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Novel quinolinone-pyrazoline hybrids: synthesis and evaluation of antioxidant and lipoxygenase inhibitory activity

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Abstract

The present project deals with the investigation of structure–activity relationship of several quinolinone–chalcone and quinolinone–pyrazoline hybrids, in an effort to discover promising antioxidant and anti-inflammatory agents. In order to accomplish this goal, four bioactive hybrid quinolinone–chalcone compounds ($\bf 8a-8d$) were synthesized via an aldol condensation reaction, which were then chemically modified, forming fifteen new pyrazoline analogues ($\bf 9a-9o$). All the synthesized analogues were in vitro evaluated in terms of their antioxidant and soybean lipoxygenase (LOX) inhibitory activity. Among all the pyrazoline derivatives, compounds $\bf 9b$ and $\bf 9m$ were found to possess the best combined activity, whereas $\bf 9b$ analogue exhibited the most potent LOX inhibitory activity, with IC₅₀ value 10 μ M. The in silico docking results revealed that the synthetic pyrazoline analogue $\bf 9b$ showed high AutoDock Vina score (-10.3 kcal/mol), while all the tested derivatives presented allosteric interactions with the enzyme.

Graphic Abstract

OHC
$$R_1$$
 OH O R_2 OH O R_2 OH O R_3 $R_3 = H$, $COCH_3$ $R_3 = H$, $COCH_3$ $R_3 = H$, $COCH_3$ $R_4 = H$, $COCH_3$ $R_5 = H$, $COCH_3$ $COCH_2$ $COCH_3$ COC

Keywords Quinolinones · Chalcones · Pyrazolines · Lipoxygenase · LOX inhibition · Antioxidant activity

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Introduction

Inflammation is part of the body's defense mechanism, and it can be beneficial as a healing process, by which the immune system recognizes and removes harmful stimuli. Acute inflammation is induced by tissue damage due to trauma, microbial invasion or noxious compounds. Chronic inflammation is a persistent phenomenon that can last from months to years and plays a central role in some of the most challenging diseases of our time, including neurodegenerative and heart diseases, rheumatoid arthritis, diabetes, asthma, and even cancer.

Lipoxygenases (LOXs) are a heterogeneous family of structurally related non-heme iron-containing enzymes that



catalyze the insertion of molecular oxygen into polyunsaturated fatty acids, such as arachidonic acid or linoleic acid. They are classified as 5-, 8-, 12, and 15- LOXs according to their selectivity to oxygenate fatty acids in a specific position [1–4].

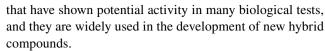
Over the years, several strategies have been developed in order to block the arachidonic acid pathway and different compounds have been identified as 5-LOX inhibitors [5–8]. Up to now, Zileuton is the only approved potent and selective 5-LOX inhibitor for the treatment of asthma [9, 10]. However, several 5-LOX inhibitors are now under clinical investigation for the treatment of cardiovascular diseases and vascular inflammation, atherosclerosis, asthma and for knee osteoarthritis [11–13]. Furthermore, recent studies examine the relationship between arachidonic acid cascade and carcinogenesis, revealing novel targets for the treatment of cancer [14–16]. It has been demonstrated that 5-LOX plays an important role in regulating cellular proliferation and there are 5-LOX inhibitor pharmacophores which can also cause cell death in prostate cancer cells [2].

Chronic inflammation and oxidative stress are two commonly associated conditions involved in the pathophysiology of cancers, diabetes, cardiovascular and pulmonary diseases, and others. Oxidative stress occurs when the balance between antioxidants and reactive oxygen species (ROS) is disrupted because of either depletion of antioxidants or accumulation of ROS.

The main feature of an antioxidant is its ability to bind free radicals, which are produced during the inflammation process by phagocytic leukocytes that invade the tissue. Moreover, ROS are involved in the biosynthesis of prostaglandins and in the cyclooxygenase (COX) and LOX mediated conversion of arachidonic acid into proinflammatory intermediates [17–19]. Therefore, the development of novel drugs that combine anti-inflammatory and antioxidant activity could be beneficial for the treatment of several diseases [20].

Hybrid molecules combine the structural frame of two or more different pharmacophores which have already been exploited in drug development, and they are designed in order to provide novel drugs with enhanced activity [21–23]. Quinolinones and chalcones are two scaffolds

Fig. 1 General structure of the quinolinone (I), chalcone (II) and pyrazoline (III) molecular scaffolds



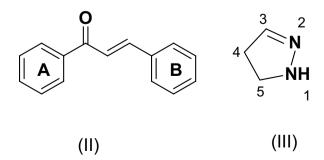
Quinolinones are heterocyclic nitrogen compounds which are mainly found as alkaloids in a variety of natural products, as well as synthetic analogues (Fig. 1) [24, 25]. Furthermore, compounds containing the heterocyclic system of 4-substituted-2-quinolinone as a building block exhibit a wide variety of biological and pharmacological properties and are useful intermediates in the synthesis of a large number of bioactive molecules [26–28].

Chalcones are natural products which have been recognized as the precursors to the biosynthesis of flavonoids and isoflavonoids [29]. They are α,β -unsaturated carbonyl compounds, which are characterized by the presence of a three carbon bridge with a double bond (Fig. 1) [30]. Chalcones and their derivatives consist "privileged structures" and they show many interesting biological properties, such as anti-inflammatory [20, 31, 32], anti-parasitic [33], antimicrobial [34], antibacterial [32, 35], antioxidant [31, 36], cytotoxic [37], anticancer [31, 38] and so on.

Following the above trend, the aim of the present research is the synthesis of bioactive hybrid quinolinone—chalcone compounds, and the chemical modification of the above system, in order to form new pyrazoline analogues (Fig. 1).

Pyrazolines (Fig. 1) are well-known and important fivemembered heterocyclic compounds, possessing two adjacent nitrogen atoms in the ring and only one endocyclic double bond. [39]. Depending on the position of the double bond, there are three possible partially reduced forms of pyrazoline structure, which are namely 1-pyrazoline, 2-pyrazoline or 3-pyrazoline which can exist in equilibrium with each other. Among them, 2-pyrazoline is more stable than the rest of the reduced forms, while it seems to be the most frequently studied form of pyrazoline [40].

Pyrazoline derivatives have been reported to exhibit a wide range of pharmacological activities such as antimicrobial [41–43], anticancer [44, 45], anti-inflammatory [46], antioxidant [47, 48], antidepressant [49] and so on. Out of all these biological activities of pyrazolines and their derivatives, the anti-inflammatory activity seems to concentrate the interest of researchers, while there is a





Scheme 1 Synthesis of 3-acetyl-4-hydroxy-2-quinolinone (3). Reagents and conditions: (i) (CH₃CO)₂O, 130 °C, 2 h (ii) CH₃COCH₂COOEt, t-BuOK, t-BuOH, rt (iii) aq Na₂CO₃/NaOH, rt

COOH (i) O (iii) O COCH₃ (iiii) O COCH₃ (iiii) O COCH₃
$$COOEt$$
 $COCH3$ $COOEt$ $COCH3$ $COCH3$

Scheme 2 Synthesis of 3-acetyl-4-hydroxy-1-methyl-2-quinolinone (6). Reagents and conditions: (i) toluene, 110 °C, 2 h (ii) NaOCH₂CH₃, 77 °C, 2 h

plethora of information available in the literature which is growing steadily over the years [50–56].

In 2019, Stefanes and coworkers synthesized a series of new pyrazoline derivatives, which were then evaluated for their anti-leukemic activity, revealing two of the compounds as promising candidates against acute leukemia [57]. Mumtaz and his group focused on the current need of new potential agents over the treatment of neurodegenerative disorders, such as Parkinson's disease or Alzheimer's disease, synthesizing several derivatives of thioureas and pyrazolines, which were then evaluated via two assays. Results showed one of the pyrazoline derivatives as the most potent acetylcholinesterase inhibitor, with $IC_{50} = 123 \pm 51$ nM [58].

Furthermore, the evaluation of the anti-inflammatory activity of pyrazolines still stands out in the literature, since several corresponding studies were published in 2019. Cai et al. [59] synthesized a number of novel steroidal derivatives bearing pyrazoline structure, among others, identifying one of them as the most potent anti-inflammatory agent, with an IC₅₀ value of 0.86 μM on NO production in LPS induced RAW 264.7 cells. Earlier the same year, Sethiya and his research group synthesized some new pyrazoline derivatives, using ultrasonic irradiation, which demonstrated remarkable anti-inflammatory activity [60]. Chandel et al. [61] synthesized coumarin-based pyrazolines and evaluated them in terms of their in vitro and in vivo anti-inflammatory activity. Experimental results revealed one of the tested compounds as highly active antiinflammatory agent and suggested it to be used as lead compound for the development of effective inhibitors.

With regard to the synthesis of pyrazolines, literature offers a wide variety of methods depending on the reactivity of molecules and the need of the chemist [62]. However, the most popular method is the one of Fischer and Knoevenagel, i.e., the reaction of α , β -unsaturated ketones with phenyl hydrazine in acetic acid under refluxing conditions [63, 64].

Table 1 New quinolinone—chalcone hybrids

Code	Y	R ₁	R ₂
8a	Н	OCH ₃	OCH ₃
8b	Н	OCH_3	OH
8c	CH_3	OCH_3	OCH_3
8d	CH_3	OCH_3	OH

Results and discussion

Chemistry

The desired starting quinolinones differ to the substituent Y attached to the nitrogen of the heterocyclic ring, and they were synthesized through different synthetic routes.

3-Acetyl-4-hydroxy-2-quinolinone (3) was synthesized using our previously developed methodology which includes C-acylation of ethyl acetoacetate by 2-methyl-3,1-benzoxazin-4-one (1) in a basic environment, followed by a cyclization reaction of the C-acylation product 2 in aqueous solution of Na_2CO_3 and NaOH (Scheme 1) [33, 65].

In order to investigate the role of the substituent Y, we synthesized quinolinone 6, which bears a methyl group at this position. The synthesis was accomplished using our previous methodology, via an acylation reaction of secondary amine 4, followed by a cyclization reaction of the acylation product 5, in basic environment (Scheme 2) [33].

The desired four quinolinone-chalcone hybrids (Table 1) were finally synthesized according to Scheme 3, using compounds 3 and 6 as starting materials in an aldol condensation reaction with two different benzaldehydes. It is a methodology that has already been developed in a previous work of our laboratory, [33].

Proceeding with the synthesis and in order to further investigate the structure-activity relationship of the final compounds, we attempted a structural modification, which



Scheme 3 Synthesis of quinolinone-chalcone hybrid compounds and of new pyrazoline analogues. Reagents and conditions: (i) Piperidine, 78 °C, 5–24 h (ii) CH₃COOH/ EtOH, reflux, 120 °C

was carried out to the α,β -unsaturated carbonyl system of compounds **8**, leading to pyrazoline derivatives **9** (Table 2).

Various conditions were investigated in order to synthesize the target pyrazoline derivatives. After optimization of the reaction conditions, we finally synthesized 15 new molecules by refluxing the corresponding quinolinone—chalcone compound (8) with a variety of hydrazine derivatives in glacial acetic acid (Scheme 3).

The structures of all the new pyrazolines were elucidated by $^1\text{H-NMR}, \, ^{13}\text{C-NMR}$ and HR-MS. In the $^1\text{H-NMR}$ of the compounds, all the three protons $H_A,\, H_B$ and H_X attached to the C_4 and C_5 carbon atoms of the pyrazoline ring, respectively, were found to give an ABX spin system. The methylene protons of C_4 resonated as a pair of doublet of doublet peaks at regions 3.43–3.66 ppm ($C_4\text{-H}_A$) and 4.14–4.33 ppm ($C_4\text{-H}_B$). The methine proton of C_5 also appeared as a doublet of doublet peak at the region 4.96–5.48 ppm, due to its vicinal coupling with the two magnetically non-equivalent protons of the position C_4 of the pyrazoline ring. Among the three signals of $H_A,\,H_B$ and H_X protons, the most deshielded one is attributed to the methine of C_5 , due to its close proximity to the nitrogen of the pyrazoline ring and to ring B of the chalcone moiety.

Biology

The new derivatives were tested for their antioxidant activity in vitro based on their capacity to scavenge the stable free radical DPPH, as well as their ability to inhibit lipid peroxidation of linoleic acid induced by AAPH radical. The first one is a fast, simple, cost-effective and widely used method, where the DPPH stable free radical reacts directly with the antioxidant and is decolorized. The scavenging effect of the synthesized compounds on the DPPH radical was evaluated according to the methods of Hadjipavlou et al. [66, 67].

