenous inhibitors of neuronal cell apoptosis in aging. *Annals of the New York Academy of Sciences*, 1088, 139–152. <https://doi.org/10.1196/ANNALS.1366.003>
- Charalampopoulos, I., Margioris, A. N., & Gravanis, A. (2008). Neurosteroid dehydroepiandrosterone exerts anti-apoptotic effects by membrane-mediated, integrated genomic and non-genomic pro-survival signaling pathways. *Journal of Neurochemistry*, 107(5), 1457–1469. <https://doi.org/10.1111/j.1471-4159.2008.05732.x>
- Charalampopoulos, I., Remboutsika, E., Margioris, A. N., & Gravanis, A. (2008). Neurosteroids as modulators of neurogenesis and neuronal survival. *Trends in Endocrinology and Metabolism*, 19(8), 300–307. <https://doi.org/10.1016/j.tem.2008.07.004>
- Charalampopoulos, I., Tsatsanis, C., Dermizaki, E., Alexaki, V. I., Castanas, E., Margioris, A. N., & Gravanis, A. (2004). Dehydroepiandrosterone and allopregnanolone protect sympathoadrenal medulla cells against apoptosis via antiapoptotic Bcl-2 proteins. *Proceedings of the National Academy of Sciences of the United States of America*, 101(21), 8209–8214. <https://doi.org/10.1073/PNAS.0306631101>
- Charou, D., Rogdakis, T., Latorrata, A., Valcarcel, M., Papadogiannis, V., Athanasiou, C., Tsengenes, A., Papadopoulou, M. A., Lypitkas, D., Lavigne, M. D., Katsila, T., Wade, R. C., Cader, M. Z., Calogeropoulou, T., Gravanis, A., & Charalampopoulos, I. (2024). Comprehensive characterization of the neurogenic and neuroprotective action of a novel TrkB agonist using mouse and human stem cell models of Alzheimer's disease. *Stem Cell Research & Therapy*, 15(1), 1–20. <https://doi.org/10.1186/s13287-024-03818-w>
- Chen, C., Guderyon, M. J., Li, Y., Ge, G., Bhattacharjee, A., Ballard, C., He, Z., Masliah, E., Clark, R. A., O'Connor, J. C., & Li, S. (2019). Non-toxic HSC transplantation-based macrophage/microglia-mediated GDNF delivery for Parkinson's disease. *Molecular Therapy: Methods &*

- Clinical Development*, 17, 83–98. <https://doi.org/10.1016/J.OMTM.2019.11.013>
- Chen, S. D., Wu, C. L., Hwang, W. C., & Yang, D. I. (2017). More insight into BDNF against neurodegeneration: Anti-apoptosis, anti-oxidation, and suppression of autophagy. *International Journal of Molecular Sciences*, 18(3), 545. <https://doi.org/10.3390/IJMS18030545>
- Chmielarz, P., & Saarna, M. (2020). Neurotrophic factors for disease-modifying treatments of Parkinson's disease: Gaps between basic science and clinical studies. *Pharmacological Reports*, 72(5), 1195–1217. <https://doi.org/10.1007/S43440-020-00120-3>
- Cohen, R. I., Marmur, R., Norton, W. T., Mehler, M. F., & Kessler, J. A. (1996). Nerve growth factor and neurotrophin-3 differentially regulate the proliferation and survival of developing rat brain oligodendrocytes. *The Journal of Neuroscience: the Official Journal of the Society for Neuroscience*, 16(20), 6433–6442. <https://doi.org/10.1523/JNEUROSCI.16-20-06433.1996>
- Colafrancesco, V., Coassin, M., Rossi, S., & Aloe, L. (2011). Effect of eye NGF administration on two animal models of retinal ganglion cells degeneration. *Annali Dell'istituto Superiore Di Sanita*, 47(3), 284–289. https://doi.org/10.4415/ANN_11_03_08
- Colucci-D'Amato, L., Speranza, L., & Volpicelli, F. (2020). Neurotrophic factor BDNF, physiological functions and therapeutic potential in depression, neurodegeneration and brain cancer. *International Journal of Molecular Sciences*, 21(20), 7777. <https://doi.org/10.3390/ijms21207777>
- Compagnone, N. A., & Mellon, S. H. (2000). Neurosteroids: Biosynthesis and function of these novel neuromodulators. *Frontiers in Neuroendocrinology*, 21(1), 1–56. <https://doi.org/10.1006/FRNE.1999.0188>
- Connor, B., Young, D., Yan, Q., Faull, R. L. M., Synek, B., & Dragunow, M. (1997). Brain-derived neurotrophic factor is reduced in Alzheimer's disease. *Molecular Brain Research*, 49(1–2), 71–81. [https://doi.org/10.1016/S0169-328X\(97\)00125-3](https://doi.org/10.1016/S0169-328X(97)00125-3)
- Copray, S., Küst, B., Emmer, B., Lin, M., Liem, R., Amor, S., de Vries, H., S. F., & Boddeke, E. (2004). Deficient p75 low-affinity neurotrophin receptor expression exacerbates experimental allergic encephalomyelitis in C57/BL6 mice. *Journal of Neuroimmunology*, 148(1–2), 41–53. <https://doi.org/10.1016/j.jneuroim.2003.11.008>
- Corse, A. M., Bilak, M. M., Bilak, S. R., Lehar, M., Rothstein, J. D., & Kuncel, R. W. (1999). Preclinical testing of neuroprotective neurotrophic factors in a model of chronic motor neuron degeneration. *Neurobiology of Disease*, 6(5), 335–346. <https://doi.org/10.1006/NBDI.1999.0253>
- Counts, S. E., Nadeem, M., Wu, J., Ginsberg, S. D., Saragovi, H. U., & Mufson, E. J. (2004). Reduction of cortical TrkA but not p75(NTR) protein in early-stage Alzheimer's disease. *Annals of Neurology*, 56(4), 520–531. <https://doi.org/10.1002/ANA.20233>
- Coyle, J., Price, D., & DeLong, M. (1983). Alzheimer's disease: A disorder of cortical cholinergic innervation. *Science*, 219(4589), 1184–1190. <https://doi.org/10.1126/science.6338589>
- Criscuolo, C., Fabiani, C., Bonadonna, C., Origlia, N., & Domenici, L. (2015). BDNF prevents amyloid-dependent impairment of LTP in the entorhinal cortex by attenuating p38 MAPK phosphorylation. *Neurobiology of Aging*, 36(3), 1303–1309. <https://doi.org/10.1016/j.neurobiolaging.2014.11.016>
- Crowley, C., Spencer, S. D., Nishimura, M. C., Chen, K. S., Pitts-Meek, S., Armanini, M. P., Ling, L. H., McMahon, S. B., Shelton, D. L., Levinson, A. D., & Phillips, H. S. (1994). Mice lacking nerve growth factor display perinatal loss of sensory and sympathetic neurons yet develop basal forebrain cholinergic neurons. *Cell*, 76(6), 1001–1011. [https://doi.org/10.1016/0092-8674\(94\)90378-6](https://doi.org/10.1016/0092-8674(94)90378-6)
- Da Penha Berzaghi, M., Cooper, J., Castr, E., Zafra, F., Sofroniew, M., Thoenen, H., & Lindholm, D. (1993). Cholinergic regulation of brain-derived neurotrophic factor (BDNF) and nerve growth factor (NGF) but not Neurotrophin-3 (NT-3) mRNA levels in the developing rat hippocampus. *The Journal of Neuroscience*, 13(g), 38183826.
- D'Amico, E., Patti, F., Zanghi, A., & Zappia, M. (2016). A personalized approach in progressive multiple sclerosis: The current status of disease modifying therapies (DMTs) and future perspectives. *International Journal of Molecular Sciences*, 17(10), 1725. <https://doi.org/10.3390/IJMS17101725>
- DaRocha-Souto, B., Coma, M., Pérez-Nievas, B. G., Scotton, T. C., Siao, M., Sánchez-Ferrer, P., Hashimoto, T., Fan, Z., Hudry, E., Barroeta, I., Serenó, L., Rodríguez, M., Sánchez, M. B., Hyman, B. T., & Gómez-Isla, T. (2012). Activation of glycogen synthase kinase-3 beta mediates β -amyloid induced neuritic damage in Alzheimer's disease. *Neurobiology of Disease*, 45(1), 425–437. <https://doi.org/10.1016/j.nbd.2011.09.002>
- De la Rosa, A., Solana, E., Corpas, R., Bartrés-Faz, D., Pallàs, M., Vina, J., Sanfeliu, C., & Gomez-Cabrera, M. C. (2019). Long-term exercise training improves memory in middle-aged men and modulates peripheral levels of BDNF and Cathepsin B. *Scientific Reports*, 9(1), 1–11. <https://doi.org/10.1038/s41598-019-40040-8>
- Demuth, H., Hosseini, S., Düsedau, H. P., Dunay, I. R., Korte, M., & Zagrebelsky, M. (2023). Deletion of p75NTR rescues the synaptic but not the inflammatory status in the brain of a mouse model for Alzheimer's disease. *Frontiers in Molecular Neuroscience*, 16, 1163087. <https://doi.org/10.3389/fnmol.2023.1163087>
- Dobson, R., & Giovannoni, G. (2019). Multiple sclerosis—A review. *European Journal of Neurology*, 26(1), 27–40. <https://doi.org/10.1111/ENE.13819>
- Durany, N., Michel, T., Kurt, J., Cruz-Sánchez, F. F., Cervós-Navarro, J., & Riederer, P. (2000). Brain-derived neurotrophic factor and neurotrophin-3 levels in Alzheimer's disease brains. *International Journal of Developmental Neuroscience*, 18(8), 807–813. [https://doi.org/10.1016/S0736-5748\(00\)00046-0](https://doi.org/10.1016/S0736-5748(00)00046-0)
- Ebrahimi, Z., Talaei, S., Aghamiri, S., Goradel, N. H., Jafarpour, A., & Negahdari, B. (2020). Overcoming the BBB in neurodegenerative disorders and brain tumours. *IET Nanobiotechnology*, 14(6), 441–448. <https://doi.org/10.1049/IET-NBT.2019.0351>
- Enterria-Morales, D., López-López, I., López-Barneo, J., & d'Anglemont de Tassigny, X. (2020). Role of glial cell line-derived neurotrophic factor in the maintenance of adult mesencephalic catecholaminergic neurons. *Movement Disorders: Official Journal of the Movement Disorder Society*, 35(4), 565–576. <https://doi.org/10.1002/MDS.27986>
- Faria, M. C., Gonçalves, G. S., Rocha, N. P., Moraes, E. N., Bicalho, M. A., Gualberto Cintra, M. T., Jardim de Paula, J., Ravic, J., de Miranda, L. F., de Souza, C., Ferreira, A., Teixeira, A. O. L., Gomes, K. B., Carvalho, M., das, G., & Sousa, L. P. (2014). Increased plasma levels of BDNF and inflammatory markers in Alzheimer's disease. *Journal of Psychiatric Research*, 53(1), 166–172. <https://doi.org/10.1016/j.jpsychires.2014.01.019>
