



We Are What We Eat: Ubiquitin–Proteasome System (UPS) Modulation Through Dietary Products

15

Eleni Panagiotidou and Niki Chondrogianni

Abstract

During lifetime, the molecular mechanisms that are responsible for cellular defense against adverse conditions such as oxidative and heat stress tend to be less efficient, thus gradually leading to the natural phenomenon of aging. Aging is linked to increased oxidative stress and is characterized by the accumulation of damaged macromolecules. The accumulation of oxidized and misfolded proteins is also accusable for various neurodegenerative pathologies that are linked to aging. Among self-defense mechanisms of cells, proteostasis network is responsible for the proper biogenesis/folding/trafficking of proteins and their elimination through proteolysis. The ubiquitin-proteasome system (UPS) is the major proteolytic mechanism that has attracted the interest of many researchers as an antiaging target. Interestingly, many natural compounds have been identified as potent UPS activators. Given that diet is a manageable environmental factor that affects aging, consumption of natural dietary products that may

potentially enhance the UPS function, would contribute to increased health span and delayed onset or progression of age-related disorders. Herein, we summarize natural compounds and extracts derived from edible products that have exhibited antiaging and anti-aggregation properties and the beneficial properties have been linked to the UPS modulation.

Keywords

Ubiquitin–proteasome system (UPS) · Proteasome modulation · Aging · Age-related diseases · Dietary compounds

Abbreviations

| | |
|-----------|--|
| A β | Amyloid β |
| AD | Alzheimer's disease |
| ALP | Autophagy-lysosome pathway |
| APP | Amyloid precursor protein |
| C-L | Caspase-like |
| CP | Core proteasome |
| CT-L | Chymotrypsin-like |
| DUB | Deubiquitinating enzymes/ deubiquitinases |
| D3T | 3H-1,2dithiole-3-thione |
| EVOO | Extra virgin olive oil |
| GHE | Guarana hydroalcoholic extract |
| HD | Huntington's disease |
| HGPS | Hutchinson-Gilford progeria syndrome |

E. Panagiotidou
Institute of Chemical Biology, National Hellenic Research
Foundation, Athens, Greece

Department of Biochemistry and Biotechnology,
University of Thessaly, Larissa, Greece
e-mail: epanagiotidou@eie.gr

N. Chondrogianni (✉)
Institute of Chemical Biology, National Hellenic Research
Foundation, Athens, Greece
e-mail: nikichon@eie.gr

| | |
|-----------------|---|
| HSP | Heat shock protein |
| HTT | Huntingtin |
| NHDF | Normal human dermal fibroblasts |
| Nrf2 | Nuclear factor (erythroid-derived 2)-like 2 |
| PD | Parkinson's disease |
| PDMs | Polyphenol digested metabolites |
| PKA | Protein kinase A |
| PolyQ | Polyglutamine |
| RP | Regulated particle |
| UPS | Ubiquitin-proteasome system |
| T-L | Trypsin-like |
| 6-OHDA | 6-hydroxydopamine |
| 18 α -GA | 18 α -glycyrrhetic acid |

15.1 Introduction

Protein integrity is vital for the normal function of all cell types. To maintain a healthy proteome, cells have developed a protein quality control system which is crucial for cell metabolism and stress adaptation. The proteostasis network is responsible for: (a) protein synthesis, stabilization, folding mostly by chaperons of the heat shock protein (HSP) family and trafficking, and (b) protein degradation mainly through the ubiquitin–proteasome system (UPS) and the autophagy–lysosome pathway (ALP) [1, 2]. Molecular chaperons recognize misfolded protein species and if the repair is impossible, they cooperate with the proteolytic systems favoring the degradation of these proteins [3, 4]. Attenuation of proteostasis mechanisms is linked to aging and is considered as one of the hallmarks of aging [5, 6]. Moreover, impaired function of proteostasis has also been reported in various age-related disorders such as Alzheimer's (AD), Parkinson's (PD) and Huntington's (HD) diseases which are characterized by chronic expression of misfolded or aggregated proteins [7].

The average life expectancy has progressively elevated in the last decades. Subsequently, the development of age-related disorders has ensued this rapid increase. Aging and age-related

neurodegeneration have been associated with increased oxidative stress but also with nutrients and their metabolites that may alleviate these toxic processes, playing a pivotal role in the battle against organismal time decline [8]. For example, a plethora of studies gives prominence to the antiaging properties of polyphenolic compounds (phenolic acids, flavonoids, stilbenes, lignans) [9]. Given that these substances mainly contribute to plant protection from adverse environmental conditions through their antioxidant activities, it is possible to be effective as antiaging factors in mammalian cells as well [10]. These compounds can be easily isolated from plants and are included in daily nutritional products like fruits, vegetables, and cereals.

Among the proteostasis mechanisms, the UPS system has been extensively investigated for its involvement in aging and age-related disorders [11]. Due to its capacity to degrade prone-to-aggregation proteins and oxidatively damaged proteins, UPS activation by natural or synthetic substances consists a desirable strategy against aging and aggregation-related disorders [12–14].

Herein, we review natural compounds that have been shown to positively affect the UPS and to exert beneficial properties against aging and age-related disorders. The reported substances can be found in edible natural products that we daily consume in the context of a normal nutrition or in dietary supplements.

15.2 The UPS System

The UPS is a key player for the degradation of short-lived, misfolded, and damaged proteins and under normal conditions it is involved in cell cycle progression, apoptosis, and cell proliferation, among others [15]. The proteasome is a large multi-subunit enzymatic complex that is the core of the UPS system catalyzing the ATP/ubiquitin-dependent and independent proteolysis (Fig. 15.1) [16, 17].

Ubiquitin is a conserved protein that modifies and tags proteins for degradation through the

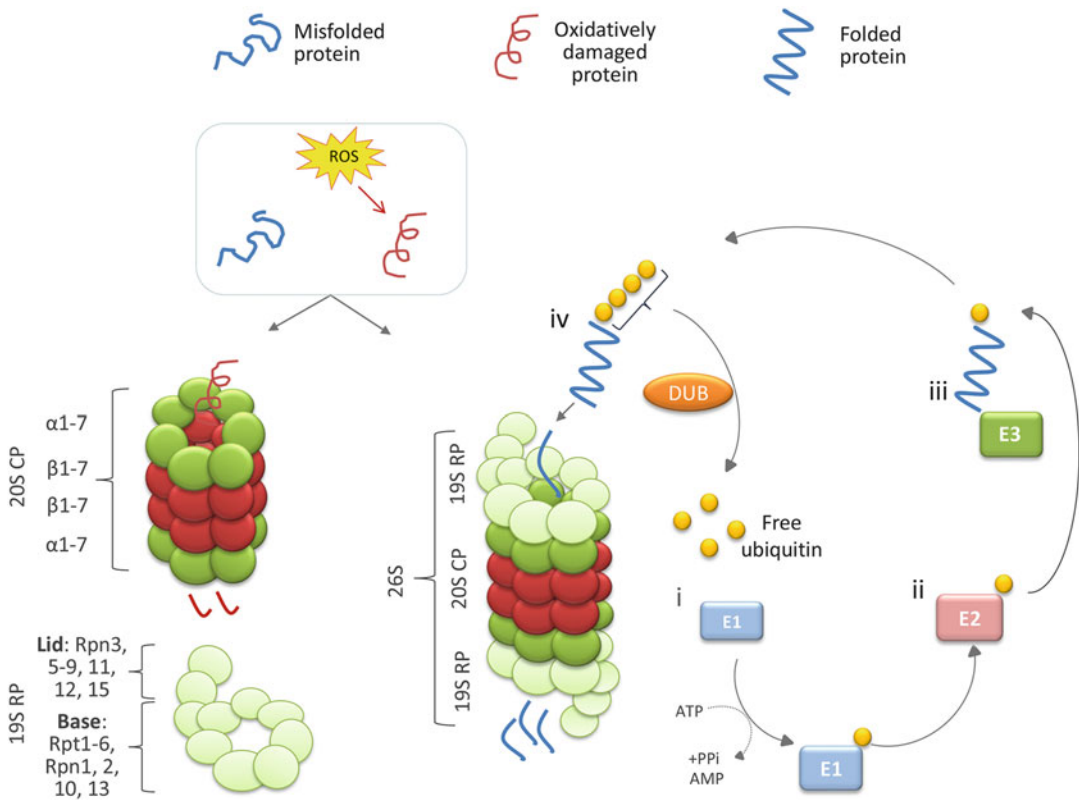


Fig. 15.1 The UPS system. Misfolded, oxidatively damaged or normally folded proteins are degraded in an ATP/ubiquitin-dependent or independent manner by the 26S and 20S proteasomes, respectively

ubiquitination process which includes three steps. (i) The E1 ubiquitin-activating enzyme activates the ubiquitin which (ii) is transferred to E2 ubiquitin-conjugating enzyme. Subsequently, the ubiquitin-charged E2 binds to (iii) the E3 ubiquitin-ligase enzyme which catalyzes the final step of ubiquitination. E3 ligase carries the substrate for degradation and thus ubiquitin is transferred to the substrate. This cycle is repeated and results in the poly-ubiquitination of the target protein which in turn (iv) is recognized and degraded by the 26S proteasome followed by the release of the ubiquitin molecules (through the action of deubiquitinases) to be reused [17].