AAPH-induced linoleic acid oxidation has been developed as a quick and reliable method for measuring the antioxidant activity. The thermal free radical producer (AAPH) generates free radicals in the solution which cause the oxidation of linoleic acid, and the method is a measure of how effective antioxidants protect against lipid peroxidation in vitro. Oxidation of exogenous linoleic acid by AAPH is followed by UV spectrophotometry in a highly diluted sample [68].

Furthermore, for the evaluation of their anti-inflammatory activity, all new molecules were tested as inhibitors of soybean LOX, which is a plant enzyme with satisfactory homology with the human 5-LOX, and the results can be considered as an indication of the anti-inflammatory activity of new analogues. The results of this study are presented in Table 3.

Evaluation of antioxidant and soybean LOX inhibitory activity

The results of the DPPH radical scavenging activity of the majority of the tested analogues showed good interaction with the DPPH radical at $100~\mu M$ concentration. In general, the interaction is altered in relation to the different substituents and more specifically in relation to the presence or not



Table 2 New pyrazoline analogues

CODE	Y	R ₁	R ₂	R ₃
9a	Н	OCH ₃	OCH ₃	3"-CN 2-1"-5" 4'''-cyanophenyl
9b	Н	OCH ₃	OCH ₃	4'''-methoxyphenyl
9c	Н	OCH ₃	OCH ₃	2"" 5"" 5"" 4"" 5"" 5"" 4"" 5"" 5"" 4"" 5"" 5
9d	Н	OCH ₃	ОН	3"-CN 2½ 5" 4"'-cyanophenyl
9e	Н	OCH ₃	ОН	3" 4" 0 4" 5" 5" 5" 4" 4" 4" 4" 4" 4" 4" 5" 5" 6" 5" 6" 6" 6" 6" 6" 6" 6" 6" 6" 6" 6" 6" 6"
9f	Н	OCH ₃	ОН	4'''-chlorophenyl
9g	СН3	OCH ₃	OCH ₃	4'''-cyanophenyl
9h	СН3	OCH ₃	OCH ₃	(3''',4'''-dimethyl)phenyl



Table 2 (continued)

9i	CH ₃	OCH_3	OCH ₃	4'''-methoxyphenyl
9j	СН3	OCH ₃	ОН	4'''-cyanophenyl
9k	CH ₃	OCH₃	ОН	(3''',4'''-dimethyl)phenyl
91	CH ₃	OCH ₃	ОН	4'''-methoxyphenyl
9m	СН3	OCH ₃	ОН	2" 4" 0 2" 5" 5" 5" 4" 4" 6" 4" 4" 6" 4" 6" 6" 6" 6" 6" 6" 6" 6" 6" 6" 6" 6" 6"
9n	СН3	OCH ₃	ОН	4'''-chlorophenyl
90	Н	OCH ₃	ОН	2"

of labile protons, such as phenolic hydroxyl groups, which react directly with the free radical.

Among the four synthesized quinolinone–chalcones, it seems that the presence of a phenolic hydroxyl group in compounds **8b** and **8d** leads to good DPPH radical scavengers (81% and 78% after 20-min incubation at a concentration of 0.1 mM, respectively) and LOX inhibitors (IC $_{50}$ 56.0 μM and 50.0 μM , respectively). The similar values of DPPH scavenging and LOX inhibition ability presented

by quinolinone–chalcones **8b** and **8d**, which differ only in the substituent at the heterocyclic N, indicate that the most important structural feature for both activities is the presence of the phenolic OH. Moreover, substitution of the phenolic OH with a OCH₃ group (compounds **8a** and **8c**) leads to complete loss of DPPH and LOX inhibitory activity. On the other hand, the lipid peroxidation ability of compounds **8b** and **8d** is low, but the replacement of the OH by a OCH₃ group (compounds **8a** and **8c**) leads to significant inhibitors



Table 3 In vitro biological evaluation of all the synthesized analogues

CODE	Interaction with the free radical DPPH (%)		Inibition of lipid peroxidation of linoleic acid induced by AAPH radical	Inhibition of soy- bean lipoxygenase
	0.1 mM/20 min	0.1 mM/60 min	(%) 0.1 mM	$IC_{50} (\mu M)$
8a	_	_	100.0	(19.2% at 0.1 mM)
8b	81.0	84.0	23.0	56.0
8c	_	_	100.0	(13.0% at 0.1 mM)
8d	78.0	82.0	31.0	50.0
9a	27.3	12.7	74.7	(27.3% at 0.1 mM)
9b	85.3	86.2	82.9	10.0
9c	86.2	98.9	93.3	(43.8% at 0.1 mM)
9d	95.0	95.3	83.8	52.0
9e	76.0	84.0	88.2	63.0
9f	63.0	71.0	71.4	74.0
9g	99.0	44.0	25.0	65.0
9h	60.0	71.0	92.3	67.5
9i	87.0	91.0	100.0	(16% at 0.1 mM)
9j	74.0	8.0	77.0	15.0
9k	77.0	79.0	87.2	57.0
91	83.0	87.0	97.0	57.5
9m	85.0	100.0	79.0	39.0
9n	67.0	73.0	59.0	50.0
90	74.5	85.5	59.1	90.0
NDGA	88.0	97.0	_	0.5
Trolox	_	_	88.0	_

of lipid peroxidation (100% inhibition at a concentration of 0.1 mM for both compounds) but inactive DPPH scavengers and LOX inhibitors. From the above observations, it can be postulated that in the case of quinolinone–chalcones, LOX inhibition is related to the antioxidant activity as measured by the DPPH scavenging ability and can probably be correlated with their H atom transfer capacity.

An important observation emerging from the evaluation of the antioxidant activity of all the new analogues is related to the 4-hydroxyl group of the quinolinone moiety. This proton creates a strong hydrogen bond with the adjacent carbonyl group, and for this reason it cannot react with the free DPPH radical, so it does not participate in the enhancement of the antioxidant activity of quinolinone-chalcones. The stability of this proton is easily proven by the ¹H NMR analysis, where it appears through a singlet peak at very low fields of the spectrum, 17-18 ppm. In the spectra of pyrazoline analogues this peak is shifted to 13–14 ppm, indicating that the proton in this case creates a weaker hydrogen bond with the nitrogen of the pyrazoline ring. This is a plausible explanation to account for the observation that the majority of the tested pyrazolines show a better DPPH radical scavenger capacity, compared to their quinolinone-chalcone precursors. As a striking example, one can compare the DPPH scavenging ability of pyrazolines 9a, **9b** and **9c**, which were prepared from the same precursor,

quinolinone–chalcone **8a**: **8a** is inactive, whereas **9b** and **9c** are among the most active DPPH scavengers (85.3% and 86.2%, respectively, at a concentration of 0.1 mM and after 20 min incubation) of the series. Pyrazoline **9a** is not a good DPPH scavenger; however, it is not completely inactive as **8a**. The same trend is followed in the case of the very good DPPH scavengers pyrazolines **9g** and **9i** (99% and 87% inhibition, respectively), which are derived from the inactive quinolinone–chalcone **8c**.

In the case of the pyrazoline analogues produced from the good radical scavengers **8b** and **8d**, namely compounds **9d–9f** and **9j–9o**, respectively, the contribution of the OH group at position 4 of the quinolinone heterocyclic ring in the DPPH scavenging ability does not seem to be as significant as the one of the phenolic OH. Thus, the above pyrazolines were found to possess analogous or even less antioxidant activity compared to their precursors **8b** and **8d**.

As far as the ability of the tested pyrazolines to inhibit lipid peroxidation of linoleic acid induced by a thermal free radical producer (AAPH), five pyrazolines, namely **9c**, **9e**, **9h**, **9i** and **9m**, were found to be potent inhibitors showing activity equal or higher than the reference compound Trolox. Insertion of the pyrazoline moiety at quinolinone—chalcone **8b** (with very low lipid peroxidation ability, 23% at 0.1 mM) resulted in compounds **9d**–**9f** which had remarkably higher activity. The same is true for pyrazolines arising from



quinolinone–chalcone **8d**: the parent compound had very low activity (31% at 0.1 mM), whereas the corresponding pyrazolines **9j–9o** showed moderate to high activity (>59%). However, pyrazolines **9a–9c** and **9g–9i** which were derived from **8a** and **8c**, respectively, showed analogous or lower activity with the parent quinolinone–chalcones.

Among all the pyrazoline analogues tested, the most potent LOX inhibitor is compound $\bf 9b$ (IC₅₀=10 μ M) which possesses a p-methoxy-phenyl substituent attached to the nitrogen of the pyrazoline moiety. In addition, pyrazoline $\bf 9b$ exhibits good antioxidant activity in both DPPH and lipid peroxidation assays; thus, it can be considered as a promising lead compound for further development. Regarding structural modifications on $\bf 9b$, replacement of the p-methoxy group by a p-cyano ($\bf 9a$) or a p-benzyloxy ($\bf 9c$) group or insertion of a methyl group on the heterocyclic NH of the quinolinone ring (compound $\bf 9i$) resulted in loss of activity.

However, the insertion of a methyl group on the nitrogen of the heterocyclic ring of the quinolinone moiety seems to enhance LOX inhibitory activity in the case of pyrazolines that share the *p*-cyano-phenyl or the *p*-benzyloxy-phenyl substituent as a common structural feature: N–H pyrazolines **9a**, **9d** and **9e** (IC₅₀=27.3% at 0,1 mM, 52.0 and 63.0 μ M, respectively) are less potent than their N-CH₃ analogues **9g**, **9j** and **9m** (IC₅₀=65.0, 15.0 and 39.0 μ M, respectively).

The comparison of pyrazoline analogues possessing two methoxy groups at the aromatic ring B (compounds **9a**, **9c**, **9g**, **9h**, **9i**) with their analogues which bear one methoxy and one hydroxyl group at ring B (compounds **9d**, **9e**, **9j**, **9k**, **9l**) shows that as a general trend, the presence of the hydroxyl group dramatically increases the ability of the compounds to inhibit LOX. In fact, compound **9j** is the second best lipoxygenase inhibitor in this series.

Molecular modeling of the synthesized derivatives in soybean LOX

All the synthesized derivatives have been subjected to in silico docking. The docking results provided useful interpretation of the experimental study. Figure 2 depicts the preferred docking pose for the most potent derivative 9b in soybean LOX (PDB code: 3PZW) presenting a high AutoDock Vina score (-10.3 kcal/mol). From the docking results, it can be concluded that the novel synthesized derivatives present allosteric interactions with the enzyme. It seems that **9b** accommodates to an extensively hydrophobic cavity with possible hydrophobic interactions (π – π stacking). It is possible that 9b extends into the hydrophobic domain and prevents access of substrates to the active site and hence prevents lipoxygenation. It is well known that LOX inhibitory activity is based on a carbon-centered radical on a lipid chain and most LOX inhibitors are antioxidants or free radical scavengers [69]. Pyrazoline **9b** is also a potent antioxidant;

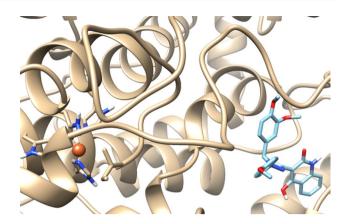


Fig. 2 Docking orientation of 9b (depicted in turquoise) bound to soybean LOX

therefore, it can serve as the starting point for the development of potential agents with combined anti-inflammatory and antioxidant activity.

Conclusions

In conclusion, this work reports the synthesis of four quinolinone-chalcones and fifteen novel structurally modified pyrazoline analogues, which to our knowledge have not been reported in the literature. In order to classify the new molecules as promising antioxidant and anti-inflammatory agents, we studied how the final biological effect is affected in relation to the different substituents at several positions of the framework, like the nitrogen heteroatom of the quinolinone ring, the aromatic ring B of the chalcone moiety, as well as the phenyl group attached to the pyrazoline ring. The evaluation of all the synthesized compounds showed that pyrazolines 9b and 9m possess the best combined activity (IC $_{50}$ 10.0 and 39.0 μM for LOX inhibition, 86% and 100% for DPPH scavenging ability and 83% and 79% in lipid peroxidation inhibition, respectively). The in silico docking results revealed that the pyrazoline analogue 9b showed high AutoDock Vina score (-10.3 kcal/mol), while all the tested derivatives presented allosteric interactions with the enzyme.

Experimental

Chemistry

General methods

The reagents and solvents used for synthesis and analysis were commercially available and used without further



purification. NMR spectra were recorded on a Varian 600 MHz spectrometer at the National Hellenic Research Foundation. The HR-MS spectrum was obtained using a UHPLC-MSn Orbitrap Velos-Thermo mass spectrometer at the National Hellenic Research Foundation. Melting points were determined on a Gallenkamp MFB-595 melting point apparatus and are uncorrected.