- Ferreira, D., Westman, E., Eyjolfssdottir, H., Almqvist, P., Lind, G., Linderöth, B., Seiger, Å., Blennow, K., Karami, A., Darreh-Shori, T., Wiberg, M., Simmons, A., Wahlund, L. O., Wahlberg, L., & Eriksdottir, M. (2015). Brain changes in Alzheimer's disease patients with implanted encapsulated cells releasing nerve growth factor. *Journal of Alzheimer's Disease: JAD*, 43(3), 1059–1072. <https://doi.org/10.3233/JAD-141068>
- Francis, P. T., Palmer, A. M., Snape, M., & Wilcock, G. K. (1999). The cholinergic hypothesis of Alzheimer's disease: A review of progress. *Journal of Neurology, Neurosurgery, and Psychiatry*, 66(2), 137–147. <https://doi.org/10.1136/JNPN.66.2.137>
- Frozza, R. L., Lourenco, M. V., & de Felice, F. G. (2018). Challenges for Alzheimer's disease therapy: Insights from novel mechanisms beyond memory defects. *Frontiers in Neuroscience*, 12(FEB), 37. <https://doi.org/10.3389/FNINS.2018.00037>
- Fulmer, C. G., VonDran, M. W., Stillman, A. A., Huang, Y., Hempstead, B. L., & Dreyfus, C. F. (2014). Astrocyte-derived BDNF supports myelin protein synthesis after cuprizone-induced

- demyelination. *Journal of Neuroscience*, 34(24), 8186–8196. <https://doi.org/10.1523/JNEUROSCI.4267-13.2014>
- Fumagalli, F., Racagni, G., & Riva, M. A. (2006). Shedding light into the role of BDNF in the pharmacotherapy of Parkinson's disease. *The Pharmacogenomics Journal*, 6(2), 95–104. <https://doi.org/10.1038/SJ.TPJ.6500360>
- Galan, A., Barcelona, P., Nedev, H., Darinic, M., Jian, Y., & Saragovi, U. (2017). Subconjunctival delivery of p75NTRAntagonists reduces the inflammatory, vascular, and neurodegenerative pathologies of diabetic retinopathy. *Investigative Ophthalmology & Visual Science*, 58, 2852–2862. <https://doi.org/10.1167/iov.16-20988>
- Gao, L., Zhang, Y., Sterling, K., & Song, W. (2022). Brain-derived neurotrophic factor in Alzheimer's disease and its pharmaceutical potential. *Translational Neurodegeneration*, 11(1), 4. <https://doi.org/10.1186/S40035-022-00279-0>
- Ge, Y. W., & Lahiri, D. K. (2002 Nov). Regulation of promoter activity of the APP gene by cytokines and growth factors: Implications in Alzheimer's disease. *Annals of the New York Academy of Sciences*, 973, 463–467. <https://doi.org/10.1111/j.1749-6632.2002.tb04684.x> PMID: 12485912
- Georgelou, K., Saridaki, E. A., Karali, K., Papagiannaki, A., Charalampopoulos, I., Gravanis, A., & Tzeranis, D. S. (2023). Micro-neurotrophin BDNF reduces astrogliosis and increases density of neurons and implanted neural stem cell-derived cells after spinal cord injury. *Biomedicines*, 11(4), 1170. <https://doi.org/10.3390/biomedicines11041170>
- Giess, R., Holtmann, B., Braga, M., Grimm, T., Müller-Myhsok, B., Toyka, K. V., & Sendtner, M. (2002). Early onset of severe familial amyotrophic lateral sclerosis with a SOD-1 mutation: Potential impact of CNTF as a candidate modifier gene. *American Journal of Human Genetics*, 70(5), 1277–1286. <https://doi.org/10.1086/340427>
- Giménez Y Ribotta, M., Revah, F., Pradier, L., Loquet, I., Mallet, J., & Privat, A. (1997). Prevention of motoneuron death by adenovirus-mediated neurotrophic factors. *Journal of Neuroscience Research*, 48(3), 281–285. [https://doi.org/10.1002/\(SICI\)1097-4547\(19970501\)48:3<281::AID-JNR11>3.3.CO;2-I](https://doi.org/10.1002/(SICI)1097-4547(19970501)48:3<281::AID-JNR11>3.3.CO;2-I)
- Ginsberg, S. D., Che, S., Wu, J., Counts, S. E., & Mufson, E. J. (2006). Down regulation of Trk but not p75NTR gene expression in single cholinergic basal forebrain neurons mark the progression of Alzheimer's disease. *Journal of Neurochemistry*, 97(2), 475–487. <https://doi.org/10.1111/J.1471-4159.2006.03764.X>
- Givalois, L., Naert, G., Rage, F., Ixart, G., Arancibia, S., & Tapia-Arancibia, L. (2004). A single brain-derived neurotrophic factor injection modifies hypothalamo-pituitary-adrenocortical axis activity in adult male rats. *Molecular and Cellular Neuroscience*, 27, 280–295. <https://doi.org/10.1016/j.mcn.2004.07.002>
- Glajch, K. E., Ferraiuolo, L., Mueller, K. A., Stopford, M. J., Prabhkar, V., Gravanis, A., Shaw, P. J., & Sadri-Vakili, G. (2016). MicroNeurotrophins improve survival in motor neuron-astrocyte co-cultures but do not improve disease phenotypes in a mutant SOD1 mouse model of amyotrophic lateral sclerosis. *PLoS One*, 11(10), 1–24. <https://doi.org/10.1371/journal.pone.0164103>
- Gold, R., Linington, C., & Lassmann, H. (2006). Understanding pathogenesis and therapy of multiple sclerosis via animal models: 70 years of merits and culprits in experimental autoimmune encephalomyelitis research. *Brain*, 129(8), 1953–1971. <https://doi.org/10.1093/BRAIN/AWL075>
- Gong, Y., Chang, Z. P., Ren, R. T., Wei, S. H., Zhou, H. F., Chen, X. F., Hou, B. K., Jin, X., & Zhang, M. N. (2012). Protective effects of adeno-associated virus mediated brain-derived neurotrophic factor expression on retinal ganglion cells in diabetic rats. *Cellular and Molecular Neurobiology*, 32(3), 467–475. <https://doi.org/10.1007/S10571-011-9779-X>
- Gravanis, A., Calogeropoulou, T., Panoutsakopoulou, V., Thermos, K., Neophytou, C., & Charalampopoulos, I. (2012). Neurosteroids and microneurotrophins signal through NGF receptors to induce prosurvival signaling in neuronal cells. *Science Signaling*, 5(246), pt8. <https://doi.org/10.1126/SCISIGNAL.2003387>
- Gravel, C., Götz, R., Lorrain, A., & Sendtner, M. (1997). Adenoviral gene transfer of ciliary neurotrophic factor and brain-derived neurotrophic factor leads to long-term survival of axotomized motor neurons. *Nature Medicine*, 3(7), 765–770. <https://doi.org/10.1038/NM0797-765>
- Greene, L. A., & Tischler, A. S. (1976). Establishment of a noradrenergic clonal line of rat adrenal pheochromocytoma cells which respond to nerve growth factor. *Proceedings of the National Academy of Sciences of the United States of America*, 73(7), 2424–2428. <https://doi.org/10.1073/PNAS.73.7.2424>
- Grondin, R., Cass, W. A., Zhang, Z., Stanford, J. A., Gash, D. M., & Gerhardt, G. A. (2003). Glial cell line-derived neurotrophic factor increases stimulus-evoked dopamine release and motor speed in aged rhesus monkeys. *The Journal of Neuroscience*, 23(5), 1974–1980. <https://doi.org/10.1523/JNEUROSCI.23-05-01974.2003>
- Grundström, E., Lindholm, D., Johansson, A., Blennow, K., & Askmark, H. (2000). GDNF but not BDNF is increased in cerebrospinal fluid in amyotrophic lateral sclerosis. *Neuroreport*, 11(8), 1781–1783. <https://doi.org/10.1097/00001756-200006050-00037>
- Guo, L., Davis, B. M., Ravindran, N., Galvao, J., Kapoor, N., Haamedi, N., Shamsheer, E., Luong, V., Fico, E., & Cordeiro, M. F. (2020). Topical recombinant human nerve growth factor (rh-NGF) is neuroprotective to retinal ganglion cells by targeting secondary degeneration. *Scientific Reports*, 10(1), 1–13. <https://doi.org/10.1038/s41598-020-60427-2>
- Hammes, H. P., Federoff, H. J., & Brownlee, M. (1995). Nerve growth factor prevents both neuroretinal programmed cell death and capillary pathology in experimental diabetes. *Molecular Medicine*, 1(5), 527–534. <https://doi.org/10.1007/BF03401589>
- Han, R., Liu, Z., Sun, N., Liu, S., Li, L., Shen, Y., Xiu, J., & Xu, Q. (2021). BDNF alleviates neuroinflammation in the hippocampus of type 1 diabetic mice via blocking the aberrant HMGB1/RAGE/NF-κB pathway. *Aging and Disease*, 10(3), 611–625. <https://doi.org/10.14336/AD.2018.0707>
- Harandi, V. M., Gaied, A. R. N., Brännström, T., Pedrosa Domellöf, F., & Liu, J. X. (2016). Unchanged neurotrophic factors and their receptors correlate with sparing in extraocular muscles in amyotrophic lateral sclerosis. *Investigative Ophthalmology & Visual Science*, 57(15), 6831–6842. <https://doi.org/10.1167/IOVS.16-20074>
- Hefti, F., & Will, B. (1987). Nerve growth factor is a neurotrophic factor for forebrain cholinergic neurons; implications for Alzheimer's disease. *Journal of Neural Transmission. Supplementa*, 24, 309–315. PMID: 2824691
- Hock, C., Heese, K., Hulette, C., Rosenberg, C., & Otten, U. (2000). Region-specific Neurotrophin imbalances in Alzheimer disease: Decreased levels of brain-derived neurotrophic factor and increased levels of nerve growth factor in hippocampus and cortical areas. *Archives of Neurology*, 57(6), 846–851. <https://doi.org/10.1001/ARCHNEUR.57.6.846>
- Holback, S., Adlerz, L., & Iverfeldt, K. (2005). Increased processing of APLP2 and APP with concomitant formation of APP intracellular domains in BDNF and retinoic acid-differentiated human neuroblastoma cells. *Journal of Neurochemistry*, 95, 1059–1068. <https://doi.org/10.1111/j.1471-4159.2005.03440.x>
- Howells, D. W., Porritt, M. J., Wong, J. Y. F., Batchelor, P. E., Kalnins, R., Hughes, A. J., & Donnan, G. A. (2000). Reduced BDNF mRNA expression in the Parkinson's disease substantia nigra. *Experimental Neurology*, 166(1), 127–135. <https://doi.org/10.1006/EXNR.2000.7483>
- Hsiao, Y. H., Hung, H. C., Chen, S. H., & Gean, X. W. (2014). Social interaction rescues memory deficit in an animal model of Alzheimer's disease by increasing BDNF-dependent hippocampal neurogenesis. *The Journal of Neuroscience*, 34(49), 16207–16219.