The 26S/30S (referred to 26S proteasome hereafter) proteasome is formed following the capping of the 20S core proteasome (CP) by one (26S) or two (30S) 19S regulated particles (RP) and is mostly responsible for the degradation of

ubiquitinated proteins. The 20S proteasome is composed of 28 subunits (14- α type and 14- β type). The 20S subunits form four rings with seven subunits of the same type per two rings in a barrel-like structure. The two inner rings are formed by β subunits (β 1-7) and exert proteolysis through β 1 (caspase-like/C-L), β 2 (trypsin-like/T-L), and β 5 (chymotrypsin-like/CT-L) subunits. The two outer rings are formed by α -type subunits and create a gated channel through which the protein substrate reaches the three catalytic centers [18]. The 19S RP consists of two sub-complexes: the lid that consists of nine subunits (Rpn3, Rpn 5-Rpn9, Rpn11, Rpn12, and Rpn15) and the base, which interacts directly with the 20S core and is composed of hexameric AAA-ATPases (Rpt1-6) and tetrameric non-ATP-ases namely Rpn1, Rpn2, Rpn10, and Rpn13. Rpn11 subunit possesses deubiquitination activity while Rpn3 and Rpn10

Table 15.1 Human genes that encode the 26S/30S and 20S proteasome subunits

| 20S CP | | | | 19S RP | | | |
|-------------------|-------|------------------|-------|---------------|-------|--------------|--------|
| α subunits | Gene | β subunits | Gene | Base subunits | Gene | Lid subunits | Gene |
| $\alpha 6$ | PSMA1 | $\beta 1$ | PSMB6 | Rpt1 | PSMC2 | Rpn3 | PSMD3 |
| $\alpha 2$ | PSMA2 | $\beta 2$ | PSMB7 | Rpt2 | PSMC1 | Rpn5 | PSMD12 |
| $\alpha 7$ | PSMA3 | $\beta 3$ | PSMB3 | Rpt3 | PSMC4 | Rpn6 | PSMD11 |
| $\alpha 3$ | PSMA4 | $\beta 4$ | PSMB2 | Rpt4 | PSMC6 | Rpn7 | PSMD6 |
| $\alpha 5$ | PSMA5 | $\beta 5$ | PSMB5 | Rpt5 | PSMC3 | Rpn8 | PSMD7 |
| $\alpha 1$ | PSMA6 | $\beta 6$ | PSMB1 | Rpt6 | PSMC5 | Rpn9 | PSMD13 |
| $\alpha 4$ | PSMA7 | $\beta 7$ | PSMB4 | Rpn1 | PSMD2 | Rpn11 | PSMD14 |
| | | | | Rpn2 | PSMD1 | Rpn12 | PSMD8 |
| | | | | Rpn10 | PSMD4 | Rpn15 | SEMI |
| | | | | Rpn13 | ADRM1 | | |

serve as ubiquitin receptors [19, 20] (Table 15.1). In the 26S proteasome ubiquitin-ATP-dependent proteolysis, proteins tagged with ubiquitin are recognized by Rpn3 and Rpn10 receptors and deubiquitinating enzymes (DUB) like Rpn11, USP14, and UCH37 remove the ubiquitin tag [21]. Subsequently, the base of the 19S complex unfolds the protein and promotes it to the catalytic centers for degradation [22]. On the other hand, the 20S proteasome has been suggested to promote the degradation of unfolded and oxidized proteins in an ATP-independent manner and does not require poly-ubiquitination of target proteins and 19S-mediated protein unfolding [16] (Fig. 15.1).

15.3 Proteasome Activation

Proteasome activation is feasible through genetic manipulation, post-translational modifications of specific proteasome subunits, or conformational alterations of the 20S core structure [14]. Treatment with specific compounds (including some that are part of our diet) can also modulate the expression and/or the function of the proteasome and they will be further presented below (Fig. 15.2).

The transcription factor Nrf2 (Nuclear factor (erythroid-derived 2)-like 2) is the key response factor against oxidative stress that regulates the induction of the expression of various antioxidant enzymes [23]. A cross talk between Nrf2 and the proteasome has been reported and several studies have shown that Nrf2 is able to modulate the 20S and 19S gene expression following oxidative

stress or dietary Nrf2 modulation [24, 25]. Apart from Nrf2, Rpn4 is another transcription factor in yeast that has been shown to regulate proteasome biogenesis. Under normal conditions, Rpn4 is short lived and rapidly degraded by the proteasome. However, upon proteasome inhibition following proteotoxic stress, Rpn4 is stabilized to upregulate the expression of proteasome genes. With regard to genetic manipulation of the proteasome, we have previously demonstrated that stable overexpression of the $\beta 5$ catalytic subunit or the hUPM1/POMP chaperone in human fibroblasts may induce all three proteasomal activities [26, 27], while PSMD11 (RPN-6 in *C. elegans*) has also been shown to be correlated with increased proteasome assembly and activity [123, 124].

Proteasome activation has also been succeeded through post-translational modifications such as phosphorylation. 26S upregulation has been achieved through cAMP-dependent protein kinase A (PKA) phosphorylation of the Rpt6 proteasome subunit [28]. Moreover, PKA-mediated phosphorylation of Rpn6 has been shown to enhance the degradation of ubiquitinated proteins [29]. Additionally, inhibition of the proteasome deubiquitinases such as USP14 and UCH37 resulted in elevated proteasome activities [30]. Finally, various peptides and compounds have been shown to favor proteasome function through conformational alterations that mainly affect the α -gated channel of the 20S core. Molecules that modulate 20S and/or 26S proteasome function are known as gate-openers or stimulators [14].

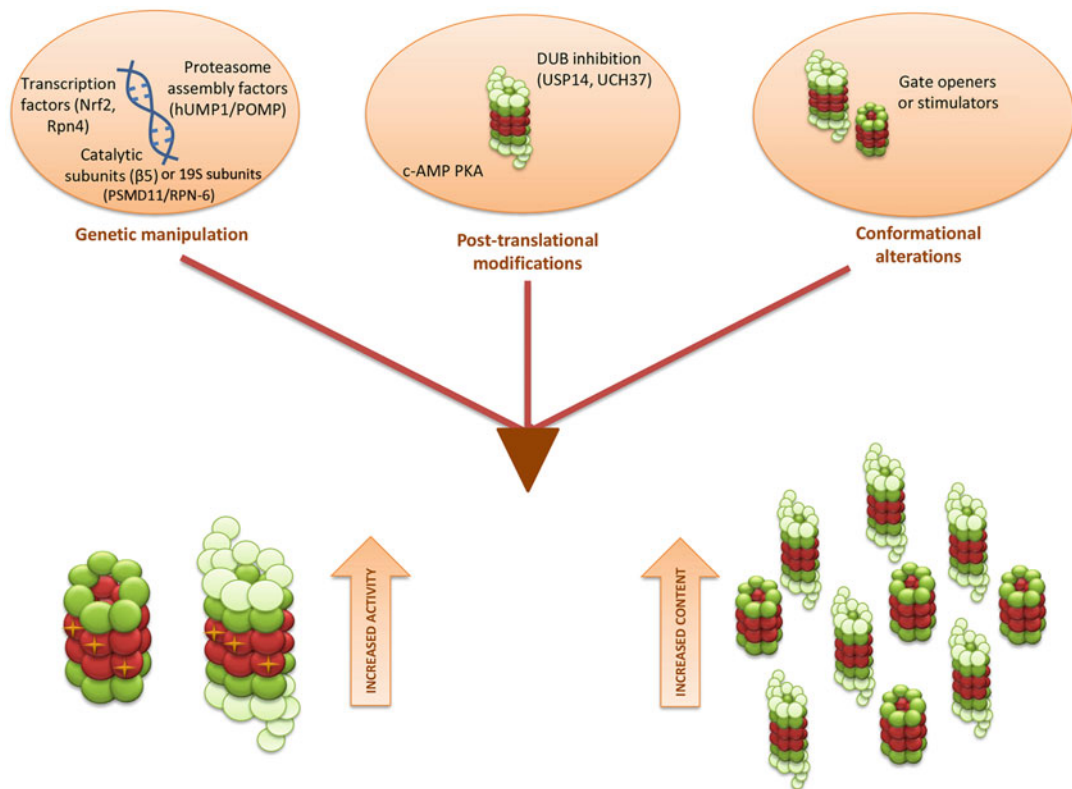


Fig. 15.2 Proteasome activation. Increased proteasome activity or content may occur through genetic manipulation, post-translational modifications, and conformational alterations. Natural or synthetic compounds can modulate

proteasome activity and function either directly as stimulators or through modulation of molecules responsible for proteasome activation

15.4 UPS Modulation in Aging and Neurodegenerative Diseases

Aging is a natural decline of the organism and its function during lifetime affects, in several rhythms, almost whole animal and plant kingdoms. This natural process is characterized by the attenuation of self-defense mechanisms and the decline of homeostatic mechanisms. Impairment of proteostasis has been associated with aging and results in reduced capacity of cells to repair misfolded or damaged proteins or to remove them through degradation pathways [31]. Proteasome activation has been shown to decelerate aging *in vivo* and *in vitro* [32]. The accumulation of misfolded proteins also characterizes many neurodegenerative disorders

that are linked to aging such as AD, HD, PD, and Hutchinson–Gilford progeria syndrome (HGPS).