Synthesis

General procedure (A) for the synthesis of quinolinone–chalcone analogues (8a–8d)

Equimolar amounts of 3-acetyl-4-hydroxy-2-(1H)-quinolinone (3, 6) and the appropriate benzaldehyde (7) are dissolved in absolute ethanol (EtOH), and a catalytic amount of piperidine is added. The reaction mixture is stirred for 5–24 h at 78 °C with a reflux condenser, while the reaction is monitored by thin-layer chromatography (TLC). After the reaction is complete, the mixture is cooled in an ice bath and acidified with aq. HCl 10% v/v. Finally, the product is obtained from the acidified aqueous solution in solid form.

(E)-3-(3-(3,4-dimethoxyphenyl)acryloyl)-4-hydroxyquino-lin-2(1H)-one (8a)

This compound was prepared following the general procedure A. 3-acetyl-4-hydroxy-2-(1H)-quinolinone (3) (1000 mg, 4,92 mmol) and 3,4-dimethoxy benzaldehyde (7a) (810 mg, 4.92 mmol) were dissolved in 20 ml EtOH and 40 drops of piperidine were added. The mixture was stirred at 78 °C for 5 h. The solid (chalcone 8a) was obtained upon acidification as yellow powder. Yield: 680 mg (68%); m.p. > 250 °C; 1 H NMR (DMSO-d₆, 600 MHz): δ ppm 18.32 (s, 1H, OH), 11.44 (s, 1H, NH), 8.54 (d, 1H, J=15 Hz, COCH=CHAr), 8.01 (d, 1H, J=4.2 Hz, Ar–H), 7.94 (d, 1H, J=15.6 Hz, COCH=CHAr), 7.67 (t, 1H, J=6.0 Hz, Ar–H), 7.37 (d, 1H, J=8.4 Hz, Ar–H), 7.27 (m, 3H, Ar–H), 7.08 (d, 1H, J=8.4 Hz, Ar–H), 3.83 (s, 6H, 2OCH₃).

(E)-4-hydroxy-3-(3-(4-hydroxy-3-methoxyphenyl)acryloyl) quinolin-2(1H)-one (**8b**)

This compound was prepared following the general procedure A. 3-acetyl-4-hydroxy-2-(1H)-quinolinone (3) (500 mg, 2.46 mmol) and 4-hydroxy-3-methoxy benzaldehyde (**7b**) (374.3 mg, 2.46 mmol) were dissolved in 10 ml EtOH, and 20 drops of piperidine were added. The mixture was stirred at 78 °C for 5 h. The solid (chalcone **8b**) was obtained upon acidification as orange powder. Yield: 290 mg (58%); m.p. > 250 °C; ¹H NMR (DMSOde, 600 MHz): δ ppm 18.46 (s, 1H, OH), 11.42 (s, 1H, NH), 9.90 (s, 1H, Ar-OH), 8.51 (d, 1H, J = 15.6 Hz,

COCH = CH-Ar), 7.00 (d, 1H, J = 8.4 Hz, Ar–H), 7.93 (d, 1H, J = 15.6 Hz, COCH = CH-Ar), 7.67 (t, 1H, J = 7.2 Hz, Ar–H), 7.29 (m, 3H, Ar–H), 7.22 (t, 1H, J = 7.8 Hz, Ar–H), 6.90 (d, 1H, J = 7.8 Hz, Ar–H), 3.85 (s, 3H, 2OCH₃);

(E)-3-(3-(3,4-dimethoxyphenyl)acryloyl)-4-hydroxy-1-methylquinolin-2(1H)-one (8c)

This compound was prepared following the general procedure A. 3-acetyl-4-hydroxy-2-(1-CH₃)-quinolinone (**6**) (500 mg, 2.30 mmol) and 3,4-dimethoxy benzaldehyde (**7a**) (382.2 mg, 2.30 mmol) were dissolved in 15 ml EtOH, and 18 drops of piperidine were added. The mixture was stirred overnight at 78 °C. The solid produced upon acidification was triturated from methanol/dichloromethane to afford chalcone **8c** as orange powder. Yield: 357 mg (71%); m.p. > 250 °C ¹H NMR (CDCl₃, 600 MHz): δ ppm 18.20 (s, 1H, OH), 8.58 (d, 1H, J=15.6 Hz, COCH=CH-Ar), 8.25 (d, 1H, J=7.8 Hz, Ar-H), 7.96 (d, 1H, J=15.6 Hz, COCH=CH-Ar), 7.68 (t, 1H, J=7.8 Hz, Ar-H), 7.32 (m, 2H, Ar-H), 7.26 (m, 2H, Ar-H), 6.89 (d, 1H, J=8.4 Hz, Ar-H), 3.97 (s, 3H, 2OCH₃), 3.93 (s, 3H, OCH₃), 3.67 (s, 3H, NCH₃);

(E)-4-hydroxy-3-(3-(4-hydroxy-3-methoxyphenyl) acryloyl)-1-methylquinolin-2(1H)-one (8d)

This compound was prepared following the general procedure A. 3-acetyl-4-hydroxy-2-(1-CH₃)-quinolinone (6) (600 mg, 2.76 mmol) and 4-hydroxy-3-methoxy benzaldehyde (**7b**) (419.9 mg, 2.76 mmol) were dissolved in 18 ml EtOH, and 22 drops of piperidine were added. The mixture was stirred overnight at 78 °C. The solid produced upon acidification was triturated from methanol/dichloromethane to afford chalcone 8d as orange powder. Yield: 286 mg (48%); m.p. > 250 °C; ¹H NMR (DMSO-d₆, 600 MHz): δ ppm 18.25 (s, 1H, OH), 9.90 (s, 1H, Ar-OH), 8.44 (d, 1H, J = 15.6 Hz, COCH = CH-Ar), 8.09 (d, 1H, J = 7.8 Hz, Ar-H), 7.89 (d, 1H, J = 15.6 Hz, COCH = CH-Ar), 7.77 (t, 1H, J = 7.8 Hz, Ar - H), 7.50 (d, 1H, J = 8.4 Hz, Ar - H),7.27 (m, 3H, Ar–H), 6.88 (d, 1H, J = 7.8 Hz, Ar–H), 3.85 $(s, 3H, OCH_3), 3.55 (s, 3H, NCH_3); ^{13}C NMR (DMSO$ d_6 , 600 MHz): δ ppm 193.0 (C, C-11), 175.7 (C, C-4), 160.7 (C, C-2), 150.4 (C, C-3', C-4'), 148.0 (CH, C-12), 146.1 (C, C-5), 141.4 (C, C-1'), 135.4 (CH, C-8), 126.3 (CH, C-13), 125.3 (CH, C-2'), 123.7 (CH, C-5'), 122.1 (CH, C-6'), 121.1 (CH, C-6), 116.0 (CH, C-7), 115.2 (CH, C-9), 112.3 (C, C-10), 104.9 (C, C-3), 55.7 (CH₃, Ar-O-<u>CH</u>₃), 29.1 (CH₃, N-<u>CH</u>₃); HR-MS *m/z* (neg): 350.10314 C₂₀H₁₇NO₅ (calcd. 351.1107).



General procedure (B) for the synthesis of pyrazoline analogues (9a-9o)

The appropriate hydrazine derivative (2.5 eq.) is dissolved in absolute ethanol. The appropriate quinolinone—chalcone **8** (1 eq.) is dissolved in acetic acid, and it is added dropwise to the ethanolic solution of hydrazine. The resulting mixture is stirred at 120 °C for 24 h. The reaction is monitored by TLC. After completion of the reaction, the mixture is poured in ice and a yellow or orange precipitate is formed. The solid is filtered and washed with ice water. If no solid precipitates, the mixture is extracted with dichloromethane, the organic extracts are dried with Na₂SO₄ and the solvent is evaporated under reduced pressure to give the pyrazoline analogue as a solid product. Final compounds are further purified by trituration with methanol and dichloromethane.

4-(5-(3,4-Dimethoxyphenyl)-3-(4-hydroxy-2-oxo-1,2-dihydroquinolin-3-yl)-4,5-dihydro-1H-pyrazol-1-yl)benzonitrile (9a)

This compound was prepared following the general procedure B. 4-cyano phenyl-hydrazine hydrochloride (89.1 mg, 0.53 mmol) was dissolved in 0.67 ml of absolute ethanol. Ouinolinone-chalcone 8a (73 mg, 0.21 mmol) was dissolved in 1.3 ml of acetic acid and it was added dropwise to the ethanolic solution hydrazine. The resulting solution was stirred at 120 °C for 24 h. The solid produced upon filtration was triturated from methanol/dichloromethane to afford the pure pyrazoline **9a** as yellow powder. Yield: 35.8 mg (49%); m.p. > 250 °C; ¹H NMR (DMSO-d₆, 600 MHz,): δ ppm 13.04 (s, 1H, OH), 11.61 (s, 1H, NH), 7.98 (br, 1H, Ar-H), 7.62 (m, 3H, Ar-H), 7.27 (m, 2H, Ar-H), 6.94 (m, 4H, Ar-H), 6.71 (br, 1H, Ar-H), 5.48 (br, 1H, C_5 - \underline{H}_x), 4.21 (br, 1H, C_4 - \underline{H}_B), 3.72 (br, 6H, 2OC \underline{H}_3), 3.59 (br, 1H, C_4 - \underline{H}_A); ¹³C NMR (DMSO-d₆, 600 MHz): δ ppm 163.7 (C, C-4'), 161.7 (C, C-3), 154.2 (C, C-2'), 149.7 (C, C-3"), 148.7 (C, C-4"), 146.1 (C, C-1"), 139.0 (C, C-10'), 133.9 (CH, C-3", C-5"), 133.8 (CH, C-8'), 132.8 (C, C-1"), 123.9 (CH, C-6'), 122.4 (CH, C-7'), 120.2 (C, C-6"), 117.7 (C, Ar– $\underline{C} \equiv N$), 115.7 (CH, C-9'), 114.3 (C, C-5') 113.1 (CH, C-2'", C-6'"), 112.6 (CH, C-5"), 110.0 (C, C-2"), 101.5 (CH, C-4""), 100.0 (C, C-3'), 60.9 (CH, C-5), 56.0 (CH₃, Ar–O-C<u>H</u>₃, Ar–O- $\underline{CH_3}$), 55.9 (CH₃, Ar-O-C<u>H</u>₃, Ar-O-C<u>H</u>₃), 47.2 (CH₂, C-4); HR-MS m/z (neg): 465.15554 $C_{27}H_{22}N_4O_4$ (calcd. 466.1641).

3-(5-(3,4-Dimethoxyphenyl)-1-(4-methoxyphenyl)-4,5-dihydro-1H-pyrazol-3-yl)-4-hydroxyquinolin-2(1H)-one (**9b**)

This compound was prepared following the general procedure B. 4-methoxy phenyl hydrazine hydrochloride (149.1 mg, 0.85 mmol) was dissolved in 0.94 ml of absolute

ethanol. Quinolinone-chalcone 8a (120 mg, 0.34 mmol) was dissolved in 1.9 ml of acetic acid, and it was added dropwise to the ethanolic solution of hydrazine. The resulting solution was stirred at 120 °C for 24 h. The solid produced upon filtration was triturated from methanol/dichloromethane to afford the pure pyrazoline **9b** as yellow powder. Yield: 82 mg (68.3%); m.p. 230 °C; 1 H NMR (DMSO-d₆, 600 MHz): δ ppm 13.54 (s, 1H, OH), 11.54 (s, 1H, NH), 7.96 (d, 1H, J = 7.8 Hz, Ar-H), 7.55 (t, 1H, J = 7.8 Hz, Ar-H), 7.29 (d, 1H, J = 8.7 Hz, Ar–H), 7.23 (t, 1H, J = 7.8 Hz, Ar–H), 6.99 (s, 1H, Ar–H), 6.86 (m, 6H, Ar–H), 5.16 (dd, 1H, J=11.4, 9.0 Hz, $C_5 - \underline{H}_X$), 4.19 (dd,1H, J = 18.6, 12.0 Hz, $C_4 - \underline{H}_B$), 3.72 (br, 6H, 2OCH₃), 3.67 (s, 3H, OCH₃), 3.44 (dd, 1H, J = 18.0, 8.4 Hz, $C_4 - H_A$); ¹³C NMR (DMSO-d₆, 600 MHz): δ ppm 162.4 (C, C-4'), 161.3 (C, C-3), 153.4 (C, C-2'), 150.6 (C, C-4"), 149.1 (C, C-3"), 148.1 (C, C-4"), 138.4 (C, C-10'), 138.2 (C, C-1"), 134.3 (CH, C-8'), 123.3 (C, C-1"), 123.1 (CH, C-6'), 122.1 (C, C-7'), 121.6 (CH, C-6"), 118.2 (CH, C-3", C-5"), 115.4 (CH, C-9'), 115.2 (CH, C-5"), 114.8 (CH, C-2", C-6"), 114.4 (C, C-5'), 112.2 (CH, C-2"), 101.4 (C, C-3'), 63.2 (CH, C-5), 55.6 (CH₃, Ar–O-<u>CH₃</u>), 55.4 (CH₃, Ar–O-<u>CH₃</u>), 55.1 (CH₃, Ar–O-<u>CH₃</u>), 46.6 (CH₂, C-4); HR-MS m/z (pos): 471.17808 $C_{27}H_{25}N_3O_5$ (calcd. 471.1794).