- Huang, E. J., & Reichardt, L. F. (2001). Neurotrophins: Roles in neuronal development and function. *Annual Review of Neuroscience*, 24, 677–736. <https://doi.org/10.1146/ANNUREV.NEURO.24.1.677>

- Hurtig, H., Joyce, J., Sladek, J. R., & Trojanowski, J. Q. (1989). Postmortem analysis of adrenal-medulla-to-caudate autograft in a patient with Parkinson's disease. *Annals of Neurology*, 25(6), 607–614. <https://doi.org/10.1002/ANA.410250613>
- Ibán-Arias, R., Lisa, S., Mastrodinou, N., Kokona, D., Koulakis, E., Iordanidou, P., Kouvarakis, A., Fothiadaki, M., Papadogkonaki, S., Sotiriou, A., Katerinopoulos, H. E., Gravanis, A., Charalampopoulos, I., & Thermos, K. (2018). The synthetic microneurotrophin BNN27 affects retinal function in rats with streptozotocin-induced diabetes. *Diabetes*, 67(2), 321–333. <https://doi.org/10.2337/db17-0391>
- Ibán-Arias, R., Lisa, S., Poulaki, S., Mastrodinou, N., Charalampopoulos, I., Gravanis, A., & Thermos, K. (2019). Effect of topical administration of the microneurotrophin BNN27 in the diabetic rat retina. *Graefes Arch Clin Exp Ophthalmol*, 257(11), 2429–2436. <https://doi.org/10.1007/s00417-019-04460-6>
- Ibrahim, A. M., Chauhan, L., Bhardwaj, A., Sharma, A., Fayaz, F., Kumar, B., Alhashmi, M., AlHajri, N., Alam, M. S., & Potttoo, F. H. (2022). Brain-derived neurotrophic factor in neurodegenerative disorders. *Biomedicine*, 10(5), 1143. <https://doi.org/10.3390/biomedicine10051143>
- Ikedo, K., Klinkosz, B., Greene, T., Cedarbaum, J. M., Wong, V., Lindsay, R. M., & Mitsumoto, H. (1995). Effects of brain-derived neurotrophic factor on motor dysfunction in wobbler mouse motor neuron disease. *Annals of Neurology*, 37(4), 505–511. <https://doi.org/10.1002/ANA.410370413>
- Ioannou, M. S., & Fahnstok, M. (2017). ProNGF, but not NGF, switches from neurotrophic to apoptotic activity in response to reductions in TrkA receptor levels. *International Journal of Molecular Sciences*, 18(3), 599. <https://doi.org/10.3390/IJMS18030599>
- Iulita, M. F., & Cuello, A. C. (2014). Nerve growth factor metabolic dysfunction in Alzheimer's disease and down syndrome. *Trends in Pharmacological Sciences*, 35(7), 338–348. <https://doi.org/10.1016/J.TIPS.2014.04.010>
- J. Allen, S., J. Watson, J., & Dawbarn, D. (2011). The neurotrophins and their role in Alzheimer's disease. *Current Neuropharmacology*, 9(4), 559–573. <https://doi.org/10.2174/157015911798376190>
- Jansen, P., Giehl, K., Nyengaard, J., Nyengaard, J. R., Teng, K., Liubinski, O., Sjoegaard, S. S., Breiderhoff, T., Gotthardt, M., Lin, F., Eilers, A., Petersen, C. M., Lewin, G. R., Hempstead, B. L., Willnow, T. E., & Nykjaer, A. (2007). Roles for the pro-neurotrophin receptor sortilin in neuronal development, aging and brain injury. *Nature Neuroscience*, 10, 1449–1457. <https://doi.org/10.1038/nn2000>
- Jean, I., Lavalie, C., Barthelais-Pouplard, A., & Fressinaud, C. (2003). Neurotrophin-3 specifically increases mature oligodendrocyte population and enhances remyelination after chemical demyelination of adult rat CNS. *Brain Research*, 972(1–2), 110–118. [https://doi.org/10.1016/S0006-8993\(03\)02510-1](https://doi.org/10.1016/S0006-8993(03)02510-1)
- Ji, Y., Lu, Y., Yang, F., Shen, W., Tang, T. T. T., Feng, L., Duan, S., & Lu, B. (2010). Acute and gradual increases in BDNF concentration elicit distinct signaling and functions in neurons. *Nature Neuroscience*, 13, 302–309. <https://doi.org/10.1038/nn.2505>
- Jiao, S.-S., Shen, L.-L., Zhu, C., Bu, X.-L., Liu, Y.-H., Liu, C.-H., Yao, X.-Q., Zhang, L.-L., Zhou, H.-D., Walker, D. G., Tan, J., Götz, J., Zhou, X.-F., & Wang, Y.-J. (2016). Brain-derived neurotrophic factor protects against tau-related neurodegeneration of Alzheimer's disease. *Translational Psychiatry*, 6(10), e907. <https://doi.org/10.1038/tp.2016.186>
- Joseph-Hernandez, S., Jmaeff, S., Pirvulescu, I., Aboukassim, T., & Saragovi, H. U. (2017). Neurotrophin receptor agonists and antagonists as therapeutic agents: An evolving paradigm. *Neurobiology of Disease*, 97(Pt B), 139–155. <https://doi.org/10.1016/j.nbd.2016.08.004>
- Just-Borràs, L., Cilleros-Mañé, V., Polischuk, A., Balanya-Segura, M., Tomàs, M., García, N., Tomàs, J., & Lanuza, M. A. (2022). TrkB signaling is correlated with muscular fatigue resistance and less vulnerability to neurodegeneration. *Frontiers in Molecular Neuroscience*, 15, 1069940. <https://doi.org/10.3389/FNMOL.2022.1069940>
- Just-Borràs, L., Hurtado, E., Cilleros-Mañé, V., Biondi, O., Charbonnier, F., Tomàs, M., García, N., Tomàs, J., & Lanuza, M. A. (2019). Running and swimming prevent the deregulation of the BDNF/TrkB neurotrophic signalling at the neuromuscular junction in mice with amyotrophic lateral sclerosis. *Cellular and Molecular Life Sciences*, 77(15), 3027–3040. <https://doi.org/10.1007/S00018-019-03337-5>
- Kalafatis, I., Patellis, A., Charalampopoulos, I., Gravanis, A., & Karageorgos, D. (2021). The beneficial role of the synthetic microneurotrophin BNN20 in a focal demyelination model. *Journal of Neuroscience Research*, 99(5), 1474–1495. <https://doi.org/10.1002/jnr.24809>
- Kalra, S., Genge, A., & Arnold, D. L. (2003). A prospective, randomized, placebo-controlled evaluation of corticoneuronal response to intrathecal BDNF therapy in ALS using magnetic resonance spectroscopy: Feasibility and results. *Amyotrophic Lateral Sclerosis and Other Motor Neuron Disorders*, 4(1), 22–26. <https://doi.org/10.1080/14660820310006689>
- Kannavou, M., Karali, K., Katsila, T., Siapi, E., Marazioti, A., Klepetsanis, P., Calogeropoulou, T., Charalampopoulos, I., & Antimisaris, S. G. (2023). Development and comparative in vitro and in vivo study of BNN27 mucoadhesive liposomes and nanoemulsions for nose-to-brain delivery. *Pharmaceutics*, 15(2), 419. <https://doi.org/10.3390/pharmaceutics15020419>
- Karimi, N., Ashourizadeh, H., Akbarzadeh Pasha, B., Haghshomar, M., Jouzdani, T., Shobeiri, P., Teixeira, A. L., & Rezaie, N. (2022). Blood levels of brain-derived neurotrophic factor (BDNF) in people with multiple sclerosis (MS): A systematic review and meta-analysis. *Multiple Sclerosis and Related Disorders*, 65, 103984. <https://doi.org/10.1016/J.MSARD.2022.103984>
- Kavirasan, K., Jithu, M., Arif Mulla, M., Sharma, T., Sivasankar, S., Das, U. N., & Angayarkanni, N. (2015). Low blood and vitreal BDNF, LXA4 and altered Th1/Th2 cytokine balance are potential risk factors for diabetic retinopathy. *Metabolism: Clinical and Experimental*, 64(9), 958–966. <https://doi.org/10.1016/J.METABOL.2015.04.005>
- Kawamoto, Y., Nakamura, S., Aikiguchi, I., & Kimura, J. (1998). Immunohistochemical localization of brain-derived neurotrophic factor in the spinal cords of amyotrophic lateral sclerosis and non-amyotrophic lateral sclerosis patients. *Journal of Neuropathology and Experimental Neurology*, 57(9), 822–830. <https://doi.org/10.1097/00005072-199809000-00003>
- Khandaker, G. M., Cousins, L., Deakin, J., Lennox, B. R., Yolken, R., & Jones, P. B. (2015). Inflammation and immunity in schizophrenia: Implications for pathophysiology and treatment. *The Lancet Psychiatry*, 2(3), 258–270. [https://doi.org/10.1016/S2215-0366\(14\)00122-9](https://doi.org/10.1016/S2215-0366(14)00122-9)
- Kirik, D., Georgievskaya, B., & Björklund, A. (2004). Localized striatal delivery of GDNF as a treatment for Parkinson disease. *Nature Neuroscience*, 7(2), 105–110. <https://doi.org/10.1038/nn1175>
- Kitiyant, N., Kitiyanant, Y., Svendsen, C. N., & Thangnipon, W. (2012). BDNF-, IGF-1- and GDNF-secreting human neural progenitor cells rescue amyloid β -induced toxicity in cultured rat septal neurons. *Neurochem Res*, 37, 143–152. <https://doi.org/10.1007/s11064-011-0592-1>
- Knowles, J. K., Rajadas, J., Nguyen, T. V. V., Yang, T., LeMieux, M. C., Vander Griend, L., Ishikawa, C., Massa, S. M., Wyss-Coray, T., & Longo, F. M. (2009). The p75 neurotrophin receptor promotes amyloid- β (1-42)-induced neuritic dystrophy in vitro and in vivo. *The Journal of Neuroscience*, 29(34), 10627–10637. <https://doi.org/10.1523/JNEUROSCI.0620-09.2009>
- Kokkali, M., Karali, K., Thanou, E., Papadopoulou, M. A., Zota, I., Tsimplis, A., Efsthathopoulos, P., Calogeropoulou, T., Li, K. W., Sidiropoulou, K., Gravanis, A., & Charalampopoulos, I. (2024). Multi-modal beneficial effects of BNN27, a nerve growth factor synthetic mimetic, in the 5xFAD mouse model of Alzheimer's disease. *Molecular*

- Psychiatry*, 30, 2265–2283. <https://doi.org/10.1038/s41380-024-02833-w>
- Kokras, N., Dioli, C., Paravatou, R., Sotiropoulos, M. G., Delis, F., Antoniou, K., Calogeropoulou, T., Charalampopoulos, I., Gravanis, A., & Dalla, C. (2020). Psychoactive properties of BNN27, a novel neurosteroid derivative, in male and female rats. *Psychopharmacology*, 237(8), 2435–2449. <https://doi.org/10.1007/s00213-020-05545-5>
- Koliatsos, V. E., Clatterbuck, R. E., Winslow, J. W., Cayouette, M. H., & Prices, D. L. (1993). Evidence that brain-derived neurotrophic factor is a trophic factor for motor neurons in vivo. *Neuron*, 10(3), 359–367. [https://doi.org/10.1016/0896-6273\(93\)90326-M](https://doi.org/10.1016/0896-6273(93)90326-M)