AD is the most common cause of dementia and is characterized by loss of synaptic connections and brain atrophy. The accumulation of amyloid β ($A\beta$) plaques and hyper-phosphorylated Tau protein in neurofibrillary tangles are distinctive AD features [33]. $A\beta$ is formed by the Amyloid Precursor Protein (APP) when the latter is cleaved by β - and γ -secretases. The proteasome is one of the major agents for $A\beta$ metabolism, albeit the $A\beta$ aggregates lead to proteasome impairment resulting in accumulation of aggregates in a vicious cycle [34]. Additionally, the E3 ubiquitin ligase HRD1 and the deubiquitinase UCHL-1 are downregulated during AD further highlighting the importance of UPS restoration in this disorder [35, 36].

Huntingtin (HTT) is the most involved protein in HD pathogenesis. Mutations in *HTT* gene are related to expansion of the CAG triplet resulting in a polyglutamine (polyQ) tract in the N-terminus of the HTT protein [37]. Like in other proteinopathies, there is evidence of the UPS implication in HD. Mutations in ubiquitin UBB⁺, PARKIN, NHLRC1 (an E3 ubiquitin ligase), and reduction of E1 and E2 ubiquitin enzyme activities along with 26S proteasome inhibition by HTT are characteristic alterations that lead to the UPS impairment in HD [38, 39].

PD is characterized by the accumulation of Lewy bodies (eosinophilic intracytoplasmic inclusions); α -synuclein is the main component [40]. α -synuclein has been shown to impede both the 20S and 26S proteasome function [41]. Moreover PARKIN, an E3 ligase, is the second most involved protein in PD pathology. PARKIN has been shown to enhance the interaction between 19S proteasome subunits underlining the importance of this ligase on the 26S proteasome activation [42]. PARKIN is affected by Lewy bodies that along with PARKIN mutations result in the degradation impairment observed in PD [43, 44].

Finally, progeroid syndromes are genetic conditions characterized by premature aging. Given that aging is accompanied by impairment of proteostatic mechanisms subsequently the UPS may also affect this type of syndromes. HGPS is a genetic condition in which a point mutation in the *LMNA* gene is observed. There are not many studies referring to UPS implication in HGPS; however, the downregulation of β 5 and β 7 proteasome subunits and reduction of CT-L proteasome activity have been reported [45]. Therefore, the UPS also appears as an attractive target against HGPS.

15.5 UPS and Diet

Several compounds have been shown to induce the UPS expression and/or function and thus to affect the progression of aging and age-related disorders. These compounds are summarized in the following sections and in Table 15.2.

15.5.1 Aggregation-Related Disorders

Well known for its stimulant activity on the central nervous system, **guarana** (*Paullinia cupana*) is consumed widely in Brazil in high-energy drinks and dietary supplements. Guarana compounds such as polyphenols, caffeine, and other methylxanthines have been associated with its beneficial biological properties which include antioxidant, antimicrobial, and chemopreventive activities in carcinogenesis [75]. Moreover, guarana powder has been shown to prevent A β aggregation in human neuronal-like cells while guarana water extract reduced the formation of polyQ aggregates expressed in *C. elegans*, suggesting a beneficial effect in AD and HD, respectively [76, 77]. According to a recent study, treatment with guarana hydroalcoholic extract (GHE) increased proteasomal and lysosomal activities and reduced intracellular ROS in *C. elegans*. It was also demonstrated that GHE eliminated the toxic phenotypes associated with protein misfolding and accumulation in nematode models of HD and AD. All the data above suggest the contribution of protein degradation mechanisms in GHE-mediated protection against polyQ protein aggregation and A β -induced toxicity [46].

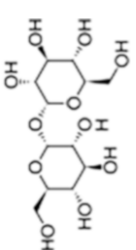
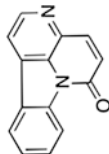
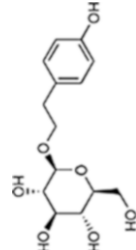
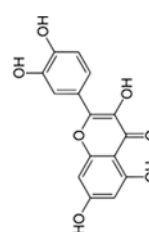
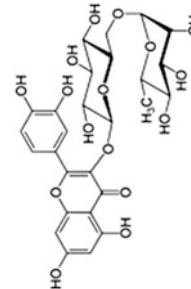
Numerous studies have extensively investigated the pivotal role of the Mediterranean Diet in the prevention of a wide range of age-related pathological conditions [78]. The extra virgin olive oil (EVOO) is one of the most characteristic components of the Mediterranean Diet and it has attracted the interest of many researchers due to its antioxidant and antiaging properties while many of EVOO's ingredients have been proposed as nutraceuticals against AD disease [79, 80]. **Oleocanthal** is one of the EVOO's phenols responsible for its bitter and pungent taste. Many studies demonstrated the potential neuroprotective and anti-aggregation effects of oleocanthal [81–83]. According to a recent study, oleocanthal upregulated the expression of *PSMD1* and *PSMB4* subunits in neuron-like SH-SY5Y cells under stress and normal conditions, suggesting the involvement of oleocanthal in the degradation of misfolded proteins rather than oxidized ones. Moreover, oleocanthal increased


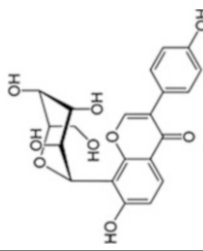
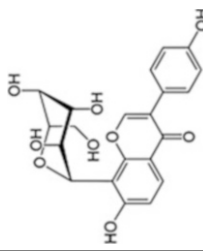
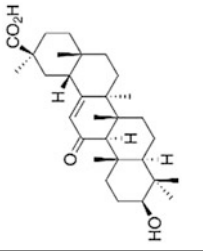
Table 15.2 Natural compounds/extracts that trigger UPS activation and affect aging and/or age-related disorders

| Natural compound/ extract | Structure | Source | UPS target | Model | Disorder | References |
|--|-----------|---|--|---|----------|------------|
| Guarana hydroalcoholic extract (GHE) ^a | | <i>Paulinia cupana</i> | CT-L activity | <i>C. elegans</i> (CL2006, CL4176, AM141) | AD/HD | [46] |
| Oleocanthal | | Extra virgin olive oil (EVOO) | <i>PSMD1</i> , <i>PSMB4</i> | SH-SY5Y cells | AD | [47] |
| Morin | | Leaves of <i>Psidium guajava</i> (guava tea), white mulberry and red wine | 26S proteasome | APP695- transfected SH-SY5Y cells | AD | [48] |
| Isoquercitrin | | <i>Mangifera indica</i> (mango), leaves of <i>Ammonia squamosa</i> and <i>Camellia sinensis</i> | 26S proteasome | APP695- transfected SH-SY5Y cells | AD | [48] |
| Resveratrol | | Blueberries, raspberries, peanuts, the skin of grapes, red wine, cocoa and various other herbs | Aβ proteasome-dependent reduction CT-L activity, UBA-1 T-L activity | APP695- transfected HEK293 cells <i>C. elegans</i> (CL2006) Mouse (3xTg- AD) (cerebral cortex) | AD | [49–51] |

(continued)

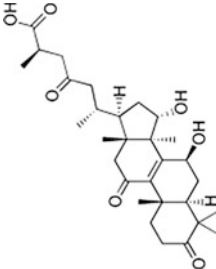
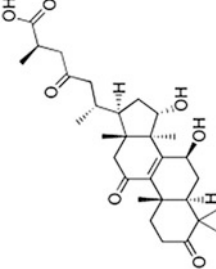

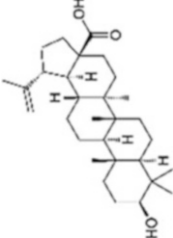
Table 15.2 (continued)



| Natural compound/ extract | Structure | Source | UPS target | Model | Disorder | References |
|------------------------------|--|--|---|--|----------|------------|
| Trehalose |  | Honey, mushrooms, lobsters, shrimps, wine | CT-L activity | Glial cultures from R6/1 mouse model of HD | HD | [52] |
| Canthin-6-one |  | Extract of many plants including <i>Zanthoxylum chiloperone</i> , <i>Eurycoma longifolia</i> and <i>Aerva lanata</i> | CT-L activity, <i>Psm1</i> | PC12 cells | PD | [54] |
| Salidroside |  | <i>Rhodiola rosea</i> | PARKIN, UCH-L1, CT-L activity | A30P α -synuclein-transfected SH-SY5Y cells | PD | [55] |
| Quercetin |  | Variety of widely consumed products including green tea, honey, red wine, and onions | <i>uba-1</i> , <i>ubq-1</i> , CT-L activity | <i>C. elegans</i> (CL2006) | AD | [56] |
| Rutin |  | Variety of plants including citrus fruits | CT-L activity | APP695-transfected SH-SY5Y cells | Aging | [58] |
| | | | CT-L activity | HFL-1 cells | HD | [59] |
| | | | CT-L activity | mHTT-transfected Neuro2a cells | AD | [57] |

| | | | | | | |
|---|--|--|---|--|-------|----------|
| Sulforaphane |  | Cruciferous vegetables | CT-L activity, T-L activity, C-L activity | mHTT-transfected HEK293 cells Fibroblasts from patients with HGPS | HD | [60] |
| Polyphenol digested metabolites (PDMs) from leaves and fruits of <i>Arbutus unedo</i> | | <i>Arbutus unedo</i> (strawberry tree) | CT-L activity, PSMC2 | hESCs (human embryonic stem cells) | Aging | [62] |
| Ginkgo biloba extract EGb761 |  | Ginkgo biloba | <i>PSMB6</i> , <i>PSMB6</i> , <i>PSMB7</i> | Yeast cells transformed with α -synuclein | PD | [63] |
| Puerarin |  | <i>Pueraria lobata</i> | CT-L activity | MPP ⁺ -treated SH-SY5Y cells | PD | [65] |
| 18 α -glycyrrhetic acid (18 α -GA) |  | <i>Glycyrrhiza radix</i> | CT-L activity | <i>C. elegans</i> (CL2006, CL4176, CL2331) SH-SY5Y cells exposed to 7PA2-conditioned medium | AD | [66] |
| | | | CT-L activity, T-L activity, PBS-5, PAS-1-7, RPT-6 CT-L activity, T-L activity, C-L activity | <i>C. elegans</i> HFL-1 cells | Aging | [66, 67] |