3-(1-(4-(Benzyloxy)phenyl)-5-(3,4-dimethoxyphenyl)-4, 5-dihydro-1H-pyrazol-3-yl)-4-hydroxyquinolin-2(1H)-one (9c)

This compound was prepared following the general procedure B. 4-benzyloxy phenyl hydrazine hydrochloride (214.1 mg, 0.85 mmol) was dissolved in 0.9 ml of absolute ethanol. Quinolinone-chalcone 8a (120 mg, 0.34 mmol) was dissolved in 1.9 ml of acetic acid, and it was added dropwise to the ethanolic solution of hydrazine. The resulting solution was stirred at 120 °C for 24 h. The solid produced upon filtration was triturated from methanol/dichloromethane to afford the pure pyrazoline 9c as yellow powder. Yield: 73.2 mg (61%); m.p. 245 °C; ¹H NMR (DMSO-d₆, 600 MHz): δ ppm 13.53 (s, 1H, OH), 11.55 (s, 1H, NH), 7.96 (d, 1H, J = 7.2 Hz, Ar-H), 7.55 (t, 1H, J = 7.2 Hz, Ar-H), 7.38 (m, 4H, Ar-H), 7.30 (m, 2H, Ar-H), 7.23 (t, 1H, J = 7.2 Hz, Ar-H), 6.98 (s, 1H, Ar-H), 6.90 (m, 5H, Ar-H), 6.81 (d, 1H, J = 7.2 Hz, Ar-H), 5.17 (br, 1H, $C_5 - \underline{H}_X$), 5.0 (s, 2H, Ar-C \underline{H}_2 -O-Ar), 4.19 (dd, 1H, J=18.6, 12.0 Hz, C_4 - H_B), 3.72 (s, 6H, OC H_3), 3.44 (dd, 1H, J= 18.0, 8.4 Hz, C_4 - H_A); ¹³C NMR (DMSO- d_6 , 600 MHz): δ ppm 162.4 (C, C-4'), 161.3 (C, C-3), 152.4 (C, C-2'), 150.6 (C, C-4'"), 149.1 (C, C-3"), 148.1 (C, C-4"), 138.5 (C, C-10'), 138.2 (C, C-1""), 137.3 (C, C-1""), 134.3 (CH, C-8'), 131.8 (C, C-1"), 128.3 (CH, C-3"", C-5""), 127.7 (CH, C-4""), 127.6 (CH, C-2"", C-6""), 123.2 (CH, C-6'), 121.8 (CH, C-7'), 118.0 (CH, C-6"), 115.6(CH, C-2", C-6"), 115.1 (CH,



C-9'), 114.8 (CH, C-3'", C-5'"), 114.0 (CH, C-5"), 112.0 (C, C-5'), 109.8 (CH, C-2"), 101.4 (C, C-3'), 69.5 (CH₂, Ar–O-<u>CH₂</u>-Ar), 63.3(CH, C-5), 55.4 (CH₃, Ar–O-<u>CH₃</u>), 30.7 (CH₂, C-4); HR-MS *m/z* (pos): 548.21717 C₃₃H₂₉N₃O₅ (calcd. 547.2107).

4-(3-(4-Hydroxy-2-oxo-1,2-dihydroquinolin-3-yl)-5-(4-hydroxy-3-methoxyphenyl)-4,5-dihydro-1H-pyrazol-1-yl)benzonitrile (**9d**)

This compound was prepared following the general procedure B. 4-cyano phenyl hydrazine hydrochloride (150.8 mg, 0.89 mmol) was dissolved in 1 ml of absolute ethanol. Quinolinone-chalcone 8b (120 mg, 0.36 mmol) was dissolved in 2 ml of acetic acid, and it was added dropwise to the ethanolic solution of hydrazine. The resulting solution was stirred at 120 °C for 24 h. The solid produced upon filtration was triturated from methanol/dichloromethane to afford the pure pyrazoline 9d as orange powder. Yield: 62 mg (51.7%); m.p. > 250 °C; ¹H NMR (DMSO-d₆, 600 MHz): δ ppm 13.08 (s, 1H, O<u>H</u>), 11.61 (s, 1H, N<u>H</u>), 9.01 (s, 1H, Ar-OH), 7.98 (d, 1H, J = 8.4 Hz, Ar-H), 7.62 (d, 2H, J=9.0 Hz, Ar-H), 7.58 (t, 1H, J=8.4 Hz, Ar-H), 7.29 (d, 1H, J = 8.4 Hz, Ar–H), 7.25 (t, 1H, J = 7.8 Hz, Ar–H), 6.98 (d, 2H, J=9.0 Hz, Ar-H), 6.90 (d, 1H, J=1.2 Hz, Ar-H),6.71 (d, 1H, J=8.4 Hz, Ar–H), 6.59 (dd, 1H, J=7.8, 1.2 Hz, Ar-H), 5.42 (dd, 1H, J = 11.4, 5.4 Hz, $C_5 - H_X$), 4.19 (dd, 1H, J = 18.6, 12.0 Hz, C_4 - H_B), 3.73 (s, 3H, OC H_3), 3.58 (dd, 1H, J = 19.2,6.0 Hz, $C_4 - \underline{H}_A$); ¹³C NMR (DMSO-d₆, 600 MHz): δ ppm 163.5 (C, C-4'), 161.6 (C, C-3), 154.2 (C, C-2'), 148.5 (C, C-3"), 146.5 (C, C-4"), 146.0 (C, C-1"), 138.9 (C, C-10'), 133.9 (CH, C-3"', C-5"'), 132.8 (C, C-1"), 132.3 (CH, C-8'), 123.8 (CH, C-6'), 122.4 (CH, C-7'), 120.3 (CH, C-6"), 118.1 (C, Ar- $\mathbb{C} = \mathbb{N}$), 116.3 (CH, C-9'), 115.7 (CH, C-5"), 114.2 (C, C-5'), 113.1 (CH, C-2"", C-6""), 110.4 (C, C-2"), 101.5 (C, C-4""), 99.9 (C, C-3'), 61.0 (CH, C-5), 56.1 (CH₃, Ar–O-<u>CH₃</u>,), 47.2 (CH₂, C-4); HR-MS *m/z* (neg): 451.13988 C₂₆H₂₀N₄O₄ (calcd. 452.1485).

3-(1-(4-(Benzyloxy)phenyl)-5-(4-hydroxy-3-methoxyphenyl)-4,5-dihydro-1H-pyrazol-3-yl)-4-hydroxyquino-lin-2(1H)-one (**9e**)

This compound was prepared following the general procedure B. 4-benzyloxy phenyl hydrazine hydrochloride (223 mg, 0.89 mmol) was dissolved in 1 ml of absolute ethanol. Quinolinone-chalcone **8b** (120 mg, 0.36 mmol) was dissolved in 2 ml of acetic acid and it was added dropwise to the ethanolic solution of hydrazine. The resulting solution was stirred at 120 °C for 24 h. The solid produced upon filtration was triturated from methanol/dichloromethane to afford the pure pyrazoline **9e** as orange powder. Yield: 68 mg (56.7%); m.p. > 250 °C; ¹H NMR (DMSO-d₆,

600 MHz): δ ppm 13.54 (s, 1H, OH), 11.54 (s, 1H, NH), 8.95 (s, 1H, Ar-OH), 7.95 (d, 1H, J = 7.8 Hz, Ar-H), 7.55 (t, 1H, J = 7.8 Hz, Ar - H), 7.41 (m, 2H, Ar - H), 7.37 (t,2H, J = 7.2 Hz, Ar - H), 7.30 (m, 2H, Ar - H), 7.23 (t, 1H, J = 7.2 Hz, Ar-H), 6.93 (m, 3H, Ar-H), 6.88 (m, 2H, Ar-H), 6.71 (m, 2H, Ar-H), 5.11 (br, 1H, C_5 - \underline{H}_x), 5.00 (s, 2H, Ar-C \mathbf{H}_2 -O-Ar), 4.17 (dd, 1H, J= 18.6, 12.0 Hz, C_4-H_B), 3.73 (s, 3H, OCH_3), 3.43 (dd, 1H, J=18.6, 8.4 Hz, C_4 - \underline{H}_A); ¹³C NMR (DMSO- d_6 , 600 MHz): δ ppm 162.3 (C, C-4'), 161.3 (C, C-3), 152.4 (C, C-2'), 150.6 (C, C-4'"), 147.9 (C, C-3"), 145.9 (C, C-4"), 138.6 (C, C-10'), 138.2 (C, C-1""), 137.3 (C, C-1""), 132.8 (CH, C-8'), 131.8 (C, C-1"), 128.3 (CH, C-3"", C-5""), 127.7 (CH, C-4""), 127.6 (CH, C-2"", C-6""), 123.1 (CH, C-6'), 121.8 (CH, C-7'), 118.4 (CH, C-6"), 115.6 (CH, C-2", C-6"), 115.1 (CH, C-9'), 114.8 (CH, C-3", C-5"), 114.0 (CH, C-5"), 112.9 (C, C-5'), 110.2 (CH, C-2"), 101.4 (C, C-3'), 69.5 (CH₂, Ar–O-<u>CH</u>₂-Ar), 63.4 (CH, C-5), 55.6 $(CH_3, Ar-O-CH_3)$, 46.6 $(CH_2, C-4)$; HR-MS m/z (pos): 533.19400 C₃₂H₂₇N₃O₅ (calcd. 533.1951).

3-(1-(4-Chlorophenyl)-5-(4-hydroxy-3-methoxyphenyl)-4, 5-dihydro-1H-pyrazol-3-yl)-4-hydroxyquinolin-2(1H)-one (9f)

This compound was prepared following the general procedure B. 4-chloro-phenyl hydrazine hydrochloride (100.9 mg, 0.56 mmol) was dissolved in 0.6 ml of absolute ethanol. Quinolinone-chalcone **8b** (76 mg, 0.2 mmol) was dissolved in 1.2 ml of acetic acid and was added dropwise to the ethanolic solution of hydrazine. The resulting solution was stirred at 120 °C for 24 h. The solid produced upon filtration was triturated from methanol/dichloromethane to afford the pure pyrazoline 9f as orange powder. Yield: 32 mg (42.1%); m.p. > 250 °C; ¹H NMR (DMSO d_6 , 600 MHz): δ ppm 13.29 (s, 1H, OH), 11.58 (s, 1H, $N\underline{H}$), 9.0 (s, 1H, Ar-O \underline{H}), 7.96 (d, 1H, J = 7.8 Hz, Ar-H), 7.56 (t, 1H, J = 7.2 Hz, Ar–H), 7.25 (m, 4H, Ar–H), 6.91 (m, 3H, Ar-H), 6.71 (d, 1H, J=8.4 Hz, Ar-H), 6.64 (dd,1H, J = 7.8, 0.6 Hz, Ar–H), 5.26 (dd, 1H, J = 12.0, 7.2 Hz, $C_5 - \underline{H}_X$, 4.18 (dd, 1H, J = 18.6, 12.0 Hz, $C_4 - \underline{H}_B$), 3.73 (s, 3H, OCH₃), 3.51 (dd, 1H, J = 19.2, 7.2 Hz, C₄-H_A); ¹³C NMR (DMSO-d₆, 600 MHz): δ ppm 162.6 (C, C-4'), 161.3 (C, C-3), 151.7 (C, C-2'), 148.0 (C, C-3"), 146.0 (C, C-4"), 142.4 (C, C-1"), 138.3 (C, C-10'), 132.4 (CH, C-8'), 132.1 (C, C-4'"), 130.1 (C, C-1"), 128.9 (CH, C-3'", C-5"), 123.2 (CH, C-6'), 122.9 (CH, C-7'), 121.9 (CH, C-6"), 119.6 (CH, C-2", C-6"), 118.0 (CH, C-9'), 115.2 (CH, C-5"), 113.9 (C, C-5'), 111.6 (CH, C-2"), 101.3 (C, C-3'), 61.8 (CH, C-5), 55.6 (CH₃, Ar–O-<u>CH₃</u>,), 46.8 (CH₂, C-4); HR-MS m/z (neg): 460.10629 $C_{25}H_{20}CIN_3O_4$ (calcd. 461.1142).