- Kopeck, B. M., Zhao, L., Rosa-Molinar, E., & Sahaan, T. J. (2020). Non-invasive brain delivery and efficacy of BDNF in APP/PS1 transgenic mice as a model of Alzheimer's disease. *Medical Research Archives*, 8(2), 2043. <https://doi.org/10.18103/mra.v8i2.2043> 32551362
- Kopra, J., Vilenius, C., Grealish, S., Härmä, M. A., Varendi, K., Lindholm, J., Castrén, E., Vöikar, V., Björklund, A., Piepponen, T. P., Saarna, M., & Andressoo, J. O. (2015). GDNF is not required for catecholaminergic neuron survival in vivo. *Nature Neuroscience*, 18(3), 319–322. <https://doi.org/10.1038/NN.3941>
- Kosuge, Y., Sekikawa-Nishida, K., Negi, H., Ishige, K., & Ito, Y. (2009). Characterization of chronic glutamate-mediated motor neuron toxicity in organotypic spinal cord culture prepared from ALS model mice. *Neuroscience Letters*, 454(2), 165–169. <https://doi.org/10.1016/J.NEULET.2009.03.017>
- Kowluru, R. A., & Chan, P. S. (2007). Oxidative stress and diabetic retinopathy. *Experimental Diabetes Research*, 2007, 43603. <https://doi.org/10.1155/2007/43603>
- Kowluru, R. A., Kowluru, A., Mishra, M., & Kumar, B. (2015). Oxidative stress and epigenetic modifications in the pathogenesis of diabetic retinopathy. *Progress in Retinal and Eye Research*, 48, 40–61. <https://doi.org/10.1016/J.PRETEYERES.2015.05.001>
- Kramár, E. A., Chen, L. Y., Lauterborn, J. C., Simmons, D. A., Gall, C. M., & Lynch, G. (2012). BDNF upregulation rescues synaptic plasticity in middle-aged ovariectomized rats. *Neurobiology of Aging*, 33(4), 708–719. <https://doi.org/10.1016/J.NEUROBIOLAGING.2010.06.008>
- Kramer, E. R., Aron, L., Ramakers, G. M. J., Seitz, S., Zhuang, X., Beyer, K., Smidt, M. P., & Klein, R. (2007). Absence of Ret signaling in mice causes progressive and late degeneration of the nigrostriatal system. *PLoS Biology*, 5(3), 616–628. <https://doi.org/10.1371/JOURNAL.PBIO.0050039>
- Kuczewski, N., Porcher, C., & Gaiarsa, J. L. (2010). Activity-dependent dendritic secretion of brain-derived neurotrophic factor modulates synaptic plasticity. *The European Journal of Neuroscience*, 32(8), 1239–1244. <https://doi.org/10.1111/J.1460-9568.2010.07378.X>
- Kumamaru, E., Numakawa, T., Adachi, N., Yagasaki, Y., Izumi, A., Niyaz, M., Kudo, M., & Kunugi, H. (2008). Glucocorticoid prevents brain-derived neurotrophic factor-mediated maturation of synaptic function in developing hippocampal neurons through reduction in the activity of mitogen-activated protein kinase. *Molecular Endocrinology*, 22(3), 546–558. <https://doi.org/10.1210/ME.2007-0264>
- Küst, B., Mantingh-Otter, I., Boddeke, E., & Copray, S. (2006). Deficient p75 low-affinity neurotrophin receptor expression does alter the composition of cellular infiltrate in experimental autoimmune encephalomyelitis in C57BL/6 mice. *Journal of Neuroimmunology*, 174(1–2), 92–100. <https://doi.org/10.1016/j.jneuroim.2006.01.020> 16519950
- Lai, S. W., Chen, J. H., Lin, H. Y., Liu, Y. S., Tsai, C. F., Chang, P. C., Lu, D. Y., & Lin, C. (2018). Regulatory effects of neuroinflammatory responses through brain-derived neurotrophic factor signaling in microglial cells. *Molecular Neurobiology*, 55(9), 7487–7499. <https://doi.org/10.1007/S12035-018-0933-Z>
- Lambiase, A., Aloe, L., Centofanti, M., Parisi, V., Mantelli, F., Colafrancesco, V., Manni, G. L., Bucci, M. G., Bonini, S., & Levi-Montalcini, R. (2009). Experimental and clinical evidence of neuroprotection by nerve growth factor eye drops: Implications for glaucoma. *Proceedings of the National Academy of Sciences of the United States of America*, 106(32), 13469–13474. <https://doi.org/10.1073/PNAS.0906678106>
- Lang, A. E., Gill, S., Patel, N. K., Lozano, A., Nutt, J. G., Penn, R., Brooks, D. J., Hotton, G., Moro, E., Heywood, P., Brodsky, M. A., Burchiel, K., Kelly, P., Dalvi, A., Scott, B., Stacy, M., Turner, D., Wooten, V. G. F., Elias, W. J., ... Traub, M. (2006). Randomized controlled trial of intraputamenal glial cell line-derived neurotrophic factor infusion in Parkinson disease. *Annals of Neurology*, 59(3), 459–466. <https://doi.org/10.1002/ANA.20737>
- Laske, C., Stellos, K., Hoffmann, N., Stransky, E., Straten, G., Eschweiler, G. W., & Leyhe, T. (2011). Higher BDNF serum levels predict slower cognitive decline in Alzheimer's disease patients. *The International Journal of Neuropsychopharmacology*, 14(3), 399–404. <https://doi.org/10.1017/S1461145710001008>
- Lattanzio, F., Carboni, L., Carretta, D., Rimondini, R., Candeletti, S., & Romualdi, P. (2014). Human apolipoprotein E4 modulates the expression of Pin1, Sirtuin 1, and Presenilin 1 in brain regions of targeted replacement apoE mice. *Neuroscience*, 256, 360–369. <https://doi.org/10.1016/j.neuroscience.2013.10.017>
- Laudiero, L. B., Aloe, L., Levi-Montalcini, R., Buttinelli, C., Schilter, D., Gillesen, S., & Otten, U. (1992). Multiple sclerosis patients express increased levels of beta-nerve growth factor in cerebrospinal fluid. *Neuroscience Letters*, 147(1), 9–12. [https://doi.org/10.1016/0304-3940\(92\)90762-V](https://doi.org/10.1016/0304-3940(92)90762-V)
- Lazaridis, I., Charalampopoulos, I., Alexaki, V.-I., Avlonitis, N., Peditakis, I., Efstathopoulos, P., Calogeropoulou, T., Castanas, E., & Gravanis, A. (2011). Neurosteroid dehydroepiandrosterone interacts with nerve growth factor (NGF) receptors, preventing neuronal apoptosis. *PLoS Biology*, 9(4), e1001051. <https://doi.org/10.1371/journal.pbio.1001051>
- Lee, D. A., Gross, L. A., Wittrock, D. A., & Windebank, A. J. (1996). Localization and expression of ciliary neurotrophic factor (CNTF) in postmortem sciatic nerve from patients with motor neuron disease and diabetic neuropathy. *Journal of Neuropathology and Experimental Neurology*, 55(8), 915–923. <https://doi.org/10.1097/00005072-199608000-00007>
- Lesné, S., Gabriel, C., Nelson, D. A., White, E., Mackenzie, E. T., Vivien, D., & Buisson, A. (2005). Akt-dependent expression of NAIP-1 protects neurons against amyloid- β toxicity. *The Journal of Biological Chemistry*, 280(26), 24941–24947. <https://doi.org/10.1074/jbc.M413495200> 15797869
- Li, W., Brakefield, D., Pan, Y., Hunter, D., Myckatyn, T. M., & Parsadanian, A. (2007). Muscle-derived but not centrally derived transgene GDNF is neuroprotective in G93A-SOD1 mouse model of ALS. *Experimental Neurology*, 203(2), 457–471. <https://doi.org/10.1016/J.EXPNEUROL.2006.08.028>
- Lim, Y. Y., Hassenstab, J., Goate, A., Fagan, A. M., Benzinger, T. L. S., Cruchaga, C., McDade, E., Chhatwal, J., Levin, J., Farlow, M. R., Graff-Radford, N. R., Laske, C., Masters, C. L., Salloway, S., Schofield, P., Morris, J. C., Maruff, P., & Bateman, R. J. (2018). Effect of BDNFVal66Met on disease markers in dominantly inherited Alzheimer's disease. *Annals of Neurology*, 84(3), 424–435. <https://doi.org/10.1002/ANA.25299>
- Lindvall, O., & Kokaia, Z. (2010). Stem cells in human neurodegenerative disorders—Time for clinical translation? *The Journal of Clinical Investigation*, 120(1), 29–40. <https://doi.org/10.1172/JCI40543>
- Linker, R. A., Lee, D. H., Demir, S., Wiese, S., Kruse, N., Siglienti, I., Gerhardt, E., Neumann, H., Sendtner, M., Lühder, F., & Gold, R. (2010). Functional role of brain-derived neurotrophic factor in neuroprotective autoimmunity: Therapeutic implications in a model of multiple sclerosis. *Brain: A Journal of Neurology*, 133(Pt 8), 2248–2263. <https://doi.org/10.1093/BRAIN/AWQ179>

- Liu, Y., Lijian, T., Fu, X., Zhao, Y., & Xu, X. (2013). BDNF protects retinal neurons from hyperglycemia through the TrkB/ERK/MAPK pathway. *Molecular Medicine Reports*, 7(6), 1773–1778. <https://doi.org/10.3892/MMR.2013.1433/HTML>
- Longo, F., Longo, F., Massa, S., & Massa, S. (2008). Small molecule modulation of p75 neurotrophin receptor functions. *CNS & Neurological Disorders - Drug Targets*, 7(1), 63–70. <https://doi.org/10.2174/187152708783885093>
- Longo, F. M., Yang, T., Knowles, J. K., Xie, Y., Moore, L. A., & Massa, S. M. (2007). Small molecule neurotrophin receptor ligands: Novel strategies for targeting Alzheimer's disease mechanisms. *Current Alzheimer Research*, 4(5), 503–506. <https://doi.org/10.2174/156720507783018316>
- Lorigados, L., Söderström, S., & Ebendal, T. (1992). Two-site enzyme immunoassay for beta NGF applied to human patient sera. *Journal of Neuroscience Research*, 32(3), 329–339. <https://doi.org/10.1002/JNR.490320305>
- Lorigados Pedre, L., Pavón Fuentes, N., Alvarez González, L., McRae, A., Serrano Sánchez, T., Blanco Lescano, L., & Macías González, R. (2002). Nerve growth factor levels in Parkinson disease and experimental parkinsonian rats. *Brain Research*, 952(1), 122–127. [https://doi.org/10.1016/S0006-8993\(02\)03222-5](https://doi.org/10.1016/S0006-8993(02)03222-5)
- Lu, X., & Hagg, T. (1997). Glial cell line-derived neurotrophic factor prevents death, but not reductions in tyrosine hydroxylase, of injured nigrostriatal neurons in adult rats. *Journal of Comparative Neurology*, 388(3), 484–494. [https://doi.org/10.1002/\(SICI\)1096-9861\(19971124\)388:3<484::AID-CNE10>3.0.CO;2-M](https://doi.org/10.1002/(SICI)1096-9861(19971124)388:3<484::AID-CNE10>3.0.CO;2-M)