(continued)

Table 15.2 (continued)

| Natural compound/ extract | Structure | Source | UPS target | Model | Disorder | References |
|---|--|---|---|-------------------------------------|----------|------------|
| Ganoderic acid DM |  | Ganoderma mushrooms | Proteasome function | T-REx293 expressing Aβ42-EGFP | AD | [68] |
| Polysaccharide 5 |  | <i>Rubia cordifolia</i> L. | Proteasome function | T-REx293 expressing Aβ42-EGFP | AD | [68] |
| 3H-1,2dithiole-3-thione (D3T) ^a |  | Cruciferous vegetables | CT-L activity, T-L activity, C-L activity, PSMA1, PSMA4, PSMB3, PSMB5, PSMB6 | Mouse | AD | [69] |
| Betulinic acid ^a |  | Many plants including <i>Betula pubescens</i> , <i>Ziziphus mauritiana</i> and <i>Prunella vulgaris</i> | CT-L activity | Purified proteasome | AD | [70] |

| | | | | | |
|--------------------|---|--|---|---------------------------|------------|
| Oleuropein |  <p>The image shows the chemical structure of Oleuropein, a polyphenolic compound. It consists of a glucose molecule linked to a hydroxytyrosol unit, which is further linked to a caffeoyl group. The caffeoyl group has a double bond and a hydroxyl group on the aromatic ring.</p> | Green olives, olive leaves and EVOO | CT-L activity, T-L activity, C-L activity | IMR90, W138 cells | Aging [71] |
| Lipoic acid |  <p>The image shows the chemical structure of Lipoic acid, which is a cyclohexane ring with two sulfur atoms at the 1 and 2 positions, and a hexanoic acid chain at the 3 position.</p> | Broccoli, spinach, tomatoes and brussels sprouts | CT-L activity, PSMB5 | NHDF cells | Aging [72] |
| Bee pollen extract | | Bee pollen | CT-L activity | HFL-1 cells | Aging [73] |
| Zinc | | Plethora of natural edible products | CT-L activity | Humans (population study) | Aging [74] |

^aCompounds that have demonstrated anti-aggregation properties, but the association, if there is any, between their potency to act as proteasome activators and their contribution in protection against age-related disorders has not been studied yet

HSP90 levels, a chaperone that is essential for proteasome integrity and function [47, 84, 85].

Morin is a flavonol present in guava leaf tea (*Psidium guajava*), white mulberry (*Morus Alba L.*), and red wine. It is known for its antioxidant activity as well for its antiapoptotic and neuroprotective properties [86, 87]. Regarding its neuroprotective effects in AD, morin restricted β - and γ -secretase activities to impede A β aggregation as well as to disaggregate preformed fibrils and to act against H₂O₂-induced stress in APP695-transfected SH-SY5Y cells. In the same cell line, morin positively affected the 26S proteasome activity under H₂O₂ stress conditions. The same results were also produced by another flavonoid namely **isoquercitrin** [48].

The potential therapeutic properties of **resveratrol** including life span extension, neuroprotection, and cytoprotection, foreshadow the potential contribution of this polyphenol in age-related disorders [88, 89]. Resveratrol's effects on neurodegenerative disorders have been studied both in cellular and organismal level in models of AD; the effects on UPS among their positive outcomes have also been investigated [90]. For instance, in APP695-transfected HEK293 cell line, resveratrol reduced A β levels through proteasomal degradation [49]. The same phenotype was reported in *C. elegans*; treatment with resveratrol reduced A β aggregates in CL2006 strain which expresses the human A β 1-42 and paralyzes as A β aggregates accumulate [91]. Reduced A β aggregates have also been linked to proteostasis factors including UBA-1 (the only ubiquitin-activating enzyme in *C. elegans*) and enhanced proteasome activity [50]. Resveratrol was also shown to activate the brain proteasome function in the 3xTg-AD mouse model. Moreover, resveratrol administration enhanced the levels of neprilysin, an amyloid-degrading enzyme and diminished the levels of the amyloidogenic secretase BACE-1 [51]. On top of these effects, the already known effect of resveratrol on deacetylase SIRT1 [92] seems to dictate a preferential degradation of phosphorylated-Tau through enhanced deacetylation of Tau [51]. The abovementioned biochemical alterations led to

ameliorated cognitive behavior and function. Natural sources of resveratrol are blueberries, raspberries, peanuts, the skin of grapes, red wine, cocoa and various other herbs [93].

Trehalose is a disaccharide that is characterized as an autophagy enhancer [94]. During the last decades, many studies have reported the longevity effects of trehalose and its neuroprotective properties against HD [95]. In vivo experiments with mouse models of HD showed that trehalose administration reversed many aspects of pathological phenotype caused by polyQ aggregates [96]. In HD human fibroblasts, trehalose increased proteasome activities and decreased HTT protein levels [53]. Similar results were observed in glial cultures from R6/1 mice [52]. Trehalose is found naturally in honey, mushrooms, lobsters, shrimps, wine, and other edible products.

Canthin-6-one is a natural derived alkaloid that can be isolated by many herbs including the edible plant *Aerva lanata*. In PD cellular models (PC12 cells), canthin-6-one promoted α -synuclein degradation through the UPS. Specifically, this alkaloid promoted the expression of *Psm1* gene by activating PKA, subsequently favoring the UPS activation [54].

Salidroside is a glycoside with antiaging, antioxidant, and neuroprotective properties. In SH-SY5Y cells treated with 6-hydroxydopamine (6-OHDA), salidroside alleviated pSer129- α -synuclein burden. Additionally, in the same PD model salidroside elevated the protein levels of PARKIN and UCH-L1 which are two of the most involved constituents of UPS in PD. Free ubiquitin protein levels were also increased due to salidroside treatment along with the 20S proteasome activity (CT-L activity). Moreover, treatment with salidroside enhanced 20S proteasome activity in SH-SY5Y cells transfected with A30P α -synuclein [55]. The herb *Rhodiola rosea* is the main source of salidroside. *Rhodiola rosea* is consumed in the form of tea, made from the roots of the plant and it has been used as an antiaging herb in traditional Chinese medicine.

Quercetin is a flavonoid found in many edible products like green tea, onions, and red wine.

Many studies reveal the positive outcome of quercetin in AD in vivo and in vitro [56]. In CL2006 transgenic strain of *C. elegans*, quercetin was able to reduce A β aggregates and the downstream paralysis phenotype. Among the defense mechanisms, quercetin promoted the UPS activation. Restriction of *uba-1* and *ubq-1* expression hindered the protective effects of quercetin against paralysis. The same research group observed enhanced proteasome activity in CL2006 animals treated with quercetin [56]. Proteasome activation by quercetin was also observed in APP695-transfected SH-SY5Y cells [57]. The same action was also revealed for **rutin**, a flavonoid mostly derived from citrus fruits that has the potential to prevent A β aggregation [97]. Both quercetin and rutin target agents related to proteasome activation, like Nrf2 [98]. Finally, in an HD cell line model, quercetin eliminated mHTT aggregates through proteasome activity enhancement [59].

Sulforaphane is an organic compound mainly found in many dietary products like onions, broccoli, cherries, and red grapes as well as in edible products like green tea and red wine. Through its strong antioxidant activity, sulforaphane exerts protective effects against neurodegenerative diseases [99, 100]. In murine neuroblastoma cells, sulforaphane enhanced proteasome activity via the Nrf2 transcription factor leading to elevated expression of proteasome subunits and thus to resistance against oxidative stress [25]. In an HD cellular model (mHTT-transfected HEK293 cells) treatment with sulforaphane enhanced mHTT degradation by the proteasome [60]. Additionally, sulforaphane was also able to induce proteasome activation in fibroblasts derived from patients with HGPS and improved the growth rate of these cells. In the same study, increased levels of HSP27 protein were also reported; sulforaphane has been shown to enhance proteasome activity through upregulation of HSP27 [61, 101].