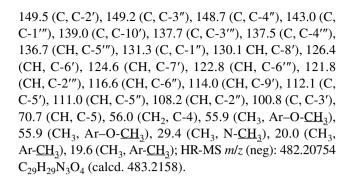


4-(5-(3,4-Dimethoxyphenyl)-3-(4-hydroxy-1-me-thyl-2-oxo-1,2-dihydroquinolin-3-yl)-4,5-dihydro-1H-pyrazol-1-yl)benzonitrile (**9g**)

This compound was prepared following the general procedure B. 4-cyano phenyl hydrazine hydrochloride (116.1 mg, 0.68 mmol) was dissolved in 0.9 ml of absolute ethanol. Quinolinone-chalcone 8c (100 mg, 0.27 mmol) was dissolved in 1.7 ml of acetic acid and it was added dropwise to the ethanolic solution of hydrazine. The resulting solution was stirred at 120 °C for 24 h. The solid produced upon filtration was triturated from methanol/dichloromethane to afford the pure pyrazoline **9g** as orange powder. Yield: 73 mg (73%); m.p. > 250 °C; ¹H NMR (DMSO-d₆, 600 MHz): δ ppm 13.07 (s, 1H, OH), 8.07 (br, 1H, Ar-H), 7.69 (br, 1H, Ar-H), 7.62 1H, Ar-H), 6.96 (m, 3H, Ar-H), 6.88 (d, 1H, J = 8.4 Hz, Ar-H), 6.71 (d, 1H, J = 8.4 Hz, Ar-H), 5.46 (br, 1H, C_5-H_X , 4.21 (dd, 1H, J=18.6, 12.6 Hz, C_4-H_B), 3.71 (br, 6H, $2OC\underline{H}_3$), 3.59 (br, 1H, C_4 - \underline{H}_Δ), 3.57 (s, 3H, $NC\underline{H}_3$); ¹³C NMR (DMSO-d₆, 600 MHz): δ ppm 162.2 (C, C-3), 160.8 (C, C-4'), 154.4 (C, C-2'), 149.6 (C, C-3"), 148.7 (C, C-4"), 146.0 (C, C-1"), 139.8 (C, C-10'), 134.0 (CH, C-3", C-5"), 133.8 (C, C-1"), 133.3 (CH, C-8'), 124.3 (CH, C-6'), 122.6 (CH, C-7'), 117.8 (C, Ar- $\underline{C} \equiv N$), 115.4 (CH, C-6"), 115.0 (CH, C-9'), 113.1 (CH, C-2'", C-6'"), 112.6 (CH, C-5"), 110.0 (C, C-5'), 101.4 (C, C-4"'), 100.1 (CH, C-2"), 99.5 (C, C-3'), 60.9 (CH, C-5), 55.9 (CH₃, Ar–O-<u>CH₃</u>), 55.9 (CH₃, Ar-O-<u>CH</u>₃), 47.4 (CH₂, C-4), 29.5 (CH₃, N-<u>CH</u>₃); HR-MS m/z (neg): 479.17102 $C_{28}H_{24}N_4O_4$ (calcd. 480.1798).

3-(5-(3,4-Dimethoxyphenyl)-1-(3,4-dimethylphenyl)-4, 5-dihydro-1H-pyrazol-3-yl)-4-hydroxy-1-methylquino-lin-2(1H)-one (**9h**)

This compound was prepared following the general procedure B. 1-(3,4-dimethylphenyl) hydrazine hydrochloride (118.2 mg, 0.68 mmol) was dissolved in 0.9 ml of absolute ethanol. Quinolinone-chalcone 8c (100 mg, 0.27 mmol) was dissolved in 1.7 ml of acetic acid, and it was added dropwise to the ethanolic solution of hydrazine. The resulting solution was stirred at 120 °C for 24 h. The solid produced upon filtration was triturated from methanol/dichloromethane to afford the pure pyrazoline **9h** as orange powder. Yield: 34 mg (34%); m.p. > 250 °C; 1 H NMR (CDCl₃, 600 MHz): δ ppm 13.74 (br, 1H, OH), 8.17 (d, 1H, J = 7.8 Hz, Ar–H), 7.56 (t, 1H, J = 7.8 Hz, Ar-H), 7.25 (m, 2H, Ar-H), 6.94 (d, 1H, J=7.8 Hz, Ar-H), 6.81 (m, 4H, Ar-H), 6.64 (d,1H, J = 7.8 Hz, Ar-H), 5.01 (dd, 1H, J = 12.0, 9.0 Hz, $C_5-\underline{H}_X$), 4.29 (dd, 1H, J=18.6, 12.6 Hz, $C_4-\underline{H}_B$), 3.82 (br, 6H, 2OCH₃), 3.76 (br, 1H, C₄-H_A), 3.61 (s, 3H, NCH₃), 2.18 (s, 3H, Ar-C \underline{H}_3), 2.145 (s, 3H, Ar-C \underline{H}_3); ¹³C NMR $(CDCl_3, 600 \text{ MHz}): \delta \text{ ppm } 162.1 \text{ (C, C-3)}, 160.8 \text{ (C, C-4')},$



3-(5-(3,4-Dimethoxyphenyl)-1-(4-methoxyphenyl)-4, 5-dihydro-1H-pyrazol-3-yl)-4-hydroxy-1-methylquinolin-2(1H)-one (9i)

This compound was prepared following the general procedure B. 4-methoxy phenyl hydrazine hydrochloride (239 mg, 1.37 mmol) was dissolved in 1.7 ml of absolute ethanol. Quinolinone-chalcone 8c (200 mg, 0.55 mmol) was dissolved in 3.5 ml of acetic acid, and it was added dropwise to the ethanolic solution of hydrazine. The resulting solution was stirred at 120 °C for 24 h. The solid produced upon filtration was triturated from methanol/dichloromethane to afford the pure pyrazoline 9i as orange powder. Yield: 152 mg (76%); m.p. > 250 °C; ¹H NMR (CDCl₃, 600 MHz): δ ppm 13.74 (s, 1H, OH), 8.17 (dd, 1H, J=7.8, 0.6 Hz, Ar-H), 7.58(t, 1H, J = 8.4 Hz, Ar–H), 7.283 (m, 2H, Ar–H), 6.92 (m, 2H, Ar-H), 6.87 (m, 2H, Ar-H), 6.81 (m, 3H, Ar-H), 4.96 (dd, 1H, J = 12.0, 9.6 Hz, $C_5 - \underline{H}_X$), 4.33 (dd, 1H, J = 18.6, 12.0 Hz, C_4 - \underline{H}_B), 3.86 (s, 3H, $OC\underline{H}_3$), 3.83 (s, 3H, $OC\underline{H}_3$), 3.74 (s, 3H, OCH₃), 3.64 (s, 3H, NCH₃), 3.60 (br, 1H, $C_4 - H_A$); ¹³C NMR (CDCl₃, 600 MHz): δ ppm 162.6 (C, C-3), 161.9 (C, C-4'), 154.1 (C, C-2'), 151.4 (C, C-4'"), 149.6 (C, C-3"), 149.5 (C, C-4"), 148.5 (C, C-10'), 139.4 (C, C-1"), 139.4 (C, C-1"), 134.8 (CH, C-8'), 124.7 (CH, C-6'), 124.6 (CH, C-7'), 121.9 (CH, C-6"), 118.8 (CH, C-9'), 116.1 (C, C-5'), 115.6 (CH, C-5"), 114.8 (CH, C-2"", C-6""), 111.7 (CH, C-3", C-5"), 109.3 (CH, C-2"), 101.7 (C, C-3'), 65.3 (CH, C-5), 56.0 (CH₃, Ar-O-CH₃), 55.8 (CH₃, Ar-O-<u>CH</u>₃), 55.6 (CH₃, Ar–O-<u>CH</u>₃), 47.4 (CH₂, C-4), 29.1 (CH₃, $N-CH_3$; HR-MS m/z (neg): 484.18742 $C_{28}H_{27}N_3O_5$ (calcd. 485.1951).

4-(3-(4-Hydroxy-1-methyl-2-oxo-1,2-dihydroquino-lin-3-yl)-5-(4-hydroxy-3-methoxyphenyl)-4,5-dihydro-1H-pyrazol-1-yl)benzonitrile (9j)

This compound was prepared following the general procedure B. 4-cyano phenyl hydrazine hydrochloride (120.7 mg, 0.71 mmol) was dissolved in 0.9 ml of absolute ethanol. Quinolinone-chalcone **8d** (100 mg, 0.28 mmol) was dissolved in 1.8 ml of acetic acid, and it was added dropwise to the ethanolic solution of hydrazine. The resulting solution



was stirred at 120 °C for 24 h. The solid produced upon filtration was triturated from methanol/dichloromethane to afford the pure pyrazoline 9j as orange powder. Yield: 82 mg (82%); m.p. > 250 °C; ¹H NMR (DMSO-d₆, 600 MHz): δ ppm 13.06 (s, 1H, OH), 9.01 (s, 1H, Ar-OH), 8.05 (d, 1H, J = 7.2 Hz, Ar-H), 7.67 (t, 1H, J = 6.6 Hz, Ar-H), 7.60 (d, 2H, J = 7.8 Hz, Ar - H), 7.48 (d, 1H, J = 7.8 Hz, Ar - H), 7.31(t, 1H, J=7.2 Hz, Ar-H), 6.95 (d, 2H, J=7.8 Hz, Ar-H),6.90 (br, 1H, Ar–H), 6.71 (d, 1H, J=7.8 Hz, Ar–H), 6.60 (d, 1H, J = 7.2 Hz, Ar-H), 5.38 (dd, 1H, J = 10.8, 4.2 Hz, $C_5-\underline{H}_X$), 4.18 (dd, 1H, J=18.6, 12.6 Hz, $C_4-\underline{H}_B$), 3.73 (s, 3H, OC \underline{H}_3), 3.57 (dd, 1H, J = 19.8, 3.6 Hz, $C_4 - \underline{H}_A$), 3.52 (s, 3H, NCH₃); 13 C NMR (DMSO-d₆, 600 MHz): δ ppm 162.2 (C, C-3), 160.8 (C, C-4'), 154.4 (C, C-2'), 148.5 (C, C-3"), 146.5 (C, C-4"), 146.0 (C, C-1"), 139.8 (C, C-10'), 133.9 (CH, C-3", C-5"), 133.1 (C, C-1"), 132.3 (CH, C-8'), 124.3 (CH, C-6'), 122.5 (CH, C-7'), 120.2 (CH, C-5"), 118.2 (C, $Ar-\underline{C}\equiv N$), 116.3 (CH, C-9'), 115.2 (CH, C-6"), 115.0 (C, C-5'), 113.1 (CH, C-2"', C-6"'), 110.4 (CH, C-2"), 101.3 (C, C-4"), 100.0 (C, C-3'), 61.0 (CH, C-5), 56.1 (CH₃, Ar-O-<u>CH</u>₃), 47.4 (CH₂, C-4), 29.4 (CH₃, N-<u>CH</u>₃); HR-MS *m/z* (neg): $465.15552 \, \text{C}_{27} \text{H}_{22} \text{N}_4 \text{O}_4$ (calcd. 466.1641).

3-(1-(3,4-Dimethylphenyl)-5-(4-hydroxy-3-methoxypheny l)-4,5-dihydro-1H-pyrazol-3-yl)-4-hydroxy-1-methylquino-lin-2(1H)-one (**9k**)

This compound was prepared following the general procedure B. 1-(3,4-dimethylphenyl) hydrazine hydrochloride (122.85 mg, 0.71 mmol) was dissolved in 0.9 ml of absolute ethanol. Quinolinone-chalcone 8d (100 mg, 0.28 mmol) was dissolved in 1.8 ml of acetic acid and it was added dropwise to the ethanolic solution of hydrazine. The resulting solution was stirred at 120 °C for 24 h. The solid produced upon filtration was triturated from methanol/dichloromethane to afford the pure pyrazoline 9k as orange powder. Yield: 67 mg (67%); m.p. > 250 °C; ¹H NMR (DMSO-d₆, 600 MHz): δ ppm 13.55 (s, 1H, OH), 8.95 (s, 1H, Ar-OH), 8.04 (d, 1H, J=7.2 Hz, Ar-H, 7.65 (t, 1H, J=7.2 Hz, Ar-H), 7.46 (d, 1H, J = 8.4 Hz, Ar–H), 7.30 (t, 1H, J = 7.2 Hz, Ar–H), 6.95 (d, 1H, J = 7.8 Hz, Ar–H), 6.91 (br, 1H, Ar–H), 6.75 (br, 1H, Ar-H), 6.70 (d, 1H, J = 7.8 Hz, Ar-H), 6.65 (d, 1H, J = 7.8 Hz, Ar-H), 6.60 (d, 1H, J = 7.2 Hz, Ar-H), 5.14 (dd, 1H, J = 10.8, 7.8 Hz, $C_5 - \underline{H}_X$), 4.14 (dd, 1H, J = 18.0, 12.0 Hz, C_4 - \underline{H}_B), 3.72 (s, 3H, $OC\underline{H}_3$), 3.52 (s, 3H, $NC\underline{H}_3$), 3.46 (dd, 1H, J = 18.6, 7.8 Hz, $C_4 - H_A$), 2.11 (br, 6H, 2Ar- CH_3); ¹³C NMR (DMSO-d₆, 600 MHz): δ ppm 161.4 (C, C-3), 160.8 (C, C-4'), 151.0 (C, C-2'), 148.3 (C, C-3"), 146.2 (C, C-4"), 142.3 (C, C-1"), 139.4 (C, C-10'), 137.1 (C, C-3"), 133.5 (C, C-4"), 132.6 (CH, C-5"), 130.4 (CH, C-8'), 127.7 (C, C-1"), 124.1 (CH, C-6'), 122.4 (CH, C-7'), 118.6 (CH, C-6"), 116.2 (CH, C-6"), 115.2 (CH, C-2"), 115.1 (CH, C-9'), 115.0 (CH, C-5"), 111.1 (C, C-5'), 110.5 (CH, C-2"), 101.6 (C, C-3'), 62.7 (CH, C-5), 56.0 (CH₃, Ar–O-<u>CH₃</u>), 47.1 (CH₂, C-4), 29.4 (CH₃, N-<u>CH₃</u>), 20.3 (CH₃, Ar-<u>CH₃</u>), 18.9 (CH₃, Ar-<u>CH₃</u>); HR-MS *m/z* (neg): 468.19315 C₂₈H₂₇N₃O₄ (calcd. 469.2002).