- Lübke, J., Idoon, F., Mohasel-Roodi, M., Alipour, F., Hami, J., Ehteshampour, A., Mostafaei, H., & Sadeghi, A. (2021). Neurotrophic factors in Alzheimer's disease: Pathogenesis and therapy. *Acta Neurobiologiae Experimentalis*, 81(4), 314–327. <https://doi.org/10.55782/ANE-2021-028>
- Lucchinetti, C. F., Brück, W., Rodriguez, M., & Lassmann, H. (1996). Distinct patterns of multiple sclerosis pathology indicates heterogeneity on pathogenesis. *Brain Pathology (Zurich, Switzerland)*, 6(3), 259–274. <https://doi.org/10.1111/J.1750-3639.1996.TB00854.X>
- M. Metcalfe, S., Bickerton, S., & Fahmy, T. (2017). Neurodegenerative disease: A perspective on cell-based therapy in the new era of cell-free nano-therapy. *Current Pharmaceutical Design*, 23(5), 776–783. <https://doi.org/10.2174/1381612822666161206141744>
- Maninger, N., Wolkowitz, O. M., Reus, V. I., Epel, E. S., & Mellon, S. H. (2009). Neurobiological and neuropsychiatric effects of dehydroepiandrosterone (DHEA) and DHEA sulfate (DHEAS). *Frontiers in Neuroendocrinology*, 30(1), 65–91. <https://doi.org/10.1016/J.YFRNE.2008.11.002>
- Mantelli, F., Lambiase, A., Colafrancesco, V., Rocco, M. L., Macchi, I., & Aloe, L. (2014). NGF and VEGF effects on retinal ganglion cell fate: New evidence from an animal model of diabetes. *European Journal of Ophthalmology*, 24(2), 247–253. <https://doi.org/10.5301/EJO.5000359>
- Masu, Y., Wolf, E., Holtmann, B., Sendtner, M., Brem, G., & Thoenen, H. (1993). Disruption of the CNTF gene results in motor neuron degeneration. *Nature*, 365(6441), 27–32. <https://doi.org/10.1038/365027A0>
- McFarland, H. F., & Martin, R. (2007). Multiple sclerosis: A complicated picture of autoimmunity. *Nature Immunology*, 8(9), 913–919. <https://doi.org/10.1038/ni1507>
- McTigue, D. M., Horner, P. J., Stokes, B. T., & Gage, F. H. (1998). Neurotrophin-3 and brain-derived neurotrophic factor induce oligodendrocyte proliferation and myelination of regenerating axons in the contused adult rat spinal cord. *The Journal of Neuroscience: the Official Journal of the Society for Neuroscience*, 18(14), 5354–5365. <https://doi.org/10.1523/JNEUROSCI.18-14-05354.1998>
- Meng, C., He, Z., & Xing, D. (2013). Low-level laser therapy rescues dendrite atrophy via upregulating BDNF expression: Implications for Alzheimer's disease. *The Journal of Neuroscience*, 33(33), 13505–13517. <https://doi.org/10.1523/JNEUROSCI.0918-13.2013>
- Michalski, B., & Fahnstock, M. (2003). Pro-brain-derived neurotrophic factor is decreased in parietal cortex in Alzheimer's disease. *Molecular Brain Research*, 111(1–2), 148–154. [https://doi.org/10.1016/S0169-328X\(03\)00003-2](https://doi.org/10.1016/S0169-328X(03)00003-2)
- Miller, K. M., Mercado, N. M., & Sortwell, C. E. (2021). Synucleinopathy-associated pathogenesis in Parkinson's disease and the potential for brain-derived neurotrophic factor. *Npj Parkinson's Disease*, 7(1), 1–9. <https://doi.org/10.1038/s41531-021-00179-6>
- Miller, R. G., Petajan, J. H., Bryan, W. W., Armon, C., Barohn, R. J., Goodpasture, J. C., Hoagland, R. J., Parry, G. J., Ross, M. A., Stromatt, S. C., & the rhCNTF ALS Study Group. (1996). A placebo-controlled trial of recombinant human ciliary neurotrophic factor (rhCNTF) in amyotrophic lateral sclerosis. *Annals of Neurology*, 39, 256–260. <https://doi.org/10.1002/ana.410390215>
- Miranda, M., Morici, J. F., Zanon, M. B., & Bekinschtein, P. (2019). Brain-derived neurotrophic factor: A key molecule for memory in the healthy and the pathological brain. *Frontiers in Cellular Neuroscience*, 13, 363. <https://doi.org/10.3389/FNCEL.2019.00363>
- Mitra, S., Behbahani, H., & Eriksdotter, M. (2019). Innovative therapy for Alzheimer's disease-with focus on biodelivery of NGF. *Frontiers in Neuroscience*, 13(FEB), 434303. <https://doi.org/10.3389/FNINS.2019.00038/BIBTEX>
- Mitroshina, E. V., Yarkov, R. S., Mishchenko, T. A., Krut', V. G., Gavrish, M. S., Epifanova, E. A., Babaev, A. A., & Vedunova, M. V. (2020). Brain-derived neurotrophic factor (BDNF) preserves the functional integrity of neural networks in the β -amyloidopathy model in vitro. *Frontiers in Cell and Developmental Biology*, 8, 582. <https://doi.org/10.3389/FCELL.2020.00582/FULL>
- Mo, L., Yang, Z., Zhang, A., & Li, X. (2010). The repair of the injured adult rat hippocampus with NT-3-chitosan carriers. *Biomaterials*, 31(8), 2184–2192. <https://doi.org/10.1016/J.BIOMATERIALS.2009.11.078>
- Mogi, M., Togari, A., Kondo, T., Mizuno, Y., Komure, O., Kuno, S., Ichinose, H., & Nagatsu, T. (1999). Brain-derived growth factor and nerve growth factor concentrations are decreased in the substantia nigra in Parkinson's disease. *Neuroscience Letters*, 270(1), 45–48. [https://doi.org/10.1016/S0304-3940\(99\)00463-2](https://doi.org/10.1016/S0304-3940(99)00463-2)
- Mohajeri, M. H., Figlewicz, D. A., & Bohn, M. C. (1999). Intramuscular grafts of myoblasts genetically modified to secrete glial cell line-derived neurotrophic factor prevent motoneuron loss and disease progression in a mouse model of familial amyotrophic lateral sclerosis. *Human Gene Therapy*, 10(11), 1853–1866. <https://doi.org/10.1089/10430349950017536>
- Monteggia, L. M., Barrot, M., Powell, C. M., Berton, O., Galanis, V., Gemelli, V., Meuth, S., Nagy, A., Greene, R. W., & Nestler, E. J. (2004). Essential role of brain-derived neurotrophic factor in adult hippocampal function. *Proceedings of the National Academy of Sciences of the United States of America*, 101(29), 10827–10832.
- Moreno-Jiménez, E. P., Flor-García, M., Terreros-Roncal, J., Rábano, A., Cafini, F., Pallas-Bazarra, N., Ávila, J., & Llorens-Martín, M. (2019). Adult hippocampal neurogenesis is abundant in neurologically healthy subjects and drops sharply in patients with Alzheimer's disease. *Nature Medicine*, 25(4), 554–560. <https://doi.org/10.1038/s41591-019-0375-9> 30911133
- Mufson, E. J., He, B., Nadeem, M., Perez, S. E., Counts, S. E., Leurgans, S., Fritz, J., Lah, J., Ginsberg, S. D., Wu, J., & Scheff, S. W. (2012). Hippocampal ProNGF signaling pathways and β -amyloid levels in mild cognitive impairment and Alzheimer disease. *Journal of Neuropathology & Experimental Neurology*, 71(11), 1018–1029. <https://doi.org/10.1097/NEN.0B013E318272CAAB>
- Mutoh, T., Sobue, G., Hamano, T., Kuriyama, M., Hirayama, M., Yamamoto, M., & Mitsuma, T. (2000). Decreased phosphorylation levels of TrkB neurotrophin receptor in the spinal cords from patients with amyotrophic lateral sclerosis. *Neurochemical Research*, 25(2), 239–245. <https://doi.org/10.1023/A:1007575504321/METRICS>

- Mysona, B. A., Matragoon, S., Stephens, M., Mohamed, I. N., Farooq, A., Bartasis, M. L., Fouda, A. Y., Shanab, A. Y., Espinosa-Heidmann, D. G., & El-Remessy, A. B. (2015). Imbalance of the nerve growth factor and its precursor as a potential biomarker for diabetic retinopathy. *BioMed Research International*, 2015, 1–12. <https://doi.org/10.1155/2015/571456>
- Nagahara, A., Merrill, D., Coppola, G., Nagahara, A. H., Merrill, D. A., Tsukada, S., Schroeder, B. E., Shaked, G. M., Wang, L., Blesch, A., Kim, A., Conner, J. M., Rockenstein, E., Chao, M. V., Koo, E. H., Geschwind, D., Masliah, E., Chiba, A. A., & Tuszynski, M. H. (2009). Neuroprotective effects of brain-derived neurotrophic factor in rodent and primate models of Alzheimer's disease. *Nature Medicine*, 15, 331–337. <https://doi.org/10.1038/nm.1912>
- Nagahara, A. H., & Tuszynski, M. H. (2011 Mar). Potential therapeutic uses of BDNF in neurological and psychiatric disorders. *Nat Rev Drug Discov.*, 10(3), 209–219. <https://doi.org/10.1038/nrd3366> PMID: 21358740
- Narducci, D., Charou, D., Rogdakis, T., Zota, I., Bafiti, V., Zervou, M., Katsila, T., Gravanis, A., Prousis, K. C., Charalampopoulos, I., & Calogeropoulou, T. (2023). A quest for the stereo-electronic requirements for selective agonism for the neurotrophin receptors TrkA and TrkB in 17-spirocyclic-dehydroepiandrosterone derivatives. *Frontiers in Molecular Neuroscience*, 16, 1244133. <https://doi.org/10.3389/fnmol.2023.1244133>
- Nigam, S. M., Xu, S., Kritikou, J. S., Marosi, K., Brodin, L., & Mattson, M. P. (2017). Exercise and BDNF reduce $\alpha\beta$ production by enhancing α -secretase processing of APP. *Journal of Neurochemistry*, 142(2), 286–296. <https://doi.org/10.1111/JNC.14034>
- Nishio, T., Sunohara, N., & Furukawa, S. (1998). Neurotrophin switching in spinal motoneurons of amyotrophic lateral sclerosis. *Neuroreport*, 9(7), 1661–1665. <https://doi.org/10.1097/00001756-199805110-00073>
- Nociti, V. (2020). What is the role of brain derived neurotrophic factor in multiple sclerosis neuroinflammation? *Neurosciences*, 7(3), 291–299. <https://doi.org/10.20517/2347-8659.2020.25>
- Numakawa, T., & Odaka, H. (2022). The role of Neurotrophin signaling in age-related cognitive decline and cognitive diseases. *International Journal of Molecular Sciences*, 23(14), 7726. <https://doi.org/10.3390/IJMS23147726>