As mentioned above polyphenols exhibit antioxidant and anti-aggregation activities. Interestingly, a recent study revealed the implication of **polyphenol digested metabolites** (PDMs) in PD [63]. PDMs from leaves and fruits of *Arbutus*

unedo limited α -synuclein aggregates in a yeast model of PD through proteostasis network modulation, including proteasome elevated activity. Moreover, PDMs influenced the transcription levels of HRD-1, an ubiquitin ligase which is induced by unfolded protein response (UPR). HRD-1 recognizes and ubiquitinates misfolded proteins in ER favoring their degradation through the proteasome indicating a cross talk between the UPR and the UPS in the clearance of α -synuclein by PDMs.

Many phenolic acids and flavonoid compounds such as myricetin, quercetin, and kaempferol along with terpenes attribute to **Ginkgo biloba extract** strong antioxidant properties [102]. Treatment of mHTT-transfected HEK293 cells with Ginkgo biloba extract EGb761 was able to decrease the amount of poly-Q proteins. Accordingly, it was reported that EGb761 promotes proteasome activation and elevated transcript levels of *PSMB5*, *PSMB6*, and *PSMB7* subunits [64].

Puerarin is mainly isolated from *Pueraria lobata* and belongs to the isoflavone family. Beverages made by roots of *Pueraria lobata* contain a small number of isoflavones including puerarin. Many studies reveal the positive effects of puerarin in neuronal survival and cytoprotection in PD organismal and cellular models [103, 104]. In 1-methyl-4-phenylpyridinium (MPP⁺)-treated SH-SY5Y cells, treatment with puerarin increased CT-L activity and lowered the amount of α -synuclein and ubiquitin-conjugated proteins, suggesting the implication of the UPS in puerarin positive results in PD [65].

18 α -glycyrrhetic acid (18 α -GA) is a triterpenoid isolated from *Glycyrrhiza radix*. We have shown that 18 α -GA enhanced proteasome activity and function through elevated protein expression of PBS-5, PAS-1-7 and RPT-6 subunits that are orthologs of *PSMB5*, α -subunits and Rpt-6 proteasome subunits, respectively in *C. elegans*. In the same study, 18 α -GA was able to promote deceleration of the paralysis phenotype in a proteasome-dependent manner in nematode models of AD [66]. 18 α -GA has been shown to enhance

proteasome activity through the induction of Nrf2 transcription factor in human primary cells or its ortholog, namely SKN-1, in *C. elegans* [66, 67]. Finally, in the same study 18 α -GA reduced A β toxicity in SH-SY5Y cells while parallel proteasome inhibition abolished this effect.

Ganoderic acid and **polysaccharide 5** isolated from Ganoderma mushrooms and the plant *Rubia cordifolia* L., respectively, were shown to induce A β clearance through the proteasome in T-REx293 embryonic kidney cell line expressing A β 42-EGFP [68].

Finally, there are two natural compounds that were shown to possess anti-aggregation properties and have also been shown to modulate the UPS, but the association, if there is any, between their potency to act as proteasome activators and their contribution in protection against aggregate accumulation has not been studied yet. **3H-1,2dithiole-3-thione** (D3T) can be found in cruciferous vegetables. In a recent study, treatment of a mouse model of AD (Tg2576) with D3T markedly decreased the levels of insoluble A β 40-42 and enhanced the expression of Nrf2 [105]. Interestingly, D3T has been previously revealed to elevate the 26S proteasome activation and increase the expression of multiple proteasomal subunits in an Nrf2-dependent manner [69]. Whether the Nrf2-mediated proteasome activation is responsible for the protection against A β -toxicity remains to be elucidated. Last but not least, **betulinic acid** is a triterpenoid present in a plethora of plants and has been studied for its neuroprotective effects [106]. Betulinic acid has been reported as stimulator that enhances 20S proteasome activity [70]; the link between betulinic acid action as proteasome stimulator and the neuroprotection that it exerts is still not investigated.

15.5.2 Aging

Oleuropein is a phenolic compound found in high amounts in green olives, olive leaves, and EVOO. Oleuropein effectively exerted antiaging activity and enhanced proteasome function suggesting for the first time that proteasome

activators are interesting targets for the development of antiaging agents. More specifically oleuropein extended the life span of IMR90 and W138 human primary embryonic fibroblasts and increased CT-L, C-L, and T-L activities. Potential conformational changes of the proteasome structure by oleuropein were suggested to be responsible for the observed proteasome enhancement [71].

Quercetin is not only efficient against age-related disorders, but it positively affects the process of aging. Quercetin has been shown to prolong the life span of *S. cerevisiae* and *C. elegans* [107]. Constant treatment of HFL-1 cells with quercetin increased CT-L proteasome activity and proteasome quantity and delayed the senescence phenotype [58].

Lipoic acid is an organosulfur compound that is naturally synthesized by animals and is an essential cofactor in oxidative metabolism [108]. Nevertheless, lipoic acid can also be found in common ingredients of our diet like broccoli, spinach, tomatoes, and brussels sprouts. Lipoic acid is a strong antioxidant acting as a radical scavenger. With regard to its antiaging properties, α -lipoic acid has been shown to increase life span in *C. elegans* and *D. melanogaster* [109–111]. In normal human dermal fibroblasts (NHDF), treatment with lipoic acid positively affected the 20S proteasome activity and upregulated the protein levels of PSMB5 subunit leading to attenuation of oxidative damage and enhanced cell proliferation [72].

Sulforaphane is an isothiocyanate and its protective effects against neurodegeneration and aging have attracted the interest of many researchers [112]. Sulforaphane exerts its protective effects through interaction with the Nrf2 pathway [113–115]. Sulforaphane treatment maintained self-renewal and polypotency and delayed differentiation of human embryonic stem cells through Nrf2 activation and elevated proteasome activity [62].

Bee pollen is widely used and consumed by humans since ancient times [116]. Due to its composition (mixture of polyphenolic compounds) bee pollen has been shown to protect cells through antioxidant and anti-inflammatory

mechanisms [117, 118]. Treatment of HFL-1 cells with bee pollen was shown to trigger the proteasome activity and to elevate protein expression levels of $\beta 2$ and $\beta 5$ subunits [73].

18 α -GA has been shown to extend the life span of treated human fibroblasts and to positively affect proteasome activity in an Nrf2-dependent process [67]. Furthermore, increased life span and enhanced CT-L and T-L activities were also observed in wild-type *C. elegans* upon 18 α -GA administration [66].

Finally, it is known that **zinc** is an essential trace element involved in the maintenance of many homeostatic mechanisms and its deficiency has been associated with aging [119, 120]. In a population study, zinc administration to the elderly increased CT-L proteasome activity and reduced proteotoxicity [74].

15.6 Conclusions

Aging is a natural, unavoidable process that affects almost all organisms. UPS dysfunction is considered as one of the hallmarks of aging and is reported in many age-related pathologies [121]. An important number of investigations reveals the contribution of diet on aging. Diets that are high in fish, fresh fruit and vegetable consumption, like the Mediterranean-type Diet, have been suggested to favor healthy aging and to lower the risk of age-related pathologies. The abovementioned dietary products are rich in phytochemicals which are known for their antioxidant properties and their positive effects on aging and many pathologies including neurodegeneration [122]. Nevertheless, clinical trials and populational studies are missing for most of these compounds. Another pitfall is the fact that all experimental procedures are performed using isolated compounds usually in the μM range. Nevertheless, these concentrations are often unreachable through diet. On top of that, other constituents of dietary extracts may also have adverse effects that can dampen the beneficial properties of a single compound. Therefore, more detailed studies are needed. Given that the life span and the population of older adults have been ele-

vated, along with the incidence of age-related disorders, consumption of dietary products with potent UPS activation properties may represent a promising strategy for health span extension and improvement.

Acknowledgments We would like to thank Mrs. Nikoletta Papaevgeniou and Mary A. Vasilopoulou for comments on the manuscript. Part of the research presented here from our lab has been cofinanced by the European Union and Greek national funds through the Operational Program Competitiveness, Entrepreneurship and Innovation under the call RESEARCH—CREATE—INNOVATE (project codes: T1EDK-00353 and T1EDK-01610). NC lab is also cofinanced under the Action “Action for the Strategic Development on the Research and Technological Sector” (project STHENOS-b, MIS 5002398).