4-Hydroxy-3-(5-(4-hydroxy-3-methoxyphenyl)-1-(4-methoxyphenyl)-4,5-dihydro-1H-pyrazol-3-yl)-1-methylquino-lin-2(1H)-one (9l)

This compound was prepared following the general procedure B. 4-methoxy phenyl hydrazine hydrochloride (124.3 mg, 0.71 mmol) was dissolved in 0.9 ml of absolute ethanol. Quinolinone-chalcone 8d (100 mg, 0.28 mmol) was dissolved in 1.8 ml of acetic acid, and it was added dropwise to the ethanolic solution of hydrazine. The resulting solution was stirred at 120 °C for 24 h. The solid produced upon filtration was triturated from methanol/dichloromethane to afford the pure pyrazoline 91 as orange powder. Yield: 48 mg (48%); m.p. > 250 °C; ¹H NMR (DMSO-d₆, 600 MHz): δ ppm 13.57 (s, 1H, OH), 8.95 (s, 1H, Ar-OH), 8.04 (d, 1H, J = 7.2 Hz, Ar-H), 7.66 (t, 1H, J = 7.2 Hz, Ar-H), 7.48 (d, 1H, J = 8.4 Hz, Ar–H), 7.31 (t, 1H, J = 7.8 Hz, Ar–H), 6.94 (br, 1H, Ar-H), 6.86 (m, 4H, Ar-H), 6.71 (m, 2H, Ar-H), 5.08 (dd, 1H, J = 11.4, 9.0 Hz, $C_5 - \underline{H}_X$), 4.16 (dd, 1H, J = 18.6, 12.0 Hz, C_4 - H_B), 3.73 (s, 3H, OC H_3), 3.67 (s, 3H, OCH_3), 3.54 (s, 3H, NCH_3), 3.44 (dd, 1H, J=18.6, 8.4 Hz, C_4 - \underline{H}_A); ¹³C NMR (DMSO- d_6 , 600 MHz): δ ppm 161.5 (C, C-3), 160.9 (C, C-4'), 153.8 (C, C-2'), 151.2 (C, C-4"), 148.4 (C, C-3"), 146.3 (C, C-4"), 139.4 (C, C-10'), 138.8 (C, C-1"), 133.2 (CH, C-8'), 132.6 (C, C-1"), 124.1 (CH, C-6'), 122.4 (CH, C-7'), 118.9 (CH, C-6"), 116.2 (CH, C-9'), 115.4 (CH, C-2'", C-6'"), 115.3 (C, C-5'), 115.2 (CH, C-5"), 115.0 (CH, C-3"', C-5"'), 110.7 (CH, C-2"), 101.6 (C, C-3'), 63.9 (CH, C-5), 56.0 (CH₃, Ar-O-CH₃), 55.7 (CH₃, Ar-O-CH₃), 47.2 (CH₂, C-4), 29.4 (CH₃, N-CH₃); HR-MS m/z (pos): 472.18606 C₂₇H₂₅N₃O₅ (calcd. 471.1794).

3-(1-(4-(Benzyloxy)phenyl)-5-(4-hydroxy-3-methoxyphenyl)-4,5-dihydro-1H-pyrazol-3-yl)-4-hydroxy-1-methylquino-lin-2(1H)-one (9m)

This compound was prepared following the general procedure B. 4-benzyloxy phenyl hydrazine hydrochloride (178.4 mg, 0.71 mmol) was dissolved in 0.9 ml of absolute ethanol. Quinolinone-chalcone **8d** (100 mg, 0.28 mmol) was dissolved in 1.8 ml of acetic acid, and it was added dropwise to the ethanol solution of hydrazine. The resulting solution was stirred at 120 °C for 24 h. The solid produced upon filtration was triturated from methanol/dichloromethane to afford the pure pyrazoline **9 m** as orange powder. Yield: 38 mg (38%); m.p. > 250 °C; 1 H NMR (DMSO-d₆, 600 MHz): δ ppm 13.51 (s, 1H, OH), 8.93 (br, 1H, Ar-OH), 8.03 (br, 1H, Ar-H), 7.64 (br, 1H, Ar-H), 7.37 (m, 7H, Ar-H), 6.89 (m, 5H, Ar-H), 6.69



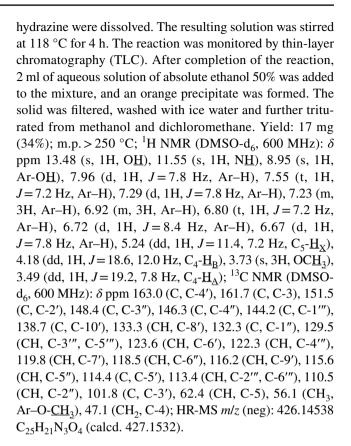
(br, 2H, Ar–H), 5.09 (br, 1H, C_5-H_X), 4.99 (s, 2H, Ar–CH₂-O–Ar), 4.17 (br, 1H, C_4-H_B), 3.72 (s, 3H, OCH₃), 3.55 (s, 3H, NCH₃), 3.45 (br, 1H, C_4-H_A); ¹³C NMR (DMSO-d₆, 600 MHz): δ ppm 161.0 (C, C-3), 160.4 (C, C-4'), 152.4 (C, C-2'), 150.8 (C, C-4'''), 147.9 (C, C-3''), 145.9 (C, C-4''), 138.9 (C, C-10'), 138.5 (C, C-1'''), 137.3 (C, C-1''''), 132.8 (CH, C-8'), 132.2 (C, C-1''), 128.4 (CH, C-3'''', C-5''), 127.7 (CH, C-4''''), 127.6 (CH, C-2''', C-6''''), 123.6 (CH, C-6'), 122.0 (CH, C-7'), 118.4 (CH, C-6''), 115.7 (CH, C-9'), 115.6 (CH, C-2''', C-6'''), 114.8 (CH, C-5''), 114.8 (CH, C-3''', C-5'''), 114.7 (C, C-5'), 110.2 (CH, C-2''), 101.1 (C, C-3'), 69.5 (CH₂, Ar–O-CH₂-Ar), 63.4 (CH, C-5), 55.6 (CH₃, Ar–O-CH₃), 46.8 (CH₂, C-4), 28.9 (CH₃, N-CH₃); HR-MS m/z (pos): 548.21901 $C_{33}H_{29}N_3O_5$ (calcd. 547.2107).

3-(1-(4-Chlorophenyl)-5-(4-hydroxy-3-methoxyphenyl)-4,5-dihydro-1H-pyrazol-3-yl)-4-hydroxy-1-methylquino-lin-2(1H)-one (9n)

This compound was prepared following the general procedure B. 4-chloro phenyl hydrazine hydrochloride (255.2 mg, 1.43 mmol) was dissolved in 1.80 ml of absolute ethanol. Quinolinone-chalcone 8d (200 mg, 0.57 mmol) was dissolved in 3.60 ml of acetic acid, and it was added dropwise to the ethanolic solution of hydrazine. The resulting solution was stirred at 120 °C for 24 h. The solid produced upon filtration was triturated from methanol/dichloromethane to afford the pure pyrazoline 9n as yellow powder. Yield: 180 mg (90%); m.p. > 250 °C; ¹H NMR (DMSO-d₆, 600 MHz): δ ppm 13.26 (s, 1H, OH), 8.99 (s, 1H, Ar-OH), 8.00 (d, 1H, J=7.8 Hz, Ar–H), 7.64 (t, 1H, J=7.2 Hz, Ar–H), 7.43 (d, 1H, J = 8.4 Hz, Ar-H), 7.28 (t, 1H, J = 7.2 Hz, Ar-H), 7.24 (d, 2H, J=8.4 Hz, Ar-H), 6.89 (br, 1H, Ar-H), 6.87 (d, 2H, H)J = 8.4 Hz, Ar-H), 6.71 (d, 1H, J = 7.8 Hz, Ar-H), 6.63 (d, 1H, J = 8.4 Hz, Ar-H), 5.18 (dd, 1H, J = 11.4, 7.2 Hz, $C_5 - H_y$), 4.15 (dd, 1H, J = 18.6, 12.0 Hz, $C_4 - \underline{H}_B$), 3.72 (s, 3H, $OC\underline{H}_3$), $3.49 (s, 3H, NCH_3), 3.47 (br, 1H, C_4-\overline{H_A}); ^{13}C NMR (DMSO-100)$ d_6 , 600 MHz): δ ppm 161.3 (C, C-3), 160.4 (C, C-4'), 151.9 (C, C-2'), 148.0 (C, C-3"), 146.0 (C, C-4"), 142.3 (C, C-1""), 139.1 (C, C-10'), 132.4 (CH, C-8'), 128.9 (CH, C-3'", C-5""), 123.7 (CH, C-6'), 122.9 (CH, C-7'), 122.1 (CH, C-6"), 118.0 (C, C-4""), 115.8 (CH, C-1"), 114.8 (C, C-9'), 114.7 (C, C-5"), 114.4 (CH, C-2", C-6"), 110.0 (C, C-5'), 101.0 (CH, C-2"), 99.1 (C, C-3'), 61.8 (CH, C-5), 55.6 (CH₃, Ar–O-<u>CH₃</u>), 47.0 (CH₂, C-4), 29.0 (CH₃, N-<u>CH₃</u>); HR-MS m/z (pos): 476.13721 C₂₆H₂₂ClN₃O₄ (calcd. 475.1299).

Experimental procedure for the synthesis of 4-hydroxy-3-(5-(4-hydroxy-3-methoxyphenyl)-1-phenyl-4,5-dihydro-1H-pyrazol-3-yl)quinolin-2(1H)-one (9o)

In 4.5 ml of acetic acid, 0.15 mmol (50 mg) of quinolinone-chalcone **8b** and 0.30 mmol (32.4 mg) of phenyl



Biological evaluation

Determination of the reducing activity of the stable radical 2,2-Diphenyl-1-picrylhydrazyl (DPPH)

To an ethanolic solution of DPPH (100 μ M) in absolute ethanol, an equal volume of the compounds dissolved in DMSO was added (100 μ M). The mixture was shaken vigorously and allowed to stand for 20 or 60 min; absorbance at 517 nm was determined spectrophotometrically, and the percentage of activity was calculated. All tests were undertaken on three replicates, and the results presented in Table 3 were averaged and compared with the appropriate standard nordihydroguaiaretic acid (NDGA) [66].

Inhibition of linoleic acid lipid peroxidation

Production of conjugated diene hydroperoxide by oxidation of linoleic acid in an aqueous dispersion is monitored at 234 nm. 2,2'-Azobis(2-amidinopropane) dihydrochloride (AAPH) is used as a free radical initiator. Ten microliters of the 16 mM linoleic acid sodium salt solution was added to the UV cuvette containing 0.93 ml of 0.05 M phosphate buffer, pH 7.4 prethermostated at 37 °C. The oxidation reaction was initiated at 37 °C under air by the addition of 50 μl of 40 mM AAPH solution. Oxidation was carried out in the presence of the synthesized compounds (10 μl, from a



stock solution of 10 mM in DMSO). In the assay without antioxidant, lipid oxidation was measured in the presence of the same level of DMSO. The rate of oxidation at 37 °C was monitored by recording the increase in absorption at 234 nm caused by conjugated diene hydroperoxides and compared with the appropriate standard trolox [66].