- Numakawa, T., Yokomaku, D., Richards, M., Hori, H., Adachi, N., & Kunugi, H. (2010). Functional interactions between steroid hormones and neurotrophin BDNF. *World Journal of Biological Chemistry*, 1(5), 133. <https://doi.org/10.4331/WJBC.V1.I5.133>
- Obeso, J. A., Rodríguez-Oroz, M. C., Goetz, C. G., Marin, C., Kordower, J. H., Rodríguez, M., Hirsch, E. C., Farrer, M., Schapira, A. H. V., & Halliday, G. (2010). Missing pieces in the Parkinson's disease puzzle. *Nature Medicine*, 16(6), 653–661. <https://doi.org/10.1038/NM.2165>
- Ola, M. S., Nawaz, M. I., El-Asrar, A. A., Abouammoh, M., & Alhomida, A. S. (2013). Reduced levels of brain derived neurotrophic factor (BDNF) in the serum of diabetic retinopathy patients and in the retina of diabetic rats. *Cellular and Molecular Neurobiology*, 33(3), 359–367. <https://doi.org/10.1007/S10571-012-9901-8>
- Ono, S., Imai, T., Igarashi, A., Shimizu, N., Nakagawa, H., & Hu, J. (1999). Decrease in the ciliary neurotrophic factor of the spinal cord in amyotrophic lateral sclerosis. *European Neurology*, 42(3), 163–168. <https://doi.org/10.1159/00008092>
- Ontaneda, D., Fox, R. J., & Chataway, J. (2015). Clinical trials in progressive multiple sclerosis: Lessons learned and future perspectives. *The Lancet. Neurology*, 14(2), 208–223. [https://doi.org/10.1016/S1474-4422\(14\)70264-9](https://doi.org/10.1016/S1474-4422(14)70264-9)
- Palfi, S., Leventhal, L., Chu, Y., Ma, S. Y., Emborg, M., Bakay, R., Déglon, N., Hantraye, P., Aebischer, P., & Kordower, J. H. (2002). Lentivirally delivered glial cell line-derived neurotrophic factor increases the number of striatal dopaminergic neurons in primate models of nigrostriatal degeneration. *The Journal of Neuroscience*, 22(12), 4942–4954. <https://doi.org/10.1523/JNEUROSCI.22-12-04942.2002>
- Pan, W., Banks, W. L., Fasold, M. B., Bluth, J., & Kastin, A. J. (1998). Transport of brain-derived neurotrophic factor across the blood–brain barrier. *Neuropharmacology*, 37(12), 1553–1561. [https://doi.org/10.1016/S0028-3908\(98\)00141-5](https://doi.org/10.1016/S0028-3908(98)00141-5)
- Panagiotakopoulou, V., Botsakis, K., Delis, F., Mourtzi, T., Tzatzarakis, M. N., Dimopoulou, A., Pouli, N., Antoniou, K., Stathopoulos, G. T., Matsokis, N., Charalampopoulos, I., Gravanis, A., & Angelatou, F. (2020). Anti-neuroinflammatory, protective effects of the synthetic microneurotrophin BNN-20 in the advanced dopaminergic neurodegeneration of “weaver” mice. *Neuropharmacology*, 165, 107919. <https://doi.org/10.1016/j.neuropharm.2019.107919>
- Papadopoulou, M. A., Rogdakis, T., Charou, D., Peteinareli, M., Ntarntani, K., Gravanis, A., Chanoumidou, K., & Charalampopoulos, I. (2023). Neurotrophin analog ENT-A044 activates the p75 Neurotrophin receptor, regulating neuronal survival in a cell context-dependent manner. *International Journal of Molecular Sciences*, 24(14), 11683. <https://doi.org/10.3390/ijms241411683>
- Pardridge, W. M. (2003). BBB drug targeting enables neuroprotection in brain ischemia following delayed intravenous administration of neurotrophins. In C. Alzheimer (Ed.), *Molecular and cellular biology of neuroprotection in the CNS*. Advances in Experimental Medicine and Biology (Vol. 513). Springer. https://doi.org/10.1007/978-1-4615-0123-7_15
- Park, S., Kim, H. T., Yun, S., Kim, I. S., Lee, J., Lee, I. S., & Kook, I. P. (2009). Growth factor-expressing human neural progenitor cell grafts protect motor neurons but do not ameliorate motor performance and survival in ALS mice. *Experimental & Molecular Medicine*, 41(7), 487–500. <https://doi.org/10.3858/emmm.2009.41.7.054>
- Pascual, A., Hidalgo-Figueroa, M., Piruat, J. I., Pintado, C. O., Gómez-Díaz, R., & López-Barneo, J. (2008). Absolute requirement of GDNF for adult catecholaminergic neuron survival. *Nature Neuroscience*, 11(7), 755–761. <https://doi.org/10.1038/nn.2136>
- Patel, N. K., Pavese, N., Javed, S., Hottot, G. R., Brooks, D. J., & Gill, S. S. (2013). Benefits of putaminal GDNF infusion in Parkinson disease are maintained after GDNF cessation. *Neurology*, 81(13), 1176–1178. <https://doi.org/10.1212/WNL.0B013E3182A55EA5>
- Pediaditakis, I., Efsthathopoulos, P., Prousis, K. C., Zervou, M., Arévalo, J. C., Alexaki, V. I., Nikolettou, V., Karagianni, E., Potamitis, C., Tavernarakis, N., Chavakis, T., Margioris, A. N., Venihaki, M., Calogeropoulou, T., Charalampopoulos, I., & Gravanis, A. (2016). Selective and differential interactions of BNN27, a novel C17-spiroepoxy steroid derivative, with TrkA receptors, regulating neuronal survival and differentiation. *Neuropharmacology*, 111, 266–282. <https://doi.org/10.1016/j.neuropharm.2016.09.007>
- Pediaditakis, I., Iliopoulos, I., Theologidis, I., Delivanoglou, N., Margioris, A. N., Charalampopoulos, I., & Gravanis, A. (2015). Dehydroepiandrosterone: An ancestral ligand of neurotrophin receptors. *Endocrinology*, 156(1), 16–23. <https://doi.org/10.1210/en.2014-1596>
- Pediaditakis, I., Kourgiantaki, A., Prousis, K. C., Potamitis, C., Xanthopoulos, K. P., Zervou, M., Calogeropoulou, T., Charalampopoulos, I., & Gravanis, A. (2016). BNN27, a 17-Spiroepoxy steroid derivative, interacts with and activates p75 neurotrophin receptor, rescuing cerebellar granule neurons from apoptosis. *Frontiers in Pharmacology*, 7, 512. <https://doi.org/10.3389/fphar.2016.00512>
- Pencea, V., Bingham, K. D., Wiegand, S. J., & Luskin, M. B. (2001). Infusion of brain-derived neurotrophic factor into the lateral ventricle of the adult rat leads to new neurons in the parenchyma of the striatum, septum, thalamus, and hypothalamus. *The Journal of Neuroscience: The Official Journal of the Society for Neuroscience*, 21(17), 6706–6717. <https://doi.org/10.1523/JNEUROSCI.21-17-06706.2001>
- Peng, S., Wu, J., Mufson, E. J., & Fahnstock, M. (2004). Increased proNGF levels in subjects with mild cognitive impairment and mild

- Alzheimer disease. *Journal of Neuropathology & Experimental Neurology*, 63(6), 641–649. <https://doi.org/10.1093/JNEN/63.6.641>
- Perez, S. E., He, B., Nadeem, M., Wu, J., Scheff, S. W., Abrahamson, E. E., Ikonovic, M. D., & Mufson, E. J. (2014). Resilience of precuneus neurotrophic signaling pathways despite amyloid pathology in prodromal Alzheimer's disease. *Biological Psychiatry*, 77(8), 693. <https://doi.org/10.1016/J.BIOPSYCH.2013.12.016>
- Perini, G., Della-Bianca, V., Politi, V., Della Valle, G., Dal-Pra, I., Rossi, F., & Armato, U. (2002). Role of p75 neurotrophin receptor in the neurotoxicity by beta-amyloid peptides and synergistic effect of inflammatory cytokines. *The Journal of Experimental Medicine*, 195(Apr 1), 907–918. <https://doi.org/10.1084/jem.20011797> 11927634
- Peterson, D. I., Price, M. L., & Small, C. S. (1989). Autopsy findings in a patient who had an adrenal-to-brain transplant for Parkinson's disease. *Neurology*, 39(2 Pt 1), 235–238. <https://doi.org/10.1212/WNL.39.2.235>
- Phillips, H. S., Hains, J. M., Armanini, M., Laramée, G. R., Johnson, S. A., & Winslow, J. W. (1991 Nov). BDNF mRNA is decreased in the hippocampus of individuals with Alzheimer's disease. *Neuron*, 7(5), 695–702. [https://doi.org/10.1016/0896-6273\(91\)90273-3](https://doi.org/10.1016/0896-6273(91)90273-3) PMID: 1742020
- Pitsikas, N., & Gravanis, A. (2017). The novel dehydroepiandrosterone (DHEA) derivative BNN27 counteracts delay-dependent and scopolamine-induced recognition memory deficits in rats. *Neurobiology of Learning and Memory*, 140, 145–153. <https://doi.org/10.1016/j.nlm.2017.03.004>
- Pitsikas, N., Zoupa, E., & Gravanis, A. (2021). The novel dehydroepiandrosterone (DHEA) derivative BNN27 counteracts cognitive deficits induced by the D1/D2 dopaminergic receptor agonist apomorphine in rats. *Psychopharmacology*, 238(1), 227–237. <https://doi.org/10.1007/s00213-020-05672-z>
- Platón-Corchado, M., Barcelona, P. F., Jmaeff, S., Marchena, M., Hernández-Pinto, A. M., Hernández-Sánchez, C., Saragovi, H. U., & de la Rosa, E. J. (2017). P75NTR antagonists attenuate photoreceptor cell loss in murine models of retinitis pigmentosa. *Cell Death & Disease*, 8(7), e2922. <https://doi.org/10.1038/cddis.2017.306> 28703796
- Poduslo, J. F., & Curran, G. L. (1996). Permeability at the blood-brain and blood-nerve barriers of the neurotrophic factors: NGF, CNTF, NT-3. *BDNF, Molecular Brain Research*, 36(2), 280–286. [https://doi.org/10.1016/0169-328X\(95\)00250-V](https://doi.org/10.1016/0169-328X(95)00250-V)
- Poon, W. W., Carlos, A. J., Aguilar, B. L., Berchtold, N. C., Kawano, C. K., Zograbyan, V., Yaoprake, T., Shelanski, M., & Cotman, C. W. (2013). B-amyloid (A β) oligomers impair brain-derived neurotrophic factor retrograde trafficking by down-regulating ubiquitin C-terminal hydrolase, UCH-L1. *The Journal of Biological Chemistry*, 288(23), 16937–16948. <https://doi.org/10.1074/jbc.M113.463711> 23599427
- Poulaki, S., Rasouli, O., Liapakis, G., Gravanis, A., & Venihaki, M. (2021). Analgesic and anti-inflammatory effects of the synthetic neurosteroid analogue BNN27 during CFA-induced hyperalgesia. *Biomedicines*, 9(9), 1185.