References

1. Ciechanover A (2017) Intracellular protein degradation: from a vague idea thru the lysosome and the ubiquitin-proteasome system and onto human diseases and drug targeting. *Best Pract Res Clin Haematol* 30(4):341–355
2. Dikic I (2017) Proteasomal and autophagic degradation systems. *Annu Rev Biochem* 86:193–224
3. Karademir B, Bozaykut P, Kartal Ozer N (2014) Heat shock proteins and proteasomal degradation in normal and tumor cells. *Free Radic Biol Med* 75(Suppl 1):S35
4. Shiber A, Ravid T (2014) Chaperoning proteins for destruction: diverse roles of Hsp70 chaperones and their co-chaperones in targeting misfolded proteins to the proteasome. *Biomolecules* 4(3):704–724
5. Lopez-Otin C, Blasco MA, Partridge L, Serrano M, Kroemer G (2013) The hallmarks of aging. *Cell* 153(6):1194–1217
6. Hipp MS, Kasturi P, Hartl FU (2019) The proteostasis network and its decline in ageing. *Nat Rev Mol Cell Biol* 20(7):421–435
7. Powers ET, Morimoto RI, Dillin A, Kelly JW, Balch WE (2009) Biological and chemical approaches to diseases of proteostasis deficiency. *Annu Rev Biochem* 78:959–991
8. Wedel S, Manola M, Cavinato M, Trougakos IP, Jansen-Durr P (2018) Targeting protein quality control mechanisms by natural products to promote healthy ageing. *Molecules* 23(5)
9. Papaevgeniou N, Chondrogianni N (2018) Anti-aging and anti-aggregation properties of polyphenolic compounds in *C. elegans*. *Curr Pharm Des* 24(19):2107–2120

10. Winkel-Shirley B (2002) Biosynthesis of flavonoids and effects of stress. *Curr Opin Plant Biol* 5 (3):218–223
11. Chondrogianni N, Voutetakis K, Kapetanou M, Delitsikou V, Papaevgeniou N, Sakellari M et al (2015) Proteasome activation: an innovative promising approach for delaying aging and retarding age-related diseases. *Ageing Res Rev* 23(Pt A):37–55
12. Lefaki M, Papaevgeniou N, Chondrogianni N (2017) Redox regulation of proteasome function. *Redox Biol* 13:452–458
13. Papaevgeniou N, Chondrogianni N (2016) UPS activation in the battle against aging and aggregation-related diseases: an extended review. *Methods Mol Biol* 1449:1–70
14. Njomen E, Tepe JJ (2019) Proteasome activation as a new therapeutic approach to target proteotoxic disorders. *J Med Chem* 62(14):6469–6481
15. Voges D, Zwickl P, Baumeister W (1999) The 26S proteasome: a molecular machine designed for controlled proteolysis. *Annu Rev Biochem* 68:1015–1068
16. Davies KJ (2001) Degradation of oxidized proteins by the 20S proteasome. *Biochimie* 83(3-4):301–310
17. Ciechanover A, Stanhill A (2014) The complexity of recognition of ubiquitinated substrates by the 26S proteasome. *Biochim Biophys Acta* 1843(1):86–96
18. Goldberg AL (2007) Functions of the proteasome: from protein degradation and immune surveillance to cancer therapy. *Biochem Soc Trans* 35(Pt 1):12–17
19. Verma R, Aravind L, Oania R, McDonald WH, Yates JR III, Koonin EV et al (2002) Role of Rpn11 metalloprotease in deubiquitination and degradation by the 26S proteasome. *Science* 298(5593):611–615
20. Groll M, Bochtler M, Brandstetter H, Clausen T, Huber R (2005) Molecular machines for protein degradation. *ChemBiochem* 6(2):222–256
21. Lee MJ, Lee BH, Hanna J, King RW, Finley D (2011) Trimming of ubiquitin chains by proteasome-associated deubiquitinating enzymes. *Mol Cell Proteomics* 10(5):R110.003871
22. Hershko A, Ciechanover A (1998) The ubiquitin system. *Annu Rev Biochem* 67:425–479
23. Nguyen T, Yang CS, Pickett CB (2004) The pathways and molecular mechanisms regulating Nrf2 activation in response to chemical stress. *Free Radic Biol Med* 37(4):433–441
24. Chapple SJ, Siow RC, Mann GE (2012) Crosstalk between Nrf2 and the proteasome: therapeutic potential of Nrf2 inducers in vascular disease and aging. *Int J Biochem Cell Biol* 44(8):1315–1320
25. Kwak MK, Cho JM, Huang B, Shin S, Kensler TW (2007) Role of increased expression of the proteasome in the protective effects of sulforaphane against hydrogen peroxide-mediated cytotoxicity in murine neuroblastoma cells. *Free Radic Biol Med* 43(5):809–817
26. Chondrogianni N, Tzavelas C, Pemberton AJ, Nezis IP, Rivett AJ, Gonos ES (2005) Overexpression of proteasome beta5 assembled subunit increases the amount of proteasome and confers ameliorated response to oxidative stress and higher survival rates. *J Biol Chem* 280(12):11840–11850
27. Chondrogianni N, Gonos ES (2007) Overexpression of hUMP1/POMP proteasome accessory protein enhances proteasome-mediated antioxidant defence. *Exp Gerontol* 42(9):899–903
28. VerPlank JJS, Lokireddy S, Zhao J, Goldberg AL (2019) 26S Proteasomes are rapidly activated by diverse hormones and physiological states that raise cAMP and cause Rpn6 phosphorylation. *Proc Natl Acad Sci U S A* 116(10):4228–4237
29. Lokireddy S, Kukushkin NV, Goldberg AL (2015) cAMP-induced phosphorylation of 26S proteasomes on Rpn6/PSMD11 enhances their activity and the degradation of misfolded proteins. *Proc Natl Acad Sci U S A* 112(52):E7176–E7185
30. Lee BH, Lee MJ, Park S, Oh DC, Elsasser S, Chen PC et al (2010) Enhancement of proteasome activity by a small-molecule inhibitor of USP14. *Nature* 467 (7312):179–184
31. Koga H, Kaushik S, Cuervo AM (2011) Protein homeostasis and aging: the importance of exquisite quality control. *Ageing Res Rev* 10(2):205–215
32. Chondrogianni N, Sakellari M, Lefaki M, Papaevgeniou N, Gonos ES (2014) Proteasome activation delays aging in vitro and in vivo. *Free Radic Biol Med* 71:303–320
33. Small DH, McLean CA (1999) Alzheimer's disease and the amyloid beta protein: What is the role of amyloid? *J Neurochem* 73(2):443–449
34. Tseng BP, Green KN, Chan JL, Blurton-Jones M, LaFerla FM (2008) Abeta inhibits the proteasome and enhances amyloid and tau accumulation. *Neurobiol Aging* 29(11):1607–1618
35. Kaneko M, Koike H, Saito R, Kitamura Y, Okuma Y, Nomura Y (2010) Loss of HRD1-mediated protein degradation causes amyloid precursor protein accumulation and amyloid-beta generation. *J Neurosci Off J Soc Neurosci* 30(11):3924–3932
36. Tramutola A, Di Domenico F, Barone E, Perluigi M, Butterfield DA (2016) It is all about (U)biqutin: role of altered ubiquitin-proteasome system and UCHL1 in Alzheimer disease. *Oxidative Med Cell Longev* 2016:2756068
37. Myers RH (2004) Huntington's disease genetics. *NeuroRx* 1(2):255–262
38. Ortega Z, Lucas JJ (2014) Ubiquitin-proteasome system involvement in Huntington's disease. *Front Mol Neurosci* 7:77
39. Diaz-Hernandez M, Valera AG, Moran MA, Gomez-Ramos P, Alvarez-Castelao B, Castano JG et al (2006) Inhibition of 26S proteasome activity by huntingtin filaments but not inclusion bodies isolated from mouse and human brain. *J Neurochem* 98 (5):1585–1596