Soybean LOX inhibition study in vitro

The tested compounds dissolved in DMSO were incubated at room temperature with sodium linoleate (0.1 ml) and 0.2 ml of enzyme solution ($1/9 \times 10^{-4}$ w/v in saline) in Tris buffer pH 9. The conversion of sodium linoleate to 13-hydroper-oxylinoleic acid at 234 nm was recorded and compared with the appropriate standard inhibitor NDGA [66].

Molecular docking simulations for soybean LOX

Molecular docking studies of soybean LOX were performed selecting 3PZW from the Protein Data Bank (PDB) and using UCSF Chimera for the visualization of the protein (PDB code: 3PZW) [70]. Water molecules were removed, missing residues were added with Modeller [71], hydrogen atoms and AMBER99SB-ILDN charges were added while the iron charge was set to +2.0, with no restraint applied to the iron atom and the ligands.

OpenBabel was used to generate and minimize Ligand 3D coordinates [72] using the MMFF94 force field [73], while ACPYPE (AnteChamber PYthon Parser interfacE) [74] was used to generate ligand topologies and parameters using Antechamber [75]. Energy minimizations were achieved using the AMBER99SB-ILDN force field [76] with GROMACS 4.6.5 [77] as the molecular dynamics simulation toolkit. Docking studies were carried out with AutoDock Vina (1.1.2) [78] using a grid box of size 100 Å, 70 Å, 70 Å in X, Y, Z dimensions for soybean LOX. UCSF Chimera was used for the generation of docking input files and analysis of the obtained docking results. Docking was carried out with an exhaustiveness value of 10 and a maximum output of 20 docking modes soybean LOX.

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Compliance with ethical standards

Conflict of interest The authors have declared no conflict of interest.

References

 Brash AR (1999) Lipoxygenases: occurrence, functions, catalysis, and acquisition of substrate. J Biol Chem 274(34):23679–23682. https://doi.org/10.1074/jbc.274.34.23679

- Wisastra R, Dekker FJ (2014) Inflammation, cancer and oxidative lipoxygenase activity are intimately linked. Cancers 6(3):1500– 1521. https://doi.org/10.3390/cancers6031500
- Yanes DA, Mosser-Goldfarb JL (2018) Emerging therapies for atopic dermatitis: the prostaglandin/leukotriene pathway. J Am Acad Dermatol 78(3 Suppl 1):S71–S75. https://doi.org/10.1016/j. iaad.2017.12.021
- Radmark O, Werz O, Steinhilber D, Samuelsson B (2007) 5-Lipoxygenase: regulation of expression and enzyme activity. Trends Biochem Sci 32(7):332–341. https://doi.org/10.1016/j.tibs.2007.06.002
- Dahlén SE (2006) Treatment of asthma with antileukotrienes: first line or last resort therapy? Eur J Pharmacol 533(1–3):40–56. https://doi.org/10.1016/j.ejphar.2005.12.070
- Lötzer K, Funk CD, Habenicht AJ (2005) The 5-lipoxygenase pathway in arterial wall biology and atherosclerosis. Biochim Biophys Acta 1736(1):30–37. https://doi.org/10.1016/j.bbali p.2005.07.001
- Fürstenberger G, Krieg P, Müller-Decker K, Habenicht AJ (2006) What are cyclooxygenases and lipoxygenases doing in the driver's seat of carcinogenesis? Int J Cancer 119(10):2247–2254. https:// doi.org/10.1002/ijc.22153
- Pontiki E, Hadjipavlou-Litina D (2013) Targeting Lipoxygenases (LOs): drug design and discovery. Curr Enzym Inhib 9(2):89–105. https://doi.org/10.2174/1573408011309020003
- Lu P, Schrag ML, Slaughter DE, Raab CE, Shou M, Rodrigues AD (2003) Mechanism-based inhibition of human liver microsomal cytochrome P450 1A2 by zileuton, a 5-lipoxygenase inhibitor. Drug Metab Dispos 31(11):1352–1360. https://doi.org/10.1124/ dmd 31.11.1352
- Zaman K, Hanigan MH, Smith A, Vaughan J, Macdonald T, Jones DR, Hunt JF, Gaston B (2006) Endogenous S-nitrosoglutathione modifies 5-lipoxygenase expression in airway epithelial cells. Am J Respir Cell Mol Biol 34(4):387–393. https://doi.org/10.1165/ rcmb.2005-0336RC
- Gaztanaga J, Farkouh M, Rudd JH, Brotz TM, Rosenbaum D, Mani V, Kerwin TC, Taub R, Tardif JC, Tawakol A, Fayad ZA (2015) A phase 2 randomized, double-blind, placebo-controlled study of the effect of VIA-2291, a 5-lipoxygenase inhibitor, on vascular inflammation in patients after an acute coronary syndrome. Atherosclerosis 240(1):53–60. https://doi.org/10.1016/j. atherosclerosis.2015.02.027
- Matsumoto S, Ibrahim R, Grégoire JC, L'Allier PL, Pressacco J, Tardif JC, Budoff MJ (2017) Effect of treatment with 5-lipoxygenase inhibitor VIA-2291 (atreleuton) on coronary plaque progression: a serial CT angiography study. Clin Cardiol 40(4):210–215. https://doi.org/10.1002/clc.22646
- Bruno F, Spaziano G, Liparulo A, Roviezzo F, Nabavi SM, Sureda A, Filosa R, D'Agostino B (2018) Recent advances in the search for novel 5-lipoxygenase inhibitors for the treatment of asthma. Eur J Med Chem 153:65–72. https://doi.org/10.1016/j.ejmec h.2017.10.020
- Tavolari S, Bonafè M, Marini M, Ferreri C, Bartolini G, Brighenti E, Manara S, Tomasi V, Laufer S, Guarnieri T (2008) Licofelone, a dual COX/5-LOX inhibitor, induces apoptosis in HCA-7 colon cancer cells through the mitochondrial pathway independently from its ability to affect the arachidonic acid cascade. Carcinogenesis 29(2):371–380. https://doi.org/10.1093/carcin/bgm265
- Merchant N, Bhaskar LVKS, Momin S, Sujatha P, Reddy ABM, Nagaraju GP (2018) 5-Lipoxygenase: its involvement in gastrointestinal malignancies. Crit Rev Oncol Hematol 127:50–55. https://doi.org/10.1016/j.critrevonc.2018.05.012
- Kim W, Son B, Lee S, Do H, Youn B (2018) Targeting the enzymes involved in arachidonic acid metabolism to improve radiotherapy. Cancer Metastasis Rev 37(2–3):213–225. https:// doi.org/10.1007/s10555-018-9742-0



- Azad MB, Chen Y, Gibson SB (2009) Regulation of autophagy by reactive oxygen species (ROS): implications for cancer progression and treatment. Antioxid Redox Signal 11(4):777–790. https://doi.org/10.1089/ars.2008.2270
- Weber V, Rubat C, Duroux E, Lartigue C, Madesclaire M, Coudert P (2005) New 3- and 4-hydroxyfuranones as anti-oxidants and anti-inflammatory agents. Bioorg Med Chem 13(14):4552–4564. https://doi.org/10.1016/j.bmc.2005.04.055
- Pontiki E, Hadjipavlou-Litina D, Patsilinakos A, Tran TM, Marson CM (2015) Pteridine-2,4-diamine derivatives as radical scavengers and inhibitors of lipoxygenase that can possess anti-inflammatory properties. Future Med Chem 7(14):1937–1951. https://doi.org/10.4155/fmc.15.104
- Detsi A, Majdalani M, Kontogiorgis CA, Hadjipavlou-Litina D, Kefalas P (2009) Natural and synthetic 2'-hydroxy-chalcones and aurones: synthesis, characterization and evaluation of the antioxidant and soybean lipoxygenase inhibitory activity. Bioorg Med Chem 17(23):8073–8085. https://doi.org/10.1016/j. bmc.2009.10.002
- Bansal Y, Silakari O (2014) Multifunctional compounds: smart molecules for multifactorial diseases. Eur J Med Chem 76:31–42. https://doi.org/10.1016/j.ejmech.2014.01.060
- Fortin S, Bérubé G (2013) Advances in the development of hybrid anticancer drugs. Expert Opin Drug Discov 8(8):1029–1047. https://doi.org/10.1517/17460441.2013.798296
- Rana A, Alex JM, Chauhan M, Joshi G, Kumar R (2015) A review on pharmacophoric designs of antiproliferative agents. Med Chem Res 24(3):903–920. https://doi.org/10.1007/s00044-014-1196-5
- Nakashima K, Oyama M, Ito T, Akao Y, Witono JR, Darnaedi D, Tanaka T, Murata J, Iinuma M (2012) Novel quinolinone alkaloids bearing a lignoid moiety and related constituents in the leaves of *Melicope denhamii*. Tetrahedron 68(10):2421–2428. https://doi. org/10.1016/j.tet.2012.01.007
- Gao H, Zhang L, Zhu T, Gu Q, Li D (2012) Unusual pyrrolyl 4-quinolinone alkaloids from the marine-derived fungus *Penicillium* sp. ghq208. Chem Pharm Bull 60(11):1458–1460. https://doi.org/10.1248/cpb.c12-00487
- Detsi A, Bouloumbasi D, Prousis KC, Koufaki M, Athanasellis G, Melagraki G, Afantitis A, Igglessi-Markopoulou O, Kontogiorgis C, Hadjipavlou-Litina DJ (2007) Design and synthesis of novel quinolinone-3-aminoamides and their alpha-lipoic acid adducts as antioxidant and antiiflammatory agents. J Med Chem 50(10):2450–2458. https://doi.org/10.1021/jm061173n
- Angeleska S, Kefalas P, Detsi A (2013) Crude peroxidase from onion solid waste as a tool for organic synthesis. Part III: synthesis of tetracyclic heterocycles (coumestans and benzofuroquinolinones). Tetrahedron Lett 54(19):2325–2328. https://doi. org/10.1016/j.tetlet.2013.02.081
- Jönsson S, Andersson G, Fex T, Fristedt T, Hedlund G, Jansson K, Abramo L, Fritzson I, Pekarski O, Runström A, Sandin H, Thuvesson I, Björk A (2004) Synthesis and biological evaluation of new 1,2-dihydro-4-hydroxy-2-oxo-3-quinolinecarboxamides for treatment of autoimmune disorders: structure-activity relationship. J Med Chem 47(8):2075–2088. https://doi.org/10.1021/jm031044w
- Rahman MA (2011) Chalcone: a valuable insight into the recent advances and potential pharmacological activities. J Chem Sci 29:1–16
- Chetana BP, Mahajan SK, Suvarna AK (2009) Chalcone: a versatile molecule. J Pharm Sci Res 1(3):11–22. https://doi. org/10.1002/chin.201025251
- Bandgar BP, Gawande SS, Bodade RG, Totre JV, Khobragade CN (2010) Synthesis and biological evaluation of simple methoxylated chalcones as anticancer, anti-inflammatory and antioxidant agents. Bioorg Med Chem 18(3):1364–1370. https://doi. org/10.1016/j.bmc.2009.11.066