- Prince, MA., Wimo, A., Guerchet, M., Gemma-Claire Ali, M., Wu, Y.-T., Prina, M., Yee Chan, K., & Xia, Z. (2015). *World Alzheimer Report 2015 The Global Impact of Dementia An Analysis of prevalence, Incidence, cost And Trends*. www.alz.co.uk/worldreport2015corrections
- Przedborski, S., Vila, M., & Jackson-Lewis, V. (2003). Neurodegeneration: What is it and where are we? *The Journal of Clinical Investigation*, 111(1), 3–10. <https://doi.org/10.1172/JCI17522>
- Qian, L., Milne, M. R., Shephard, S., Rogers, M. L., Medeiros, R., & Coulson, E. J. (2019). Removal of p75 neurotrophin receptor expression from cholinergic basal forebrain neurons reduces amyloid- β plaque deposition and cognitive impairment in aged APP/PS1 mice. *Molecular Neurobiology*, 56(7), 4639–4652. <https://doi.org/10.1007/s12035-018-1404-2> 30374941
- Ramser, E. M., Ganb, K. J., Deckera, H., Fana, E. Y., Suzukia, M., Ferreirac, S. T., & Silverman, M. A. (2013). Amyloid- β oligomers induce tau-independent disruption of BDNF axonal transport via calcineurin activation in cultured hippocampal neurons. *Molecular Biology of the Cell*, 24(16), 2494–2505. <https://doi.org/10.1091/mbc.e12-12-0858>
- Ransohoff, R. M. (2016). How neuroinflammation contributes to neurodegeneration. *Science (New York, N.Y.)*, 353(6301), 777–783. <https://doi.org/10.1126/SCIENCE.AAG2590>
- Rei, N., Valente, C. A., Vaz, S. H., Farinha-Ferreira, M., Ribeiro, J. A., & Sebastião, A. M. (2022). Changes in adenosine receptors and neurotrophic factors in the SOD1G93A mouse model of amyotrophic lateral sclerosis: Modulation by chronic caffeine. *PLoS ONE*, 17(12), e0272104. <https://doi.org/10.1371/JOURNAL.PONE.0272104>
- Reichardt, L. F. (2006). Neurotrophin-regulated signalling pathways. *Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences*, 361(1473), 1545–1564. <https://doi.org/10.1098/RSTB.2006.1894>
- Riccio, A., Ahn, S., Davenport, C. M., Blendy, J. A., & Ginty, D. D. (1999). Mediation by a CREB family transcription factor of NGF-dependent survival of sympathetic neurons. *Science (New York, N.Y.)*, 286(5448), 2358–2361. <https://doi.org/10.1126/SCIENCE.286.5448.2358>
- Riolo, G., Ricci, C., De Angelis, N., Marzocchi, C., Guerrera, G., Borsellino, G., Giannini, F., & Battistini, S. (2022). BDNF and pro-BDNF in amyotrophic lateral sclerosis: A new perspective for biomarkers of neurodegeneration. *Brain Sciences*, 12(5), 617. <https://doi.org/10.3390/BRAINSCI12050617>
- Rodrigues-Amorim, D., Rivera-Baltanás, T., Bessa, J., Sousa, N., Vallejo-Curto, M. C., Rodríguez-Jamardo, C., de Las Heras, M. E., Díaz, R., Agís-Balboa, R. C., Olivares, J. M., & Spuch, C. (2018). The neurobiological hypothesis of neurotrophins in the pathophysiology of schizophrenia: A meta-analysis. *Journal of Psychiatric Research*, 106, 43–53. <https://doi.org/10.1016/j.jpsychires.2018.09.007> 30269004
- Rogdakis, T., Charou, D., Latorrata, A., Papadimitriou, E., Tsengenesis, A., Athanasiou, C., Papadopoulou, M., Chalikiopoulou, C., Katsila, T., Ramos, I., Prousis, K. C., Wade, R. C., Sidiropoulou, K., Calogeropoulou, T., Gravanis, A., & Charalampopoulos, I. (2022). Development and biological characterization of a novel selective TrkA agonist with neuroprotective properties against amyloid toxicity. *Biomedicines*, 10(3), 614. <https://doi.org/10.3390/biomedicines10030614>
- Rosa, E., & Fahnstock, M. (2015). CREB expression mediates amyloid β -induced basal BDNF downregulation. *Neurobiology of Aging*, 36(8), 2406–2413. <https://doi.org/10.1016/j.neurobiolaging.2015.04.014> 26025137
- Rosenblad, C., Kirik, D., & Björklund, A. (2000). Sequential administration of GDNF into the substantia nigra and striatum promotes dopamine neuron survival and axonal sprouting but not striatal reinnervation or functional recovery in the partial 6-OHDA lesion model. *Experimental Neurology*, 161(2), 503–516. <https://doi.org/10.1006/EXNR.1999.7296>
- Saadipour, K., Yang, M., Lim, Y., Georgiou, K., Sun, Y., Keating, D., Liu, J., Wang, Y. R., Gai, W. P., Zhong, J. H., Wang, Y. J., & Zhou, X. F. (2013). Amyloid beta 1-42 (A β 42) up-regulates the expression of sortilin via the p75 NTR/RhoA signaling pathway. *Journal of Neurochemistry*, 127(2), 152–162. <https://doi.org/10.1111/JNC.12383>
- Saijo, K., & Glass, C. K. (2011). Microglial cell origin and phenotypes in health and disease. *Nature Reviews. Immunology*, 11(11), 775–787. <https://doi.org/10.1038/NRI3086>
- Salta, E., Lazarov, O., Fitzsimons, C. P., Tanzi, R., Lucassen, P. J., & Choi, S. H. (2023). Adult hippocampal neurogenesis in Alzheimer's disease: A roadmap to clinical relevance. *Cell Stem Cell*, 30(2), 120–136. <https://doi.org/10.1016/j.stem.2023.01.002>
- Sandrini, L., Di Minno, A., Amadio, P., Ieraci, A., Tremoli, E., & Barbieri, S. S. (2018). Association between obesity and circulating brain-derived neurotrophic factor (BDNF) levels: Systematic review of literature and meta-analysis. *International Journal of Molecular Sciences*, 19(8), 2281. <https://doi.org/10.3390/IJMS19082281>
- Schumacher, M., Weill-Engerer, S., Liere, P., Robert, F., Franklin, R. J. M., Garcia-Segura, L. M., Lambert, J. J., Mayo, W., Melcangi, R. C.,

- Parducz, A., Suter, U., Carelli, C., Baulieu, E. E., & Akwa, Y. (2003). Steroid hormones and neurosteroids in normal and pathological aging of the nervous system. *Progress in Neurobiology*, 71(1), 3–29. <https://doi.org/10.1016/J.PNEUROBIO.2003.09.004>
- Seki, M., Tanaka, T., Nawa, H., Usui, T., Fukuchi, T., Ikeda, K., Abe, H., & Takei, N. (2004). Involvement of brain-derived neurotrophic factor in early retinal neuropathy of streptozotocin-induced diabetes in rats: Therapeutic potential of brain-derived neurotrophic factor for dopaminergic amacrine cells. *Diabetes*, 53(9), 2412–2419. <https://doi.org/10.2337/DIABETES.53.9.2412>
- Sendtner, M., Arakawa, Y., Stockli, K. A., Kreutzberg, G. W., & Thoenen, H. (1991). Effect of ciliary neurotrophic factor (CNTF) on motoneuron survival. *Journal of Cell Science*, 15, 103–109. https://doi.org/10.1242/JCS.1991.SUPPLEMENT_15.14
- Sendtner, M., Dittrich, F., Hughes, R. A., & Thoenen, H. (1994). Actions of CNTF and neurotrophins on degenerating motoneurons: Preclinical studies and clinical implications. *Journal of the Neurological Sciences*, 124(SUPPL), 77–83. [https://doi.org/10.1016/0022-510X\(94\)90187-2](https://doi.org/10.1016/0022-510X(94)90187-2)
- Sendtner, M., Holtmann, B., Kolbeck, R., Thoenen, H., & Barde, Y. A. (1992). Brain-derived neurotrophic factor prevents the death of motoneurons in newborn rats after nerve section. *Nature*, 360(6406), 757–759. <https://doi.org/10.1038/360757A0>
- Sendtner, M., Kreutzberg, G. W., & Thoenen, H. (1990). Ciliary neurotrophic factor prevents the degeneration of motor neurons after axotomy. *Nature*, 345(6274), 440–441. <https://doi.org/10.1038/345440A0>
- Shanks, H. R. C., Chen, K., Reiman, E. M., Blennow, K., Cummings, J. L., Massa, S. M., Longo, F. M., Börjesson-Hanson, A., Windisch, M., & Schmitz, T. W. (2024). P75 neurotrophin receptor modulation in mild to moderate Alzheimer disease: A randomized, placebo-controlled phase 2a trial. *Nature Medicine*, 30(6), 1761–1770. <https://doi.org/10.1038/s41591-024-02977-w> 38760589
- Shen, L. L., Li, W. W., Xu, Y. L., Gao, S. H., Xu, M. Y., Bu, X. L., Liu, Y. H., Wang, J., Zhu, J., Zeng, F., Yao, X. Q., Gao, C. Y., Xu, Z. Q., Zhou, X. F., & Wang, Y. J. (2019). Neurotrophin receptor p75 mediates amyloid β -induced tau pathology. *Neurobiology of Disease*, 132, 104567. <https://doi.org/10.1016/J.NBD.2019.104567>
- Shingo, T., Date, I., Yoshida, H., & Ohmoto, T. (2002). Neuroprotective and restorative effects of intrastriatal grafting of encapsulated GDNF-producing cells in a rat model of Parkinson's disease. *Journal of Neuroscience Research*, 69(6), 946–954. <https://doi.org/10.1002/JNR.10375>
- Singh, S., Ahmad, R., Mathur, D., Kumar Sagar, R., Krishana, B., Arora, R., & Kumar Sharma, R. (2006). Neuroprotective effect of BDNF in young and aged 6-OHDA treated rat model of Parkinson disease. *Indian Journal of Experimental Biology*, 44, 699–704.