40. Spillantini MG, Schmidt ML, Lee VM, Trojanowski JQ, Jakes R, Goedert M (1997) Alpha-synuclein in Lewy bodies. *Nature* 388(6645):839–840
41. Tanaka Y, Engender S, Igarashi S, Rao RK, Wanner T, Tanzi RE et al (2001) Inducible expression of mutant alpha-synuclein decreases proteasome activity and increases sensitivity to mitochondria-dependent apoptosis. *Hum Mol Genet* 10(9):919–926
42. Um JW, Im E, Lee HJ, Min B, Yoo L, Yoo J et al (2010) Parkin directly modulates 26S proteasome activity. *J Neurosci Off J Soc Neurosci* 30(35):11805–11814
43. Giasson BI, Lee VM (2001) Parkin and the molecular pathways of Parkinson's disease. *Neuron* 31(6):885–888
44. Dawson TM, Dawson VL (2010) The role of parkin in familial and sporadic Parkinson's disease. *Mov Disord* 25(Suppl 1):S32–S39
45. Stewart CL, Kozlov S, Fong LG, Young SG (2007) Mouse models of the laminopathies. *Exp Cell Res* 313(10):2144–2156
46. Boasquiuiv PF, Silva GMM, Paiva FA, Cavalcanti RM, Nunez CV, de Paula Oliveira R (2018) Guarana (*Paullinia cupana*) extract protects *Caenorhabditis elegans* models for Alzheimer disease and huntington disease through activation of antioxidant and protein degradation pathways. *Oxidative Med Cell Longev* 2018:9241308
47. Giusti L, Angeloni C, Barbalace MC, Lacerenza S, Ciregia F, Ronci M et al (2018) A proteomic approach to uncover neuroprotective mechanisms of oleocanthal against oxidative stress. *Int J Mol Sci* 19(8)
48. Carmona V, Martin-Aragon S, Goldberg J, Schubert D, Bermejo-Bescos P (2018) Several targets involved in Alzheimer's disease amyloidogenesis are affected by morin and isoquercitrin. *Nutr Neurosci*:1–16 (Epub ahead of print)
49. Marambaud P, Zhao H, Davies P (2005) Resveratrol promotes clearance of Alzheimer's disease amyloid-beta peptides. *J Biol Chem* 280(45):37377–37382
50. Regitz C, Fitzenberger E, Mahn FL, Dussling LM, Wenzel U (2016) Resveratrol reduces amyloid-beta (A β (1)(-)(4)(2))-induced paralysis through targeting proteostasis in an Alzheimer model of *Caenorhabditis elegans*. *Eur J Nutr* 55(2):741–747
51. Corpas R, Grinan-Ferre C, Rodriguez-Farre E, Pallas M, Sanfeliu C (2019) Resveratrol induces brain resilience against Alzheimer neurodegeneration through proteostasis enhancement. *Mol Neurobiol* 56(2):1502–1516
52. Perucho J, Gomez A, Munoz MP, de Yebenes JG, Mena MA, Casarejos MJ (2016) Trehalose rescues glial cell dysfunction in striatal cultures from HD R6/1 mice at early postnatal development. *Mol Cell Neurosci* 74:128–145
53. Fernandez-Estevez MA, Casarejos MJ, Lopez Sendon J, Garcia Caldentey J, Ruiz C, Gomez A et al (2014) Trehalose reverses cell malfunction in fibroblasts from normal and Huntington's disease patients caused by proteasome inhibition. *PLoS One* 9(2):e90202
54. Yuan NN, Cai CZ, Wu MY, Zhu Q, Su H, Li M et al (2019) Canthin-6-one accelerates alpha-synuclein degradation by enhancing ups activity: drug target identification by CRISPR-Cas9 whole genome-wide screening technology. *Front Pharmacol* 10:16
55. Li T, Feng Y, Yang R, Wu L, Li R, Huang L et al (2018) Salidroside promotes the pathological alpha-synuclein clearance through ubiquitin-proteasome system in SH-SY5Y cells. *Front Pharmacol* 9:377
56. Regitz C, Dussling LM, Wenzel U (2014) Amyloid-beta (A β (1)(-)(4)(2))-induced paralysis in *Caenorhabditis elegans* is inhibited by the polyphenol quercetin through activation of protein degradation pathways. *Mol Nutr Food Res* 58(10):1931–1940
57. Martin-Aragon S, Jimenez-Aliaga KL, Benedi J, Bermejo-Bescos P (2016) Neurohormetic responses of quercetin and rutin in a cell line over-expressing the amyloid precursor protein (APP^{swe} cells). *Phytomedicine* 23(12):1285–1294
58. Chondrogianni N, Kapeta S, Chinou I, Vassilatou K, Papassideri I, Gonos ES (2010) Anti-ageing and rejuvenating effects of quercetin. *Exp Gerontol* 45(10):763–771
59. Chakraborty J, Rajamma U, Jana N, Mohanakumar KP (2015) Quercetin improves the activity of the ubiquitin-proteasomal system in 150Q mutated huntingtin-expressing cells but exerts detrimental effects on neuronal survivability. *J Neurosci Res* 93(10):1581–1591
60. Liu Y, Hettlinger CL, Zhang D, Rezvani K, Wang X, Wang H (2014) Sulforaphane enhances proteasomal and autophagic activities in mice and is a potential therapeutic reagent for Huntington's disease. *J Neurochem* 129(3):539–547
61. Gabriel D, Roedel D, Gordon LB, Djabali K (2015) Sulforaphane enhances progerin clearance in Hutchinson-Gilford progeria fibroblasts. *Aging Cell* 14(1):78–91
62. Jang J, Wang Y, Kim HS, Lalli MA, Kosik KS (2014) Nrf2, a regulator of the proteasome, controls self-renewal and pluripotency in human embryonic stem cells. *Stem Cells* 32(10):2616–2625
63. Macedo D, Jardim C, Figueira I, Almeida AF, McDougall GJ, Stewart D et al (2018) (Poly)phenol-digested metabolites modulate alpha-synuclein toxicity by regulating proteostasis. *Sci Rep* 8(1):6965
64. Stark M, Behl C (2014) The Ginkgo biloba Extract EGb 761 Modulates Proteasome Activity and Polyglutamine Protein Aggregation. *Evid Based Complement Alternat Med* 2014:940186
65. Cheng YF, Zhu GQ, Wang M, Cheng H, Zhou A, Wang N et al (2009) Involvement of ubiquitin proteasome system in protective mechanisms of Puerarin to MPP (+)-elicited apoptosis. *Neurosci Res* 63(1):52–58
66. Papaevgeniou N, Sakellari M, Jha S, Tavernarakis N, Holmberg CI, Gonos ES et al (2016) 18alpha-glycyrrhetic acid proteasome activator decelerates aging and Alzheimer's disease progression in

- Caenorhabditis elegans* and neuronal cultures. *Antioxid Redox Signal* 25(16):855–869
67. Kapeta S, Chondrogianni N, Gonos ES (2010) Nuclear erythroid factor 2-mediated proteasome activation delays senescence in human fibroblasts. *J Biol Chem* 285(11):8171–8184
 68. Chakrabortee S, Liu Y, Zhang L, Matthews HR, Zhang H, Pan N et al (2012) Macromolecular and small-molecule modulation of intracellular Abeta42 aggregation and associated toxicity. *Biochem J* 442(3):507–515
 69. Kwak MK, Wakabayashi N, Greenlaw JL, Yamamoto M, Kensler TW (2003) Antioxidants enhance mammalian proteasome expression through the Keap1-Nrf2 signaling pathway. *Mol Cell Biol* 23(23):8786–8794
 70. Coleman RA, Trader DJ (2018) Development and application of a sensitive peptide reporter to discover 20S proteasome stimulators. *ACS Comb Sci* 20(5):269–276
 71. Katsiki M, Chondrogianni N, Chinou I, Rivett AJ, Gonos ES (2007) The olive constituent oleuropein exhibits proteasome stimulatory properties in vitro and confers life span extension of human embryonic fibroblasts. *Rejuvenation Res* 10(2):157–172
 72. Sikdar S, Papadopoulou M, Dubois J (2017) Effect of -lipoic acid on proteasomal induction: protection against oxidative damage in human skin fibroblasts cell line NHDF. *Pharmacol Pharm* 08(09):292–305
 73. Graikou K, Kapeta S, Aligiannis N, Sotiroidis G, Chondrogianni N, Gonos E et al (2011) Chemical analysis of Greek pollen – antioxidant, antimicrobial and proteasome activation properties. *Chem Cent J* 5(1):33
 74. Cabreiro F, Perichon M, Jatje J, Malavolta M, Mocchegiani E, Friguet B et al (2008) Zinc supplementation in the elderly subjects: effect on oxidized protein degradation and repair systems in peripheral blood lymphocytes. *Exp Gerontol* 43(5):483–487
 75. Leite RP, Wada RS, Monteiro JC, Predes FS, Dolder H (2011) Protective effect of Guarana (*Paullinia cupana* var. *sorbilis*) pre-treatment on cadmium-induced damages in adult Wistar testis. *Biol Trace Elem Res* 141(1-3):262–274
 76. Bittencourt Lda S, Zeidan-Chulia F, Yatsu FK, Schnorr CE, Moresco KS, Kolling EA et al (2014) Guarana (*Paullinia cupana* Mart.) prevents beta-amyloid aggregation, generation of advanced glycation-end products (AGEs), and acrolein-induced cytotoxicity on human neuronal-like cells. *Phytother Res* 28(11):1615–1624
 77. Peixoto H, Roxo M, Rohrig T, Richling E, Wang X, Wink M (2017) Anti-aging and antioxidant potential of *Paullinia cupana* var. *sorbilis*: findings in *Caenorhabditis elegans* indicate a new utilization for roasted seeds of guarana. *Medicines* 4(3)
 78. Korre M, Tsoukas MA, Frantzeskou E, Yang J, Kales SN (2014) Mediterranean diet and workplace health promotion. *Curr Cardiovasc Risk Rep* 8(12):416
 79. Sabatini N, Perri E, Rongai D (2018) Olive oil antioxidants and aging. *Food quality: balancing health and disease. Handb Food Bioeng* (13):145–157
 80. Rigacci S (2015) Olive oil phenols as promising multi-targeting agents against Alzheimer's disease. *Adv Exp Med Biol* 863:1–20
 81. Li W, Sperry JB, Crowe A, Trojanowski JQ, Smith AB III, Lee VM (2009) Inhibition of tau fibrillization by oleocanthal via reaction with the amino groups of tau. *J Neurochem* 110(4):1339–1351
 82. Abuznait AH, Qosa H, Busnena BA, El Sayed KA, Kaddoumi A (2013) Olive-oil-derived oleocanthal enhances beta-amyloid clearance as a potential neuroprotective mechanism against Alzheimer's disease: in vitro and in vivo studies. *ACS Chem Neurosci* 4(6):973–982
 83. Batarseh YS, Mohamed LA, Al Rihani SB, Mousa YM, Siddique AB, El Sayed KA et al (2017) Oleocanthal ameliorates amyloid-beta oligomers' toxicity on astrocytes and neuronal cells: in vitro studies. *Neuroscience* 352:204–215
 84. Theodoraki MA, Caplan AJ (2012) Quality control and fate determination of Hsp90 client proteins. *Biochim Biophys Acta* 1823(3):683–688
 85. Imai J, Maruya M, Yashiroda H, Yahara I, Tanaka K (2003) The molecular chaperone Hsp90 plays a role in the assembly and maintenance of the 26S proteasome. *EMBO J* 22(14):3557–3567
 86. Alberdi E, Sanchez-Gomez MV, Ruiz A, Cavaliere F, Ortiz-Sanz C, Quintela-Lopez T et al (2018) Mangiferin and morin attenuate oxidative stress, mitochondrial dysfunction, and neurocytotoxicity, induced by amyloid beta oligomers. *Oxidative Med Cell Longev* 2018:2856063
 87. Kataria R, Sobarzo-Sanchez E, Khatkar A (2018) Role of morin in neurodegenerative diseases: a review. *Curr Top Med Chem* 18(11):901–907
 88. Malhotra A, Bath S, Elbarbry F (2015) An organ system approach to explore the antioxidative, anti-inflammatory, and cytoprotective actions of resveratrol. *Oxidative Med Cell Longev* 2015:803971
 89. Sun AY, Wang Q, Simonyi A, Sun GY (2010) Resveratrol as a therapeutic agent for neurodegenerative diseases. *Mol Neurobiol* 41(2-3):375–383
 90. Tellone E, Galtieri A, Russo A, Giardina B, Ficarra S (2015) Resveratrol: a focus on several neurodegenerative diseases. *Oxidative Med Cell Longev* 2015:392169
 91. Link CD (1995) Expression of human beta-amyloid peptide in transgenic *Caenorhabditis elegans*. *Proc Natl Acad Sci U S A* 92(20):9368–9372
 92. Mohar DS, Malik S (2012) The sirtuin system: the holy grail of resveratrol? *J Clin Exp Cardiol* 3(11)
 93. Burns J, Yokota T, Ashihara H, Lean ME, Crozier A (2002) Plant foods and herbal sources of resveratrol. *J Agric Food Chem* 50(11):3337–3340