- 32. Bandgar BP, Patil SA, Korbad BL, Nile SH, Khobragade CN (2010) Synthesis and biological evaluation of beta-chloro vinyl chalcones as inhibitors of TNF-alpha and IL-6 with antimicrobial activity. Eur J Med Chem 45(6):2629–2633. https://doi.org/10.1016/j.eimech.2010.01.050
- Roussaki M, Hall B, Lima SC, Cordeiro da Silva A, Wilkinson S, Detsi A (2013) Synthesis and anti-parasitic activity of a novel quinolinone–chalcone series. Bioorg Med Chem Lett 23(23):6436– 6441. https://doi.org/10.1016/j.bmcl.2013.09.047
- Siddiqui ZN, Musthafa TNM, Ahmad A, Khan AU (2011) Thermal solvent-free synthesis of novel pyrazolyl chalcones and pyrazolines as potential antimicrobial agents. Bioorg Med Chem Lett 21(10):2860–2865
- Dave SS, Rahatgaonkar AM (2011) Syntheses and anti-microbial evaluation of new quinoline scaffold derived pyrimidine derivatives. Arab J Chem Res 9:S451–S456. https://doi.org/10.1016/j. arabjc.2011.06.009
- Sonawane V, Hiwarale D, Sonawane B, Kamble S, Sonawane K, Bhosale R (2018) Synthesis, characterization and biological evaluation of novel imidazo thiazole chalcones as antioxidant agent. Int J Pharm Sci Res 10(5):2439–2446. https://doi.org/10.13040/ LIPSR.0975-8232
- Ramalho SD, Bernades A, Demetriusa G, Perez CN, Vieirab PC, Santos CY, Silva JA, Moraes MO, Mousinhod KC (2013) Synthetic chalcone derivatives as inhibitors of cathepsins K and B, and their cytotoxic evaluation. Chem Biodivers 10:1999–2006. https://doi.org/10.1002/cbdv.201200344
- Champelovier P, Chauchet X, Puch FH, Vergnaud S, Garrel C, Laporte F, Boutonnat J, Boumendjel A (2013) Cellular and molecular mechanisms activating the cell death processes by chalcones: critical structural effects. Toxicol In Vitro 27(8):2305–2315. https://doi.org/10.1016/j.tiv.2013.09.021
- Yusuf M, Jain P (2014) Synthetic and biological studies of pyrazolines and related heterocyclic compounds. Arab J Chem 7(5):553–596. https://doi.org/10.1016/j.arabjc.2011.09.013
- Singh P, Singh J, Pant GJ, Rawat MSM (2018) 2-Pyrazolines as biologically active and fluorescent agents, an overview. Anticancer Agents Med Chem 18(10):1366–1385. https://doi. org/10.2174/1871520618666180313153407
- Almahdi MM, Saeed AEM, Metwally NH (2019) Synthesis and antimicrobial activity of some new pyrazoline derivatives bearing sulfanilamido moiety. Eur J Chem 10(1):30–36. https://doi. org/10.5155/eurjchem.10.1.30-36.1791
- Abdel-Wahab BF, Abdel-Latif E, Mohamed HA, Awad GE (2012) Design and synthesis of new 4-pyrazolin-3-yl-1,2,3-triazoles and 1,2,3-triazol-4-yl-pyrazolin-1-ylthiazoles as potential antimicrobial agents. Eur J Med Chem 52:263–268. https://doi.org/10.1016/j.ejmech.2012.03.023
- Azarifar D, Shaebanzadeh M (2002) Synthesis and characterization of new 3,5-dinaphthyl substituted 2-pyrazolines and study of their antimicrobial activity. Molecules 7(12):885–895. https://doi.org/10.3390/71200885
- 44. Rathore P, Yaseen S, Ovais S, Bashir R, Yaseen R, Hameed AD, Samim M, Gupta R, Hussain F, Javed K (2014) Synthesis and evaluation of some new pyrazoline substituted benzenesulfonylureas as potential antiproliferative agents. Bioorg Med Chem Lett 24(7):1685–1691. https://doi.org/10.1016/j.bmcl.2014.02.059
- 45. Wang H, Zheng J, Xu W, Chen C, Wei D, Ni W, Pan Y (2017) A new series of cytotoxic pyrazoline derivatives as potential anticancer agents that induce cell cycle arrest and apoptosis. Molecules 22(10):1635. https://doi.org/10.3390/molecules22101635
- Khalil NA, Ahmed EM, El-Nassan HB, Ahmed OK, Al-Abd AM (2012) Synthesis and biological evaluation of novel pyrazoline derivatives as anti-inflammatory and antioxidant agents. Arch Pharm Res 35(6):995–1002. https://doi.org/10.1007/s1227 2-012-0606-9



- 47. Shringare SN, Chavan HV, Bhale PS, Dongare SB, Mule YB, Patil SB, Bandgar BP (2018) Synthesis and pharmacological evaluation of combretastatin-A4 analogs of pyrazoline and pyridine derivatives as anticancer, anti-inflammatory and antioxidant agents. Med Chem Res 27(4):1226–1237. https://doi.org/10.1007/s00044-018-2142-8
- Jeong TS, Kim KS, Kim JR, Cho KH, Lee S, Lee WS (2004) Novel 3,5-diaryl pyrazolines and pyrazole as low-density lipoprotein (LDL) oxidation inhibitor. Bioorg Med Chem Lett 14(11):2719–2723. https://doi.org/10.1016/j.bmcl.2004.03.072
- Kaplancikli ZA, Ozdemir A, Turan-Zitouni G, Altintop MD, Can OD (2010) New pyrazoline derivatives and their antidepressant activity. Eur J Med Chem 45(9):4383–4387. https://doi. org/10.1016/j.ejmech.2010.06.011
- Bandgar BP, Adsul LK, Chavan HV, Jalde SS, Shringare SN, Shaikh R, Meshram RJ, Gacche RN, Masand V (2012) Synthesis, biological evaluation, and docking studies of 3-(substituted)-aryl-5-(9-methyl-3-carbazole)-1H-2-pyrazolines as potent anti-inflammatory and antioxidant agents. Bioorg Med Chem Lett 22(18):5839–5844. https://doi.org/10.1016/j.bmcl.2012.07.080
- Kharbanda C, Alam MS, Hamid H, Javed K, Bano S, Dhulap A, Ali Y, Nazreen S, Haider S (2014) Synthesis and evaluation of pyrazolines bearing benzothiazole as anti-inflammatory agents. Bioorg Med Chem 22(21):5804–5812. https://doi.org/10.1016/j. bmc.2014.09.028
- Viveka S, Dinesha Shama P, Nagaraja GK, Ballav S, Kerkar S (2015) Design and synthesis of some new pyrazolyl-pyrazolines as potential anti-inflammatory, analgesic and antibacterial agents. Eur J Med Chem 101(28):442–451. https://doi.org/10.1016/j.ejmech.2015.07.002
- He J, Ma L, Wei Z, Zhu J, Peng F, Shao M, Lei L, He L, Tang M, He L, Wu Y, Chen L (2015) Synthesis and biological evaluation of novel pyrazoline derivatives as potent anti-inflammatory agents. Bioorg Med Chem Lett 25(11):2429–2433. https://doi.org/10.1016/j.bmcl.2015.03.087
- Eid NM, George RF (2018) Facile synthesis of some pyrazoline-based compounds with promising anti-inflammatory activity. Future Med Chem 10(2):183–199. https://doi.org/10.4155/ fmc-2017-0144
- Turkan F, Cetin A, Taslimi P, Karaman HS, Gulçin İ (2019) Synthesis, characterization, molecular docking and biological activities of novel pyrazoline derivatives. Arch Pharm (Weinheim) 352(6):e1800359. https://doi.org/10.1002/ardp.201800359
- Ozmen Ozgun D, Gul HI, Yamali C, Sakagami H, Gulcin I, Sukuroglu M, Supuran CT (2019) Synthesis and bioactivities of pyrazoline benzensulfonamides as carbonic anhydrase and acetylcholinesterase inhibitors with low cytotoxicity. Bioorg Chem 84:511–517. https://doi.org/10.1016/j.bioorg.2018.12.028
- 57. Stefanes NM, Toigo J, Maioral MF, Jacques AV, Chiaradia-Delatorre LD, Perondi DM, Ribeiro AAB, Bigolin Á, Pirath IMS, Duarte BF, Nunes RJ, Santos-Silva MC (2019) Synthesis of novel pyrazoline derivatives and the evaluation of death mechanisms involved in their antileukemic activity. Bioorg Med Chem 27(2):375–382. https://doi.org/10.1016/j.bmc.2018.12.012
- Mumtaz A, Majeed A, Zaib S, Ur Rahman S, Hameed S, Saeed A, Rafique H, Mughal E, Maalik A, Hussain I, Iqbal J (2019) Investigation of potent inhibitors of cholinesterase based on thiourea and pyrazoline derivatives: synthesis, inhibition assay and molecular modeling studies. Bioorg Chem 90:103036. https://doi.org/10.1016/j.bioorg.2019.103036
- Cai X, Zhao S, Cai Zheng J, Zhu Z, Wei D, Zheng Z, Zhu H, Chen Y (2019) Synthesis and evaluation of novel D-ring substituted steroidal pyrazolines as potential anti-inflammatory agents. Steroids 146:70–78. https://doi.org/10.1016/j.steroids.2019.03.012
- Sethiya JP, Bhavsar SP, Shahare HV (2019) Synthesis of potential anti-inflammatory pyrazoline derivatives under ultrasonic

- irradiation. Int J Pharm Sci Res 10:3290–3294. https://doi. org/10.13040/IJPSR.0975-8232.10(7).3290-94
- Chandel P, Kumar A, Singla N, Kumar A, Singh G, Gill RK (2019) Rationally synthesized coumarin based pyrazolines ameliorate carrageenan induced inflammation through COX-2/proinflammatory cytokine inhibition. Medchemcomm 10(3):421–430. https://doi.org/10.1039/c8md00457a
- 62. Sharma S, Kaur S, Bansal T, Gaba J (2014) Review on synthesis of bioactive pyrazoline derivatives. Chem Sci Trans 3(3):861–875. https://doi.org/10.7598/cst2014.796
- Weber FG, Brosehe K, Seedorf Ch, Itinow A (1969) 1,3,5-Triaryl-Δ²-pyrazoline. Monatsh Chem 100(6):1924–1927. https://doi.org/10.1007/BF01151742
- 64. Bhatnagar I, George MV (1968) Oxidation with metal oxides—II: oxidation of chalcone phenylhydrazones, pyrazolines, o-aminobenzylidine anils and o-hydroxy benzylidine anils with manganese dioxide. Tetrahedron 24(3):1293–1298. https://doi.org/10.1016/0040-4020(68)88080-9
- Detsi A, Bardakos V, Markopoulos J, Igglessi-Markopoulou O (1996) Reactions of 2-methyl-3,1-benzoxazin-4-one with active methylene compounds: a new route to 3-substituted 4-hydrox-yquinolin-2(1H)-ones. J Chem Soc Perkin Trans 1(24):2909–2913. https://doi.org/10.1039/P19960002909
- Hadjipavlou D, Garnelis T, Athanassopoulos CM, Papaioannou D (2009) Kukoamine a analogs with lipoxygenase inhibitory activity. J Enzyme Inhib Med Chem 24(5):1188–1193. https://doi.org/10.1080/14756360902779193
- 67. Kontogiorgis C, Hadjipavlou-Litina D (2003) Biological evaluation of several coumarin derivatives designed as possible anti-inflammatory/antioxidant agents. J Enzyme Inhib Med Chem 18(1):63–69. https://doi.org/10.1080/1475636031000069291
- Roussaki M, Kontogiorgis CA, Hadjipavlou-Litina D, Hamilakis S, Detsi A (2010) A novel synthesis of 3-aryl coumarins and evaluation of their antioxidant and lipoxygenase inhibitory activity. Bioorg Med Chem Lett 20(13):3889–3892. https://doi.org/10.1016/j.bmcl.2010.05.022
- Denisov ET, Afanas'ev IB (2005) Oxidation and antioxidants in organic chemistry and biology. CRC Press, Boca Raton
- Pettersen EF, Goddard TD, Huang CC, Couch GS, Greenblatt DM, Meng EC, Ferrin TE (2004) UCSF Chimera-a visualization system for exploratory research and analysis. J Comput Chem 25(13):1605-1612. https://doi.org/10.1002/jcc.20084
- Fiser A, Sali A (2003) Modeller: generation and refinement of homology-based protein structure models. Methods Enzymol 374:461–491. https://doi.org/10.1016/S0076-6879(03)74020-8
- O'Boyle NM, Banck M, James CA, Morley C, Vandermeersch T, Hutchison GR (2011) Open Babel: an open chemical toolbox. J Cheminform 3:33. https://doi.org/10.1186/1758-2946-3-33
- 73. Halgren TA (1996) Merck molecular force field. I. Basis, form, scope, parameterization, and performance of MMFF94. J Comput Chem 17(5–6):490–515. https://doi.org/10.1002/(sici)1096-987x(199604)17:5/6%3c490::aid-jcc1%3e3.0.co;2-p
- Sousa de Silva AW, Vranken WF (2012) ACPYPE—AnteChamber PYthon Parser interface. BMC Res Notes 5:367. https://doi.org/10.1186/1756-0500-5-367
- Wang J, Wang W, Kollman PA, Case DA (2006) Automatic atom type and bond type perception in molecular mechanical calculations. J Mol Graph Model 25(2):247–260. https://doi. org/10.1016/j.jmgm.2005.12.005
- Lindorff-Larsen K, Piana S, Palmo K, Maragakis P, Klepeis JL, Dror RO, Shaw DE (2010) Improved side-chain torsion potentials for the Amber ff99SB protein force field. Proteins 78(8):1950–1958. https://doi.org/10.1002/prot.22711
- Hess B, Kutzner C, van der Spoel D, Lindhal E (2008)
 GROMACS 4: algorithms for highly efficient, load-balanced,



- and scalable molecular simulation. J Chem Theory Comput 4(3):435-447. https://doi.org/10.1021/ct700301q
- Trott O, Olson AJ (2010) AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading. J Comput Chem 31(2):455–461. https://doi.org/10.1002/jcc.21334

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