- Sperling, R., Mormino, E., & Johnson, K. (2014). The evolution of preclinical Alzheimer's disease: Implications for prevention trials. *Neuron*, 84(3), 608–622. <https://doi.org/10.1016/J.NEURON.2014.10.038>
- Sun, M., Kong, L., Wang, X., Lu, X. G., Gao, Q., & Geller, A. I. (2005). Comparison of the capability of GDNF, BDNF, or both, to protect nigrostriatal neurons in a rat model of Parkinson's disease. *Brain Research*, 1052(2), 119–129. <https://doi.org/10.1016/J.BRAINRES.2005.05.072>
- Suzuki, M., McHugh, J., Tork, C., Shelley, B., Klein, S. M., Aebischer, P., & Svendsen, C. N. (2007). GDNF secreting human neural progenitor cells protect dying motor neurons, but not their projection to muscle, in a rat model of familial ALS. *PLoS ONE*, 2(8), e689. <https://doi.org/10.1371/JOURNAL.PONE.0000689>
- Takano, R., Hisahara, S., Namikawa, K., Kiyama, H., Okano, H., & Miurat, M. (2000). Nerve growth factor protects oligodendrocytes from tumor necrosis factor- α -induced injury through Akt-mediated signaling mechanisms. *The Journal of Biological Chemistry*, 275(21), 16360–16365. <https://doi.org/10.1074/JBC.M910419199>
- Teixeira, A. L., Barbosa, I. G., Diniz, B. S., & Kummer, A. (2010). Circulating levels of brain-derived neurotrophic factor: Correlation with mood, cognition and motor function. *Biomarkers in Medicine*, 4(6), 871–887. <https://doi.org/10.2217/BMM.10.111>
- Thau, N., Jungnickel, J., Knippenberg, S., Ratzka, A., Dengler, R., Petri, S., & Grothe, C. (2012). Prolonged survival and milder impairment of motor function in the SOD1 ALS mouse model devoid of fibroblast growth factor 2. *Neurobiology of Disease*, 47(2), 248–257. <https://doi.org/10.1016/J.NBD.2012.04.008>
- Tremolizzo, L., Pellegrini, A., Conti, E., Arosio, A., Gerardi, F., Lunetta, C., Magni, P., Appollonio, I., & Ferrarese, C. (2016). BDNF serum levels with respect to multidimensional assessment in amyotrophic lateral sclerosis. *Neuro-Degenerative Diseases*, 16(3–4), 192–198. <https://doi.org/10.1159/000441916>
- Tsiperson, V., Huang, Y., Bagayogo, I., Song, Y., VonDran, M. W., DiCicco-Bloom, E., & Dreyfus, C. F. (2015). Brain-derived neurotrophic factor deficiency restricts proliferation of oligodendrocyte progenitors following cuprizone-induced demyelination. *ASN Neuro*, 7(1): 1–11. <https://doi.org/10.1177/1759091414566878>
- Tsoka, P., Matsumoto, H., Maidana, D. E., Kataoka, K., Naoumidis, I., Gravanis, A., Vavvas, D. G., & Tsilimbaris, M. K. (2018). Effects of BNN27, a novel C17-spiroepoxy steroid derivative, on experimental retinal detachment-induced photoreceptor cell death. *Scientific Reports*, 8(1), 1–12. <https://doi.org/10.1038/s41598-018-28633-1>
- Turner, B. J., Murray, S. S., Piccenna, L. G., Lopes, E. C., Kilpatrick, T. J., & Cheema, S. S. (2004). Effect of p75 neurotrophin receptor antagonist on disease progression in transgenic amyotrophic lateral sclerosis mice. *Journal of Neuroscience Research*, 78, 193–199. <https://doi.org/10.1002/jnr.20256>
- Turner, M. R., Parton, M. J., & Leigh, P. N. (2001). Clinical trials in ALS: An overview. *Seminars in Neurology*, 21(2), 167–176. <https://doi.org/10.1055/s-2001-15262>
- Tuszynski, M. H., Yang, J. H., Barba, D., Hoi-Sang, U., Bakay, R. A. E., Pay, M. M., Masliah, E., Conner, J. M., Kobalka, P., Roy, S., & Nagahara, A. H. (2015). Nerve growth factor gene therapy: Activation of neuronal responses in Alzheimer disease. *JAMA Neurology*, 72(10), 1139–1147. <https://doi.org/10.1001/JAMANEUROL.2015.1807>
- Valdo, P., Stegagno, C., Mazzucco, S., Zuliani, E., Zanusso, G., Moretto, G., Raine, C. S., & Bonetti, B. (2002). Enhanced expression of NGF receptors in multiple sclerosis lesions. *Journal of Neuropathology and Experimental Neurology*, 61(1), 91–98. <https://doi.org/10.1093/JNEN/61.1.91>
- Vilar, M., & Mira, H. (2016). Regulation of neurogenesis by neurotrophins during adulthood: Expected and unexpected roles. *Frontiers in Neuroscience*, 10, 26. <https://doi.org/10.3389/fnins.2016.00026>
- Villoslada, P., Hauser, S. L., Bartke, I., Unger, J., Heald, N., Rosenberg, D., Cheung, S. W., Mobley, W. C., Fisher, S., & Genain, C. P. (2000). Human nerve growth factor protects common marmosets against autoimmune encephalomyelitis by switching the balance of T helper cell type 1 and 2 cytokines within the central nervous system. *The Journal of Experimental Medicine*, 191(10), 1799–1806. <https://doi.org/10.1084/JEM.191.10.1799>
- Volkman, R., & Offen, D. (2017). Concise review: Mesenchymal stem cells in neurodegenerative diseases. *Stem Cells*, 35(8), 1867–1880. <https://doi.org/10.1002/STEM.2651>
- Vondran, M. W., Singh, H., Honeywell, J. Z., & Dreyfus, C. F. (2011). Levels of BDNF impact oligodendrocyte lineage cells following a cuprizone lesion. *The Journal of Neuroscience*, 31(40), 14182–14190. <https://doi.org/10.1523/JNEUROSCI.6595-10.2011>
- Wakabayashi, K., Tanji, K., Mori, F., & Takahashi, H. (2007). The Lewy body in Parkinson's disease: Molecules implicated in the formation and degradation of α -synuclein aggregates. *Neuropathology*, 27(5), 494–506. <https://doi.org/10.1111/J.1440-1789.2007.00803.X>
- Wan, R., Weigand, L. A., Bateman, R., Griffioen, K., Mendelowitz, D., & Mattson, M. P. (2014). Evidence that BDNF regulates heart rate by a mechanism involving increased brainstem parasympathetic neuron

- excitability. *Journal of Neurochemistry*, 129, 573–580. <https://doi.org/10.1111/jnc.12656>
- Wang, J., Hu, W., Feng, Z., & Feng, M. (2021). BDNF-overexpressing human umbilical cord mesenchymal stem cell-derived motor neurons improve motor function and prolong survival in amyotrophic lateral sclerosis mice. *Neurological Research*, 43(3), 199–209. <https://doi.org/10.1080/01616412.2020.1834775>
- Wang, J., Hu, W. W., Jiang, Z., & Feng, M. J. (2020). Advances in treatment of neurodegenerative diseases: Perspectives for combination of stem cells with neurotrophic factors. *World Journal of Stem Cells*, 12(5), 323–338. <https://doi.org/10.4252/WJSC.V12.I5.323>
- Wang, Y. J., Wang, X., Lu, J. J., Li, Q. X., Gao, C. Y., Liu, X. H., Sun, Y., Yang, M., Lim, Y., Evin, G., Zhong, J. H., Masters, C., & Zhou, X. F. (2011). p75NTR regulates A β deposition by increasing A β production but inhibiting A β aggregation with its extracellular domain. *Journal of Neuroscience*, 31(6), 2292–2304. <https://doi.org/10.1523/JNEUROSCI.2733-10.2011>
- Weill-Engerer, S., David, J. P., Sazdovitch, V., Liere, P., Eychenne, B., Planos, A., Schumacher, M., Delacourte, A., Baulieu, E. E., & Akwa, Y. (2002). Neurosteroid quantification in human brain regions: Comparison between Alzheimer's and nondemented patients. *The Journal of Clinical Endocrinology & Metabolism*, 87(11), 5138–5143. <https://doi.org/10.1210/JC.2002-020878>
- Weinstein, G., Beiser, A. S., Choi, S. H., Preis, S. R., Chen, T. C., Vargha, D., Au, R., Pikula, A., Wolf, P. A., DeStefano, A. L., Vasan, R. S., & Seshadri, S. (2014). Serum brain-derived neurotrophic factor and the risk for dementia. *JAMA Neurology*, 71(1), 55. <https://doi.org/10.1001/jamaneurol.2013.4781>
- Wellmer, A., Misra, V. P., Sharief, M. K., Kopelman, P. G., & Anand, P. (2001). A double-blind placebo-controlled clinical trial of recombinant human brain-derived neurotrophic factor (rhBDNF) in diabetic polyneuropathy. *Journal of the Peripheral Nervous System : JPNS*, 6(4), 204–210. <https://doi.org/10.1046/J.1529-8027.2001.01019.X>
- Whone, A., Luz, M., Boca, M., Woolley, M., Mooney, L., Dharia, S., Broadfoot, J., Cronin, D., Schroers, C., Barua, N. U., Longpre, L., Barclay, C. L., Boiko, C., Johnson, G. A., Fibiger, H. C., Harrison, R., Lewis, O., Pritchard, G., Howell, M., ... Gill, S. S. (2019). Randomized trial of intermittent intraputamenal glial cell line-derived neurotrophic factor in Parkinson's disease. *Brain: A Journal of Neurology*, 142(3), 512–525. <https://doi.org/10.1093/BRAIN/AWZ023>
- Wong, B. X., Tsatsanis, A., Lim, L. Q., Adlard, P. A., Bush, A. I., & Duce, J. A. (2014). B-Amyloid precursor protein does not possess ferroxidase activity but does stabilize the cell surface ferrous iron exporter ferroportin. *PLoS ONE*, 9(12), e114174. <https://doi.org/10.1371/journal.pone.0114174>
- Xie, Y., Meeker, R. B., Massa, S. M., & Longo, F. M. (2019). Modulation of the p75 neurotrophin receptor suppresses age-related basal forebrain cholinergic neuron degeneration. *Scientific Reports*, 9(1), 5273. <https://doi.org/10.1038/s41598-019-41654-8>
- Xu, J., Lou, S., Huang, H., Xu, J., & Luo, F. (2024). Regulation and crosstalk of cells and factors in the pancreatic cancer microenvironment. *Current Cancer Drug Targets*, 100(9), 1029–1048. <https://doi.org/10.2174/0115680096317840240723071018>
- Yaar, M., Zhai, S., Pilch, P. F., Doyle, S. M., Eisenhauer, P. B., Fine, R. E., & Gilchrist, B. A. (1997). Binding of beta-amyloid to the p75 neurotrophin receptor induces apoptosis. A possible mechanism for Alzheimer's disease. *The Journal of Clinical Investigation*, 100(9), 2333–2340. <https://doi.org/10.1172/JCI119772>
- Yan, Q., Elliott, J., & Snider, W. D. (1992). Brain-derived neurotrophic factor rescues spinal motor neurons from axotomy-induced cell death. *Nature*, 360(6406), 753–755. <https://doi.org/10.1038/360753a0>
- Yan, Z., Shi, X., Wang, H., Si, C., Liu, Q., & Du, Y. (2021). Neurotrophin-3 promotes the neuronal differentiation of BMSCs and improves cognitive function in a rat model of Alzheimer's disease. *Frontiers in Cellular Neuroscience*, 15, 629356. <https://doi.org/10.3389/fncel.2021.629356>
- Yang, C., Liu, Y., Ni, X., Li, N., Zhang, B., & Fang, X. (2014). Enhancement of the nonamyloidogenic pathway by exogenous NGF in an Alzheimer transgenic mouse model. *Neuropeptides*, 48(4), 233–238. <https://doi.org/10.1016/J.NPEP.2014.04.005>
- Yao, X. Q., Jiao, S. S., Saadipour, K., Zeng, F., Wang, Q. H., Zhu, C., Shen, L. L., Zeng, G. H., Liang, C. R., Wang, J., Liu, Y. H., Hou, H. Y., Xu, X., Su, Y. P., Fan, X. T., Xiao, H. L., Lue, L. F., Zeng, Y. Q., Giunta, B., ... Wang, Y. J. (2015). p75NTR ectodomain is a physiological neuroprotective molecule against amyloid-beta toxicity in the brain of Alzheimer's disease. *Molecular Psychiatry*, 20(11), 1301–1310. <https://doi.org/10.1038/mp.2015.49>
- Yi, X., Manickam, D. S., Brynskikh, A., & Kabanov, A. V. (2014). Agile delivery of protein therapeutics to CNS. *Journal of Controlled Release : Official Journal of the Controlled Release Society*, 190, 637–663. <https://doi.org/10.1016/J.JCONREL.2014.06.017>
- Yilmaz, C., Rogdakis, T., Latorrata, A., Thanou, E., Karadima, E., Papadimitriou, E., Siapi, E., Li, K. W., Katsila, T., Calogeropoulou, T., Charalampopoulos, I., & Alexaki, V. I. (2022). ENT-A010, a novel steroid derivative, displays neuroprotective functions and modulates microglial responses. *Biomolecules*, 12(3), 1–22. <https://doi.org/10.3390/biom12030424>
- Yu, Y., Chen, H., & Su, S. B. (2015). Neuroinflammatory responses in diabetic retinopathy. *Journal of Neuroinflammation*, 12(1), 1–15. <https://doi.org/10.1186/S12974-015-0368-7>
- Zeng, F., Lu, J. J., Zhou, X. F., & Wang, Y. J. (2011). Roles of p75NTR in the pathogenesis of Alzheimer's disease: A novel therapeutic target. *Biochemical Pharmacology*, 82(10), 1500–1509. <https://doi.org/10.1016/J.BCP.2011.06.040>
- Zlokovic, B. V. (2008). The BBB in health and chronic neurodegenerative disorders. *Neuron*, 57(2), 178–201. <https://doi.org/10.1016/J.NEURON.2008.01.003>
- Zota, I., Chanoumidou, K., Charalampopoulos, I., & Gravanis, A. (2024). Dynamics of myelin deficits in the <sc>5x</sc>FAD mouse model for Alzheimer's disease and the protective role of <sc>BDNF</sc>. *Glia*, 72(4), 809–827. <https://doi.org/10.1002/glia.24505>
- Zoupa, E., Gravanis, A., & Pitsikas, N. (2019). The novel dehydroepiandrosterone (DHEA) derivative BNN27 counteracts behavioural deficits induced by the NMDA receptor antagonist ketamine in rats. *Neuropharmacology*, 151, 74–83. <https://doi.org/10.1016/J.NEUROPHARM.2019.04.001>

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