94. Lee HJ, Yoon YS, Lee SJ (2018) Mechanism of neuroprotection by trehalose: controversy surrounding autophagy induction. *Cell Death Dis* 9(7):712
95. Seo Y, Kingsley S, Walker G, Mondoux MA, Tissenbaum HA (2018) Metabolic shift from glycogen to trehalose promotes lifespan and healthspan in *Caenorhabditis elegans*. *Proc Natl Acad Sci U S A* 115(12):E2791–EE800
96. Tanaka M, Machida Y, Niu S, Ikeda T, Jana NR, Doi H et al (2004) Trehalose alleviates polyglutamine-mediated pathology in a mouse model of Huntington disease. *Nat Med* 10(2):148–154
97. Jimenez-Aliaga K, Bermejo-Bescos P, Benedi J, Martin-Aragon S (2011) Quercetin and rutin exhibit anti-amyloidogenic and fibril-disaggregating effects in vitro and potent antioxidant activity in APPsw cells. *Life Sci* 89(25–26):939–945
98. Arredondo F, Echeverry C, Abin-Carriquiry JA, Blasina F, Antunez K, Jones DP et al (2010) After cellular internalization, quercetin causes Nrf2 nuclear translocation, increases glutathione levels, and prevents neuronal death against an oxidative insult. *Free Radic Biol Med* 49(5):738–747
99. Morroni F, Tarozzi A, Sita G, Bolondi C, Zolezzi Moraga JM, Cantelli-Forti G et al (2013) Neuroprotective effect of sulforaphane in 6-hydroxydopamine-lesioned mouse model of Parkinson's disease. *Neurotoxicology* 36:63–71
100. Kim HV, Kim HY, Ehrlich HY, Choi SY, Kim DJ, Kim Y (2013) Amelioration of Alzheimer's disease by neuroprotective effect of sulforaphane in animal model. *Amyloid* 20(1):7–12
101. Gan N, Wu YC, Brunet M, Garrido C, Chung FL, Dai C et al (2010) Sulforaphane activates heat shock response and enhances proteasome activity through up-regulation of Hsp27. *J Biol Chem* 285(46):35528–35536
102. Horakova L, Licht A, Sandig G, Jakstadt M, Durackova Z, Grune T (2003) Standardized extracts of flavonoids increase the viability of PC12 cells treated with hydrogen peroxide: effects on oxidative injury. *Arch Toxicol* 77(1):22–29
103. Shiyong L, Xinhui Q, Guanghua J, Feng N, Feng L, Shumei C et al (2018) Puerarin promoted proliferation and differentiation of dopamine-producing cells in Parkinson's animal models. *Biomed Pharmacother* 106:1236–1242
104. Zhang X, Xiong J, Liu S, Wang L, Huang J, Liu L et al (2014) Puerarin protects dopaminergic neurons in Parkinson's disease models. *Neuroscience* 280:88–98
105. Cui Y, Ma S, Zhang C, Li D, Yang B, Lv P et al (2018) Pharmacological activation of the Nrf2 pathway by 3H-1, 2-dithiole-3-thione is neuroprotective in a mouse model of Alzheimer disease. *Behav Brain Res* 336:219–226
106. Navabi SP, Sarkaki A, Mansouri E, Badavi M, Ghadiri A, Farbood Y (2018) The effects of betulinic acid on neurobehavioral activity, electrophysiology and histological changes in an animal model of the Alzheimer's disease. *Behav Brain Res* 337:99–106
107. Belinha I, Amorim MA, Rodrigues P, de Freitas V, Moradas-Ferreira P, Mateus N et al (2007) Quercetin increases oxidative stress resistance and longevity in *Saccharomyces cerevisiae*. *J Agric Food Chem* 55(6):2446–2451
108. Reed LJ, De BB, Gunsalus IC, Hornberger CS Jr (1951) Crystalline alpha-lipoic acid; a catalytic agent associated with pyruvate dehydrogenase. *Science* 114(2952):93–94
109. Rahman K (2007) Studies on free radicals, antioxidants, and co-factors. *Clin Interv Aging* 2(2):219–236
110. Brown MK, Evans JL, Luo Y (2006) Beneficial effects of natural antioxidants EGCG and alpha-lipoic acid on life span and age-dependent behavioral declines in *Caenorhabditis elegans*. *Pharmacol Biochem Behav* 85(3):620–628
111. Bauer JH, Goupil S, Garber GB, Helfand SL (2004) An accelerated assay for the identification of lifespan-extending interventions in *Drosophila melanogaster*. *Proc Natl Acad Sci U S A* 101(35):12980–12985
112. Santin-Marquez R, Alarcon-Aguilar A, Lopez-Diazguerrero NE, Chondrogianni N, Konigsberg M (2019) Sulfoaphane – role in aging and neurodegeneration. *GeroScience* 41(5):655–670
113. Hu C, Eggler AL, Mesecar AD, van Breemen RB (2011) Modification of Keap1 cysteine residues by sulforaphane. *Chem Res Toxicol* 24(4):515–521
114. Pu D, Zhao Y, Chen J, Sun Y, Lv A, Zhu S et al (2018) Protective effects of sulforaphane on cognitive impairments and AD-like lesions in diabetic mice are associated with the upregulation of Nrf2 transcription activity. *Neuroscience* 381:35–45
115. Bai Y, Cui W, Xin Y, Miao X, Barati MT, Zhang C et al (2013) Prevention by sulforaphane of diabetic cardiomyopathy is associated with up-regulation of Nrf2 expression and transcription activation. *J Mol Cell Cardiol* 57:82–95
116. Xi X, Li J, Guo S, Li Y, Xu F, Zheng M et al (2018) The potential of using bee pollen in cosmetics: a review. *J Oleo Sci* 67(9):1071–1082
117. Atsalakis E, Chinou I, Makropoulou M, Karabourmioti S, Graikou K (2017) Evaluation of phenolic compounds in *Cistus creticus* bee pollen from Greece. Antioxidant and antimicrobial properties. *Nat Prod Commun* 12(11):1813–1816
118. Campos MG, Webby RF, Markham KR, Mitchell KA, Da Cunha AP (2003) Age-induced diminution of free radical scavenging capacity in bee pollens and the contribution of constituent flavonoids. *J Agric Food Chem* 51(3):742–745
119. Mocchegiani E, Romeo J, Malavolta M, Costarelli L, Giacconi R, Diaz LE et al (2013) Zinc: dietary intake and impact of supplementation on immune function in elderly. *Age* 35(3):839–860
120. Cabrera AJ (2015) Zinc, aging, and immunosenescence: an overview. *Pathobiol Aging Age Relat Dis* 5:25592

121. Labbadia J, Morimoto RI (2015) The biology of proteostasis in aging and disease. *Annu Rev Biochem* 84:435–464
122. Kumar GP, Khanum F (2012) Neuroprotective potential of phytochemicals. *Pharmacogn Rev* 6(12):81–90
123. Vilchez D, Boyer L, Morante I, Lutz M, Merkwirth C, Joyce D, Spencer B, Page L, Masliah E, Berggren WT, Gage FH, Dillin A (2012) Increased proteasome activity in human embryonic stem cells is regulated by PSMD11. *Nature* 489(7415):304–308
124. Vilchez D, Morante I, Liu Z, Douglas PM, Merkwirth C, Rodrigues APC, Manning G, Dillin A (2012) RPN-6 determines *C. elegans* longevity under proteotoxic stress conditions. *Nature* 489(7415):263–268